RELATIVE BIOAVAILABILITY OF LEAD IN SOILS FROM THE VBI70 SITE

March 2001

Stan W. Casteel, DVM, PhD, DABVT
Principal Investigator
Veterinary Medical Diagnostic Laboratory
College of Veterinary Medicine
University of Missouri, Columbia
Columbia, Missouri

Christopher P. Weis, PhD, DABT
Study Design and Technical Advisor
Bonita Lavelle
Remedial Project Manager
US Environmental Protection Agency – Region 8
Denver, Colorado

William J. Brattin, PhD Tracy L. Hammon, MS Technical Consultants Syracuse Research Corporation Denver, Colorado







ACKNOWLEDGEMENTS

The work described in this report is the product of a team effort involving a number of people. In particular, the authors would like to acknowledge the efforts and support of the following:

Tim J. Evans, DVM, University of Missouri-Columbia, provided support in all aspects of the implementation of this study.

Stevey R. Morris, BS, University of Missouri-Columbia, coordinated sample preparation, doughball preparation, and sample tracking activities.

John Drexler, PhD, University of Colorado, provided speciation for test materials utilized in this investigation.

Edward J. Hinderberger, Jr., PhD, L.E.T., Inc, provided lead analysis for samples collected during this investigation.

Washington Group, assisted in the connection and preparation of the test materials for this investigation.

EXECUTIVE SUMMARY

A study using young swine as test animals was performed to measure the gastrointestinal absorption of lead in two soils from the Vasquez Boulevard/I-70 site (VB-I70) located in Denver, Colorado. Young swine were selected for use in the study primarily because the gastrointestinal physiology and overall size of young swine are similar to that of young children, who are the population of prime concern for exposure to soil lead.

The test materials were prepared by combining soil samples collected from residential properties within the study area. The soil samples were selected to represent both an Eastern and Western area. The lead concentration in these samples was 723 parts per million (ppm) for test material #1 (Eastern Sample) and 987 ppm for test material #2 (Western Sample). Groups of 5 swine were given average oral doses of 103.7, 311.2 or 691.6 mg/kg-d of test material #1 or 76.0, 228.0, or 506.6 mg/kg-d of test material #2 for 15 days. This corresponded to target average doses of 75, 225, or 500 ug/kg/day of lead. Other groups of animals were given a standard lead reference material (lead acetate) orally at doses of 0, 25, 75, or 225 ug Pb/kg-day. The amount of lead absorbed by each animal was evaluated by measuring the amount of lead in the blood (measured on days 0, 1, 2, 3, 5, 7, 9, 12, and 15), and the amount of lead in liver, kidney and bone (measured on day 15 at study termination). The amount of lead present in blood or tissues of animals exposed to test soils was compared to that for animals exposed to lead acetate, and the results were expressed as relative bioavailability (RBA). For example, a relative bioavailability of 50% means that 50% of the lead in soil was absorbed equally as well as lead from lead acetate, and 50% behaved as if it were not available for absorption. Thus, if lead acetate were 40% absorbed, the test material would be 20% absorbed.

The RBA results for the two samples from the VB-I70 site are summarized below:

Measurement Endpoint	Test Material #1 Eastern Sample	Test Material #2 Western Sample
Blood Lead Area Under Curve	87%	85%
Liver Lead	98%	70%
Kidney Lead	97%	78%
Bone Lead	69%	56%

Because the estimates of RBA based on blood, liver, kidney, and bone do not agree in all cases, judgment must be used in interpreting the data. In general, EPA recommends greatest emphasis be placed on the RBA estimates derived from the blood lead data. This is because blood lead data are more robust and less susceptible to random errors than the tissue lead data, so there is greater confidence in RBA estimates based on blood lead. In addition, absorption into the central compartment is an early indicator of lead exposure, is

the most relevant index of central nervous system exposure, and is the standard measurement endpoint in investigations of this sort. However, data from the tissue endpoints (liver, kidney, bone) also provide valuable information. EPA considers the <u>plausible range</u> to extend from the RBA based on blood AUC to the mean of the other three tissues (liver, kidney, bone). The <u>preferred range</u> is the interval from the RBA based on blood to the mean of the blood RBA and the tissue mean RBA. Our <u>suggested point</u> <u>estimate</u> is the mid-point of the preferred range. These values are presented below:

Relative Bioavailability of Lead	Test Material #1 Eastern Sample	Test Material #2 Western Sample
Plausible Range	87-88%	68-85%
Preferred Range	87-88%	76-85%
Suggested Point Estimate	87%	81%

These RBA estimates may be used to help assess lead risk at this site by refining the estimate of absolute bioavailability (ABA) of lead in soil, as follows:

$$ABA_{soil} = ABA_{soluble} * RBA_{soil}$$

Available data indicate that fully soluble forms of lead are about 50% absorbed by a child. Thus, the estimated absolute bioavailability of lead in the site samples is as follows:

Absolute Bioavailability of Lead	Test Material #1 Eastern Sample	Test Material #2 Western Sample
Plausible Range	43-44%	34-42%
Preferred Range	43-44%	38-42%
Suggested Point Estimate	43%	40%

These absolute bioavailability estimates are appropriate for site-specific use in EPA's IEUBK model, although it is clear that there is both natural variability and uncertainty associated with these estimates. This variability and uncertainty arises from several sources, including: 1) the inherent variability in the responses of different individual animals to lead exposure, 2) uncertainty in the relative accuracy and applicability of the different measurement endpoints, 3) the extrapolation of measured RBA values in swine to young children, and 4) the potential effect of food in the stomach on lead absorption. Thus, the values reported above are judged to be reasonable estimates of typical lead absorption by children at this site, but should be interpreted with the understanding that the values are not certain.

TABLE OF CONTENTS

1.0	INTROL	DUCTION	1
2.0	STUE 2.1 2.2 2.3	PY DESIGN	<u>3</u>
	2.4	Dosing	1
	2.5	Collection of Biological Samples	
	2.6	Preparation of Biological Samples for Analysis	<u>3</u>
	2.7	Lead Analysis	3
3.0	3.1 3.2 3.3 3.4	ANALYSIS Overview 1 Fitting the Curves Responses Below Quantitation Limit Quality Assurance LTS Blood Lead vs Time Dose-Response Patterns Calculated RBA Values Estimated Absolute Bioavailability in Children 1 2 2 2 2 2 2 3 4 5 5 6 7 7 7 7 7 7 7 7 7 7 7 7	5 5 6 6 1 1 1 7 8
	4.5	Uncertainty	9
5.0	REFE	RENCES <u>3</u>	0
APP	ENDIX	TITLE	
APP	ENDIX	A DETAILED DATA AND CALCULATIONS	
APP	ENDIX I	B TEST MATERIAL SELECTION AND PREPARATION	

LIST OF TABLES

TABLE	IIILE
2-1 2-2 2-3	LEAD AND ARSENIC ANALYSIS OF TEST MATERIALS
	LIST OF FIGURES
FIGURE	TITLE
2-1 2-2 2-3	FREQUENCY OF LEAD PARTICLES
3-1 3-2	QA DUPS 18 CHECK STANDARDS 20
4-1	GROUP MEAN BLOOD VALUES
4-2 4-3	AUC
4-4 4-5	LIVER

1.0 INTRODUCTION

Absolute and Relative Bioavailability

Bioavailability is a concept that relates to the absorption of chemicals and how absorption depends upon the physical-chemical properties of the chemical and its medium (e.g., dust, soil, rock, food, water, etc.) and the physiology of the exposed receptor. Bioavailability is normally described as the fraction (or percentage) of a chemical which enters into the blood following an exposure of some specified amount, duration and route (usually oral). In some cases, bioavailability may be measured using chemical levels in peripheral tissues such as liver, kidney, and bone, rather than blood. The fraction or percentage absorbed may be expressed either in absolute terms (absolute bioavailability, ABA) or in relative terms (relative bioavailability, RBA). Absolute bioavailability is measured by comparing the amount of chemical entering the blood (or other tissue) following oral exposure to test material with the amount entering the blood (or other tissue) following intravenous exposure to an equal amount of some dissolved form of the chemical. Similarly, relative bioavailability is measured by comparing oral absorption of test material to oral absorption of some fully soluble form of the chemical (e.g., either the chemical dissolved in water, or a solid form that is expected to fully dissolve in the stomach). For example, if 100 ug of dissolved lead were administered in drinking water and a total of 50 ug entered the blood, the ABA would be 0.50 (50%). Likewise, if 100 ug of lead in soil were administered and 30 ug entered the blood, the ABA for soil would be 0.30 (30%). If the lead dissolved in water were used as the reference substance for describing the relative amount of lead absorbed from soil, the RBA would be 0.30/0.50 = 0.60 (60%). These values (50% absolute bioavailability of dissolved lead and 30% absolute absorption of lead in soil) are the values currently employed as defaults in EPA's IEUBK model.

It is important to recognize that simple <u>solubility</u> of a test material in water or some other fluid (e.g., a weak acid intended to mimic the gastric contents of a child) may not be a reliable estimator of <u>bioavailability</u> due to the non-equilibrium nature of the dissolution and transport processes that occur in the gastrointestinal tract (Mushak 1991). For example, fluid volume and pH are likely to be changing as a function of time, and transport of lead across the gut will prevent an approach to equilibrium concentrations, especially for poorly soluble lead compounds. However, information on the solubility of lead in different materials is useful in interpreting the importance of solubility as a determinant of bioavailability. To avoid confusion, the term "bioaccessability" is used to refer to the relative amount of lead that dissolves under a specified set of test conditions.

For additional discussion about the concept and application of bioavailability see Goodman et al. (1990), Klaassen et al. (1996), and/or Gibaldi and Perrier (1982).

Using Bioavailability Data to Improve Exposure Calculations for Lead

When data are available on the bioavailability of lead in soil, dust, or other soil-like waste material at a site, this information can often be used to improve the accuracy of exposure and risk calculations at that site. The basic equation for estimating the site-specific ABA of a test soil is as follows:

$$ABA_{soil} = ABA_{soluble} \cdot RBA_{soil}$$

where:

ABA_{soil} = Absolute bioavailability of lead in soil ingested by a child

 $ABA_{soluble}$ = Absolute bioavailability in children of some dissolved or fully

soluble form of lead

 RBA_{soil} = RBA for soil measured in swine

Based on available information on lead absorption in humans and animals, the EPA estimates that the absolute bioavailability of lead from water and other fully soluble forms of lead is usually about 50% in children. Thus, when a reliable site-specific RBA value for soil is available, it may be used to estimate a site-specific absolute bioavailability as follows:

$$ABA_{soil} = 50\% \cdot RBA_{soil}$$

In the absence of site-specific data, the absolute absorption of lead from soil, dust and other similar media is estimated by EPA to be about 30%. Thus, the default RBA used by EPA for lead in soil and dust compared to lead in water is 30%/50% = 60%. When the measured RBA in soil or dust at a site is found to be less than 60% compared to some fully soluble form of lead, it may be concluded that exposures to and risks from lead in these media at that site are probably lower than typical default assumptions. If the measured RBA is higher than 60%, absorption of and risk from lead in these media may be higher than usually assumed.

2.0 STUDY DESIGN

A standardized study protocol for measuring absolute and relative bioavailability of lead was developed based upon previous study designs and investigations that characterized the young pig model (Weis et al. 1995). The study was performed as nearly as possible within the spirit and guidelines of Good Laboratory Practices (GLP: 40 CFR 792). Standard Operating Procedures (SOPs) that included detailed methods for all aspects of the study were prepared, approved, and distributed to all study members prior to the study (USEPA 2000).

2.1 Test Materials

Soil samples were collected from various residential properties within the VBI70 site which were selected for specific concentrations of lead and arsenic (Washington Group, 2000; attached as appendix B). The two soils were prepared to represent both Eastern (Test Material #1) and Western (Test Material #2) neighborhoods within this site. Six individual soil samples were combined to make the Eastern sample, and five individual soil samples were combined to make the Western sample. Each sample was dried in a laboratory over at 105 C, bulk sieved with a 2-mm screen and fine sieved with a 250-mm screen. Further details regarding the selection and preparation of test materials can be found in a separate technical memorandum (Washington Group, 2000).

Table 2-1 summarizes the lead and arsenic content of the test soils measured using ICP method SW6010. As seen, average lead concentrations in the Eastern and Western Test Materials are 723 and 987 mg/kg, respectively.

TABLE 2-1 LEAD AND ARSENIC ANALYSIS OF TEST MATERIALS

Sample	Replicate	Arsenic (mg/kg)	Lead (mg/kg)
Eastern Sample (TM#1)	#1	19	700
	#2	19	710
	#3	20	760
	Average	19	723
Western Sample (TM#2)	#1	26	970
·	#2	25	1000
	#3	24	990
	Average	25	987

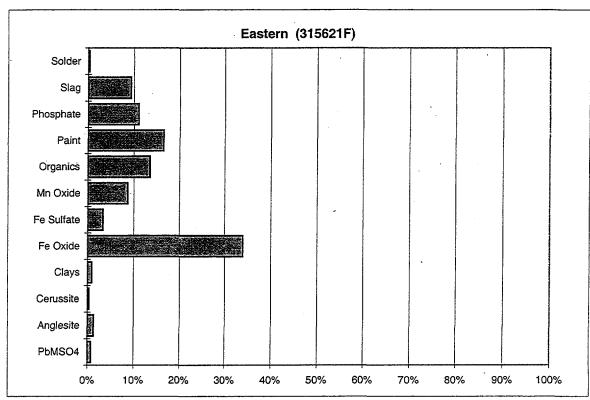
Each sample of test material was well mixed and analyzed by electron microprobe in order to identify a) how frequently particles of various lead minerals were observed, b) how frequently different types of mineral particles occur entirely inside particles of rock or slag ("included") and how often they occur partially or entirely outside rock or slag particles ("liberated"), c) the size distribution of particles of each mineral class, and d) approximately how much of the total amount of lead in the sample occurs in each mineral type. This is referred to as "relative lead mass". The results are summarized in Figures 2-1 to 2-3.

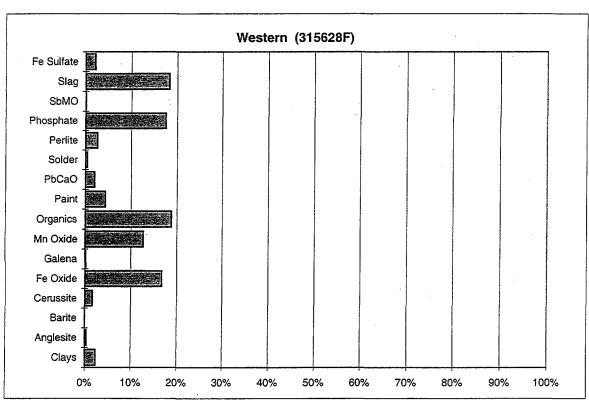
As seen in Figure 2-1 (top panel), the most common lead-bearing particle type (i.e., those which are observed most often) for the Eastern Sample (Test Material #1) was Iron Oxide, accounting for about 34% of all lead-bearing particles. However, as shown in Figure 2-2 (upper panel)), because the concentration of lead in iron oxide is relatively low, this phase accounted for only about 7.3% of the lead mass in this sample. The remainder of the lead in the eastern sample occurred mainly in particles of phosphate (41.4%), anglesite (15.6%) and paint (12%). Also shown in Figure 2-1 (bottom panel) are the results for the Western Sample (Test Material #2). As seen, the most common lead-bearing particle types were slag, phosphate, organics and iron oxide, accounting for about 18.3%, 17.5%, 18.8% and 16.7% of all lead-bearing particles, respectively. The majority of lead mass (Figure 2-2, bottom panel) in this sample was found in the phosphate (52.7%) and cerussite (18.3%) phases.

Figure 2-3 shows the distribution of the size of lead-bearing particles in the sample. As seen, there was a fairly broad distribution of lead-bearing particle sizes in both test materials, mainly ranging from 50-200 um. As noted above, small particles are often assumed to be more likely to adhere to the hand and be ingested and/or transported into the house. Further, small particles have larger surface area-to-volume ratios than larger particles, and so may tend to dissolve more rapidly in the acidic contents of the stomach than larger particles. Thus, small particle (e.g., less than 50-100 um) are thought to be of greater potential concern to humans than larger particles (e.g., 100 -250 um or larger).

Another property of lead particles that may be important in determining bioaccessability and/or bioavailability is the degree to which they are partially or entirely free from surrounding matrix ("liberated"). Based on the measured frequency of each type of particle existing in a liberated state, it can be calculated that of the total relative lead present in the samples, about 97.2% exists in liberated particles in the Eastern Sample (TM1) and 95.5% exists in liberated particles in the Western Sample (TM2). These high percentages of partially or entirel liberated grains may tend to increase the bioavailability of lead in the samples.

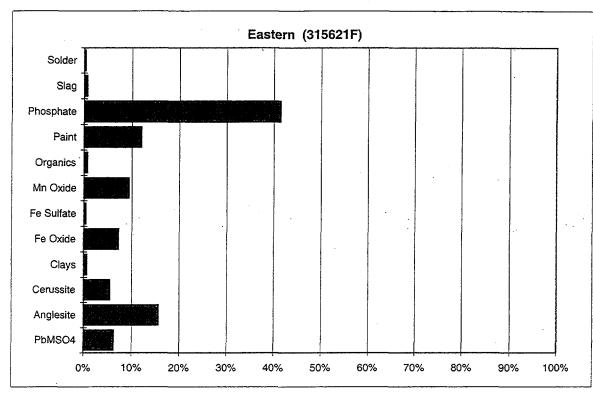
FIGURE 2-1 FREQUENCY OF LEAD PARTICLES





Report Figures.xls: Particle Freq 2/23/2001

FIGURE 2-2 LEAD MASS



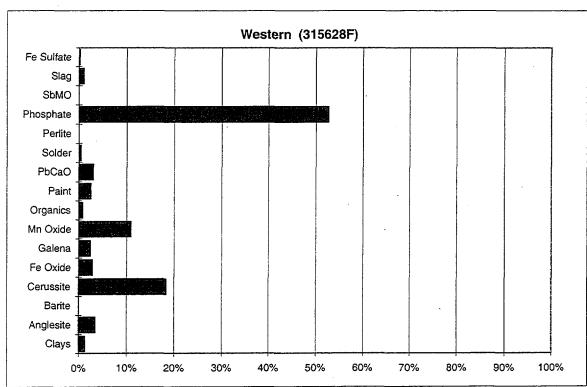
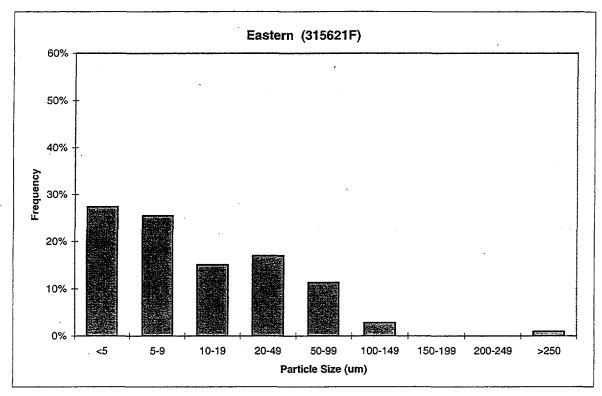
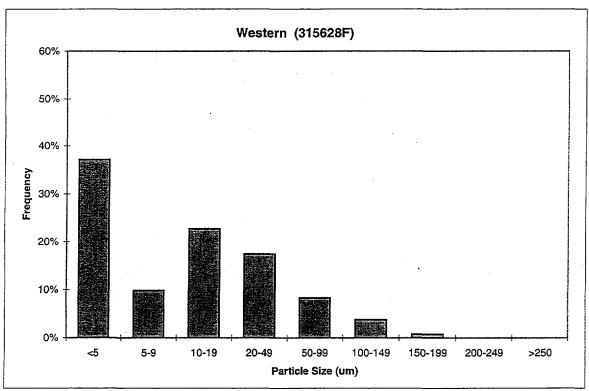


FIGURE 2-3 PARTICLE SIZE DISTRIBUTION





2.2 Experimental Animals

Young swine were selected for use in these studies because they are considered to be a good physiological model for gastrointestinal absorption in children (Weis and LaVelle 1991). The animals were intact males of the Pig Improvement Corporation (PIC) genetically defined Line 26, and were purchased from Chinn Farms, Clarence, MO. The animals were held under quarantine to observe their health for one week before beginning exposure to test materials. To minimize weight variations between animals and groups, the number of animals purchased from the supplier was six more than needed for the study, and the six animals most different in body weight on day -4 (either heavier or lighter) were excluded from further study. The remaining animals were assigned to dose groups at random. When exposure began (day zero), the animals were about 5-6 weeks old (juveniles, weaned at 3 weeks) and weighed an average of about 9.7 kg. Animals were weighed every three days during the course of the study. The group mean body weights over the course of the study are shown in Figure 2-4. On average, animals gained about 0.5 kg/day, and the rate of weight gain was comparable in all groups.

All animals were housed in individual lead-free stainless steel cages. Each animal was examined by a certified veterinary clinician (swine specialist) prior to being placed on study, and all animals were examined daily by an attending veterinarian while on study. Blood samples were collected for hematological analysis on days -4, 7, and 15 to assist in clinical health assessments. In this study, there was one animal that was removed from the study due to concerns over poor health.

2.3 Diet

Animals provided by the supplier were weaned onto standard pig chow purchased from MFA Inc., Columbia, MO. In order to minimize lead exposure from the diet, the animals were gradually transitioned from the MFA feed to a special low-lead feed (guaranteed less than 0.2 ppm lead, purchased from Zeigler Brothers, Inc., Gardners, PA) over the time interval from day -7 to day -3, and this feed was then maintained for the duration of the study. The feed was nutritionally complete and met all requirements of the National Institutes of Health-National Research Council. The typical nutritional components and chemical analysis of the feed is presented in Table 2-2. Typically, the feed contained approximately 5.7% moisture, 1.7% fiber, and provided about 3.4 kcal of metabolizable energy per gram. Analysis of two feed samples during this experiment indicated the mean lead level was 0.15 ppm.

Each day every animal was given an amount of feed equal to 5% of the mean body weight of all animals on study. Feed was administered in two equal portions of 2.5% of the mean body weight at each feeding. Feed was provided at 11:00 AM and 5:00 PM daily. Drinking water was provided ad libitum via self-activated watering nozzles within each cage. Analysis of samples from randomly selected drinking water nozzles indicated the mean lead concentration (treating non-detects at one-half the quantitation limit) was less than 1 ug/L.

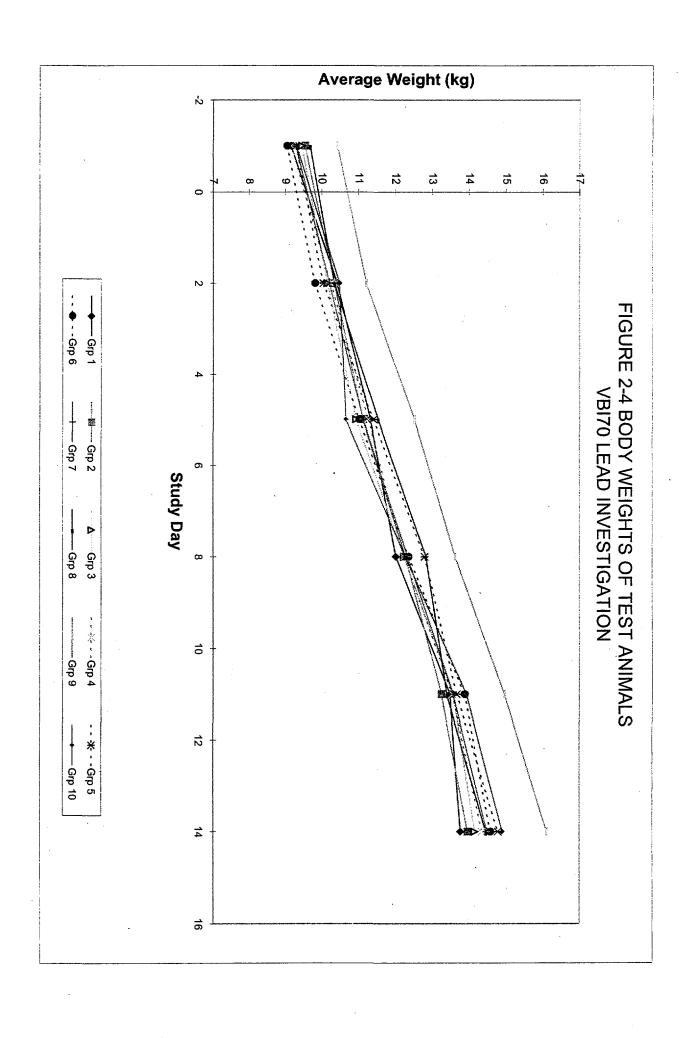


TABLE 2-2 TYPICAL FEED COMPOSITION^a

Nutrient Name	Amount	Nutrient Name	Amount
Protein	20.1021%	Chlorine	0.1911%
Arginine	1.2070%	Magnesium	0.0533%
Lysine	1.4690%	Sulfur	0.0339%
Methionine	0.8370%	Manganese	20.4719 ppm
Met+Cys	0.5876%	Zinc	118.0608 ppm
Tryptophan	0.2770%	Iron	135.3710 ppm
Histidine	0.5580%	Copper	8.1062 ppm
Leucine	1.8160%	Cobalt	0.0110 ppm
Isoleucine	1.1310%	Iodine	0.2075 ppm
Phenylalanine	1.1050%	Selenium	0.3196 ррт
Phe+Tyr	2.0500%	Nitrogen Free Extract	60.2340%
Threonine	0.8200%	Vitamin A	5.1892 kIU/kg
Valine	1.1910%	Vitamin D3	0.6486 kIU/kg
Fat	4.4440%	Vitamin E	87.2080 IU/kg
Saturated Fat	0.5590%	Vitamin K	0.9089 ррт
Unsaturated Fat	3.7410%	Thiamine	9.1681 ppm
Linoleic 18:2:6	1.9350%	Riboflavin	10,2290 ppm
Linoleic 18:3:3	0.0430%	Niacin	30.1147 ppm
Crude Fiber	3.8035%	Pantothenic Acid	19.1250 ppm
Ash	4.3347%	Choline	1019.8600 ppm
Calcium	0.8675%	Pyridoxine	8.2302 ppm
Phos Total	0.7736%	Folacin	2.0476 ррт
Available Phosphorous	0.7005%	Biotin	0.2038 ppm
Sodium	0.2448%	Vitamin B12	23.4416 ppm
Potassium	0.3733%		

^a Nutritional values provided by Zeigler Bros., Inc.

2.4 Dosing

The protocol for exposing animals to lead is shown in Table 2-3. The dose levels for lead acetate were based on experience from previous swine investigations that showed that doses of 25-225 ug Pb/kg/day gave clear and measurable increases in lead levels in all endpoints measured (blood, liver, kidney, bone). The doses of test materials were set at the same level as lead acetate, with one higher dose (500 ug Pb/kg-day) included in case the test materials were found to yield very low responses.

Animals were exposed to lead acetate or test material for 15 days, with the dose for each day being administered in two equal portions given at 9:00 AM and 3:00 PM (two hours before feeding). Doses were based on measured group mean body weights, and were adjusted every three days to account for animal growth. For animals exposed by the oral route, dose material was placed in the center of a small portion (about 5 grams) of moistened feed, and this was administered to the animals by hand. In this study, all doses were consumed by the animals without delay or spillage. However, on day 3, one animal in Group 8 was inadvertently given a dose for Group 9 in addition to it's own dose during the morning dosing. This dosing discrepancy was accounted for in further calculations.

Actual mean doses, calculated from the administered doses and the measured body weights, are also shown in Table 2-3.

2.5 Collection of Biological Samples

Samples of blood were collected from each animal on the first day of exposure (day 0), and on days 1, 2, 3, 5, 7, 9, 12, and 15 following the start of exposure. All blood samples were collected by vena-puncture of the anterior vena cava, and samples were immediately placed in purple-top Vacutainer® tubes containing EDTA as anticoagulant. Although EDTA is a chelator of metals, its presence in the sampling tubes will not impact the analytical results for lead. This is because the nitric acid digest used in the analysis destroys the organic constituents in the blood, thereby freeing all lead for analysis. Blood samples were collected each sampling day beginning at 8:00 AM, approximately one hour before the first of the two daily exposures to lead on the sampling day and 17 hours after the last lead exposure the previous day. This blood collection time was selected because the rate of change in blood lead resulting from the preceding exposures is expected to be relatively small after this interval (LaVelle et al. 1991, Weis et al. 1993), so the exact timing of sample collection relative to last dosing is not likely to be critical.

Following collection of the final blood sample at 8:00 AM on day 15, all animals were humanely euthanized and samples of liver, kidney and bone (the right femur) were removed and stored in lead-free plastic bags for lead analysis. Samples of all biological samples collected were archived in order to allow for reanalysis and verification of lead levels, if needed. All animals were also subjected to detailed examination at necropsy by a certified veterinary pathologist in order to assess overall animal health.

TABLE 2-3 DOSING PROTOCOL

	Number	Dose	_	Lead Dose ((ug Pb/kg-d)
Group	of Animals	Material Administered	Exposure Route	Target	Actual ^a
1	5	Lead Acetate	Oral	25	26.0
2	5	Lead Acetate	Oral	75	78.0
3	5	Lead Acetate	Oral	225	233.1
4	5	Eastern Sample (TM#1)	Oral	75	77.2
5	5	Eastern Sample (TM#1)	Oral	225	232.0
6	5	Eastern Sample (TM#1)	Oral	500	513.1
7	5	Western Sample (TM#2)	Oral	75	77.1
8	5	Western Sample (TM#2)	Oral	225	236.3
9	5	Western Sample (TM#2)	Oral	500	511.5
10	3 ^b	Control	Oral	0	0

Doses were administered in two equal portions given at 9:00 AM and 3:00 PM each day. Doses were based on the mean weight of the animals in each group, and were adjusted every three days to account for weight gain.

- ^a Calculated as the administered daily dose divided by the measured or extrapolated daily body weight, averaged over days 0-14 for each animal and each group.
- Three control animals were used in this study due to contraints on facility size. Based on previous investigations, this approach resulted in reliable results.

2.6 Preparation of Biological Samples for Analysis

Blood

One mL of whole blood was removed from the purple-top Vacutainer and added to 9.0 mL of "matrix modifier", a solution recommended by the Centers for Disease Control and Prevention (CDCP) for analysis of blood samples for lead. The composition of matrix modifier is 0.2% (v/v) ultrapure nitric acid, 0.5% (v/v) Triton X-100, and 0.2% (w/v) dibasic ammonium phosphate in deionized and ultrafiltered water. Samples of the matrix modifier were routinely analyzed for lead to ensure the absence of lead contamination.

Liver and Kidney

One gram of soft tissue (liver or kidney) was placed in a lead-free screw-cap teflon container with 2 mL of concentrated (70%) nitric acid and heated in an oven to 90°C overnight. After cooling, the digestate was transferred to a clean lead-free 10 mL volumetric flask and diluted to volume with deionized and ultrafiltered water.

Bone

The right femur of each animal was removed and defleshed, and dried at 100°C overnight. The dried bones were then placed in a muffle furnace and dry-ashed at 450°C for 48 hours. Following dry ashing, the bone was ground to a fine powder using a lead-free mortar and pestle, and 200 mg was removed and dissolved in 10.0 mL of 1:1 (v:v) concentrated nitric acid/water. After the powdered bone was dissolved and mixed, 1.0 mL of the acid solution was removed and diluted to 10.0 mL by addition of 0.1% (w/v) lanthanum oxide (La₂O₃) in deionized and ultrafiltered water.

2.7 Lead Analysis

Samples of biological tissue (blood, liver, kidney, bone) and other materials (food, water, reagents and solutions, etc.) were arranged in a random sequence and provided to the analytical laboratory in a blind fashion (identified to the laboratory only by a chain of custody tag number). Each sample was analyzed for lead using a Perkin Elmer Model 5100 graphite furnace atomic absorption spectrophotometer. Internal quality assurance samples were run every tenth sample, and the instrument was recalibrated every 15th sample. A blank, duplicate and spiked sample were run every 20th sample.

All results from the analytical laboratory were reported in units of ug Pb/L of prepared sample. The quantitation limit was defined as three-times the standard deviation of a set of seven replicates of a low-lead sample (typically about 2-5 ug/L). The standard deviation was usually about 0.3 ug/L, so the quantitation limit was usually about 0.9-1.0 ug/L (ppb). For prepared blood samples (diluted 1/10), this corresponds to a quantitation limit of 10 ug/L (1 ug/dL). For soft tissues (liver and kidney, diluted 1/10), this corresponds to a

quantitation limit of 10~ug/kg (ppb) wet weight, and for bone (final dilution = 1/500) the corresponding quantitation limit is 0.5~ug/g (ppm) ashed weight.

3.0 DATA ANALYSIS

3.1 Overview

Studies on the absorption of lead are often complicated because some biological responses to lead exposure may be non-linear functions of dose (i.e., tending to flatten out or plateau as dose increases). The cause of this non-linearity is uncertain but might be due either to non-linear absorption kinetics and/or to non-linear biological response per unit dose absorbed. When the dose-response curve for either the reference material (lead acetate) and/or the test material is non-linear, RBA is equal to the ratio of doses that produce equal responses (not the ratio of responses at equal doses). This is based on the simple but biologically plausible assumption that equal absorbed doses yield equal biological responses. Applying this assumption leads to the following general methods for calculating RBA from a set of non-linear experimental data:

- 1. Plot the biological responses of individual animals exposed to a series of oral doses of soluble lead (e.g., lead acetate). Fit an equation which gives a smooth line through the observed data points.
- 2. Plot the biological responses of individual animals exposed to a series of doses of test material. Fit an equation which gives a smooth line through the observed data.
- 3. Using the best fit equations for reference material and test material, calculate RBA as the ratios of doses of test material and reference material which yield equal biological responses. Depending on the relative shape of the best-fit lines through the lead acetate and test material dose response curves, RBA may either be constant (dose-independent) or variable (dose-dependent).

The principal advantage of this approach is that it is not necessary to understand the basis for a non-linear dose response curve (non-linear absorption and/or non-linear biological response) in order to derive valid RBA estimates. Also, it is important to realize that this method is very general, as it will yield correct results even if one or both of the dose-response curves are linear. In the case where both curves are linear, RBA is dose-independent and is simply equal to the ratio of the slopes of the best-fit linear equations.

3.2 Fitting the Curves

There are a number of different mathematical equations which can yield reasonable fits with the dose-response data sets obtained in this study. Conceptually, any equation which gives a smooth fit would be acceptable, since the main purpose is to allow for interpolation of responses between test doses. In selecting which equations to employ, the following principles were applied: 1) mathematically simple equations were preferred over mathematically complex equations, 2) the shape of the curves had to be smooth and

biologically realistic, without inflection points, maxima or minima, and 3) the general form of the equations had to be able to fit data not only from this one study, but from all the studies that are part of this project. After testing a wide variety of different equations, it was found that all data sets could be well fitted using one of the following three forms:

<u>Linear (LIN):</u> Response = a + b·Dose

Exponential (EXP): Response = $a + c \cdot (1-exp(-d \cdot Dose))$

<u>Combination (LIN+EXP)</u>: Response = $a + b \cdot Dose + c \cdot (1-exp(-d \cdot Dose))$

Although underlying mechanism was not considered in selecting these equations, the linear equation allows fitting data that do not show evidence of saturation in either uptake or response, while the exponential and mixed equations allow evaluation of data that appear to reflect some degree of saturation in uptake and/or response.

Each dose-response data set was fit to each of the equations above. If one equation yielded a fit that was clearly superior (as judged by the value of the adjusted correlation coefficient R²) to the others, that equation was selected. If two or more models fit the data approximately equally well, then the simplest model (that with the fewest parameters) was selected. In the process of finding the best-fits of these equations to the data, the values of the parameters (a, b, c, and d) were subjected to some constraints, and some data points (those that were outside the 95% prediction limits of the fit) were excluded. These constraints and outlier exclusion steps are detailed in Appendix A (Section 3). In general, most blood lead AUC dose-response curves were best fit by the exponential equation, and most dose-response curves for liver, kidney and bone were best fit by linear equations. In evaluating spleen results, it was determined that data were best fit by the exponential equation.

3.3 Responses Below Quantitation Limit

In some cases, most or all of the responses in a group of animals were below the quantitation limit for the endpoint being measured. For example, this was normally the case for blood lead values in unexposed animals (both on day -4 and day 0), and in control animals. In these cases, samples were assigned a response equal to one-half the quantitation limit.

3.4 Quality Assurance

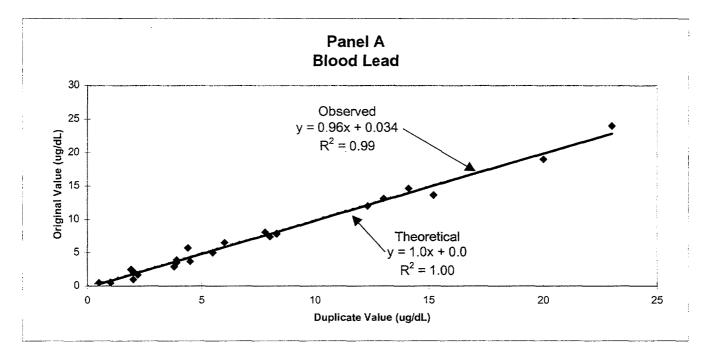
A number of steps were taken throughout this study and the other studies in this project to ensure the quality of the results. These steps are summarized below.

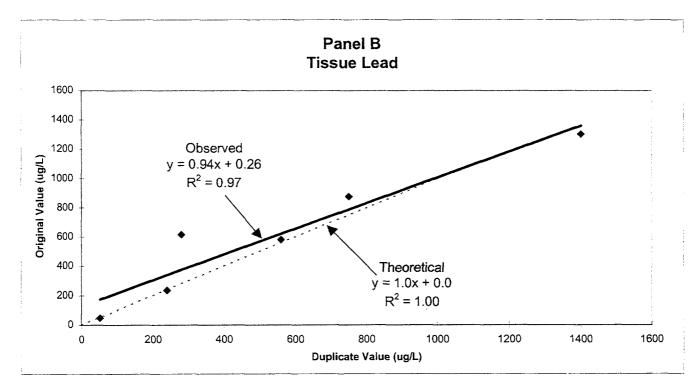
Duplicates

A randomly selected set of about 5% of all samples generated during the study were submitted to the laboratory in a blind fashion for duplicate analysis. The raw data are presented in Appendix A, and Figure 3-1 plots the results for blood (Panel A, upper) and for bone, liver, and kidney (Panel B, lower).

FIGURE 3-1 COMPARISION OF DUPLICATE ANALYSES

VBI70 LEAD





As seen, there was good intra-laboratory reproducibility between duplicate samples for both blood and tissues, with linear regression lines having a slope near 1.0, an intercept near zero, and an R² value near 1.0. One blood sample (not represented in the graph) was determined to be an outlier (original value 6.5; duplicate value 28.7).

Standards

The Centers for Disease Control and Prevention (CDCP) provides a variety of blood lead "check samples" for use in quality assurance programs for blood lead studies. Each time a group of blood samples was prepared and sent to the laboratory for analysis, several CDCP check samples of different concentrations were included in random order and in a blind fashion.

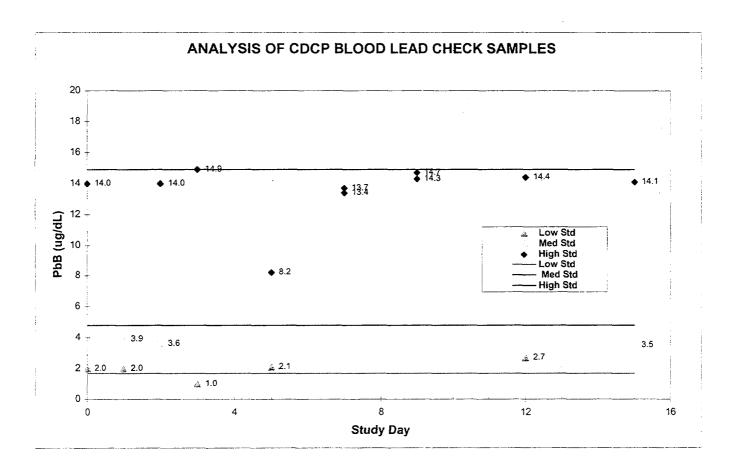
The results for the samples submitted during this study are presented in Appendix A, and the values are plotted in Figure 3-2. For the "low" standard (nominal = 1.7 ug/dL), the average measured value was 2.0 ug/dL. For the "medium" and "high" standards, the means of the measured values were 3.7 ug/dL (nominal = 4.8 ug/dL) and 13.6 ug/dL (nominal = 14.9 ug/dL).

Data Audits and Spreadsheet Validation

All analytical data generated by EPA's analytical laboratory were validated prior to being released in the form of a database file. These electronic data files were "decoded" (linking the sample tag to the correct animal and day) using Microsoft's database system ACCESS®. To ensure that no errors occurred in this process, original electronic files were printed out and compared to printouts of the tag assignments and the decoded data.

All spreadsheets used to manipulate the data and to perform calculations (see Appendix A) were validated by hand-checking random cells for accuracy.

FIGURE 3-2 CDCP CHECK SAMPLES VBI70 LEAD



4.0 RESULTS

The following sections provide results based on the group means for each dose group investigated in this study. Appendix A provides detailed data for each individual animal. Results from this study will be compared and contrasted with the results from other studies in a subsequent report.

4.1 Blood Lead vs Time

Figure 4-1 shows the group mean blood lead values as a function of time during the study. As seen, blood lead values began at or below quantitation limits (about 1 ug/dL) in all groups, and remained at or below quantitation limits in control animals (Group 10). In animals given repeated oral doses of lead acetate (Groups 1-3), Eastern soil (Groups 4-6), or Western soil (Groups 7-9), blood levels began to rise within 1-2 days, and tended to plateau by the end of the study (day 15).

4.2 Dose-Response Patterns

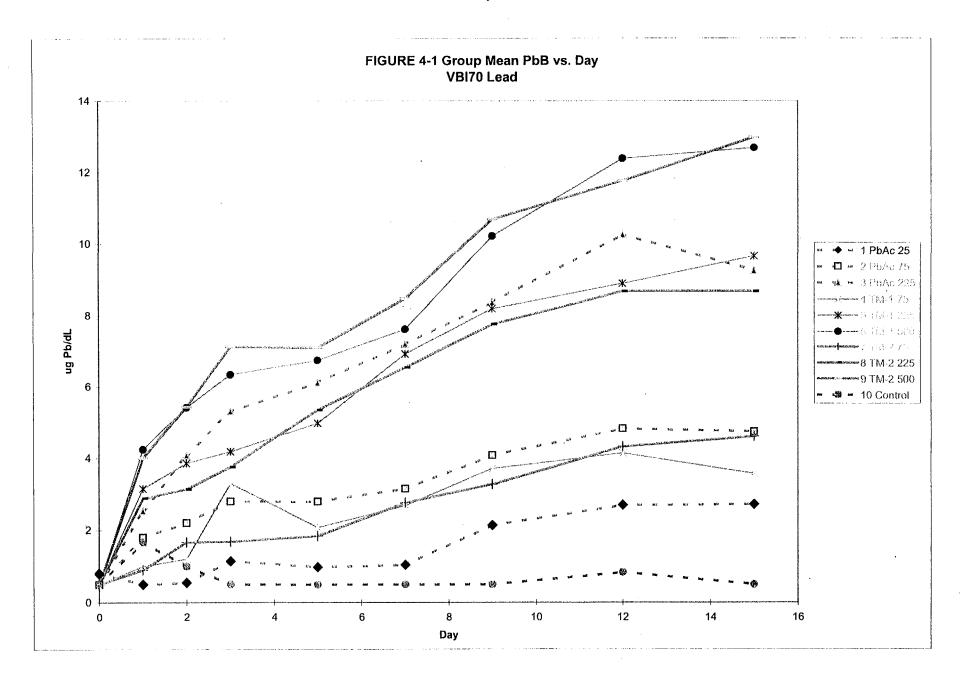
Blood Lead

The measurement endpoint used to quantify the blood lead response was the area under the curve (AUC) for blood lead vs time (days 0-15). AUC was selected because it is the standard pharmacokinetic index of chemical uptake into the blood compartment, and is relatively insensitive to small variations in blood lead level by day. The AUC was calculated using the trapezoidal rule to estimate the AUC between each time point that a blood lead value was measured (days 0, 1, 2, 3, 5, 7, 9, 12, and 15), and summing the areas across all time intervals in the study. The detailed data and calculations are presented in Appendix A, and the results are shown graphically in Figure 4-2. Each data point reflects the group mean exposure and group mean response, with the variability in dose and response shown by standard error bars. The figure also shows the best-fit equation through each data set.

As seen, the dose response pattern is non-linear for both the soluble reference material (lead acetate, abbreviated "PbAc"), and for each of the two test soils. Dose response curves for both soils are similar to those seen for lead acetate.

Tissue Lead

The dose-response data for lead levels in bone, liver, and kidney (measured at sacrifice on day 15) are detailed in Appendix A, and are shown graphically in Figures 4-3 through 4-5, respectively. As seen, all of these dose response curves for tissues are fit by linear equations, both for lead acetate and each of the two test soils.



Page 1

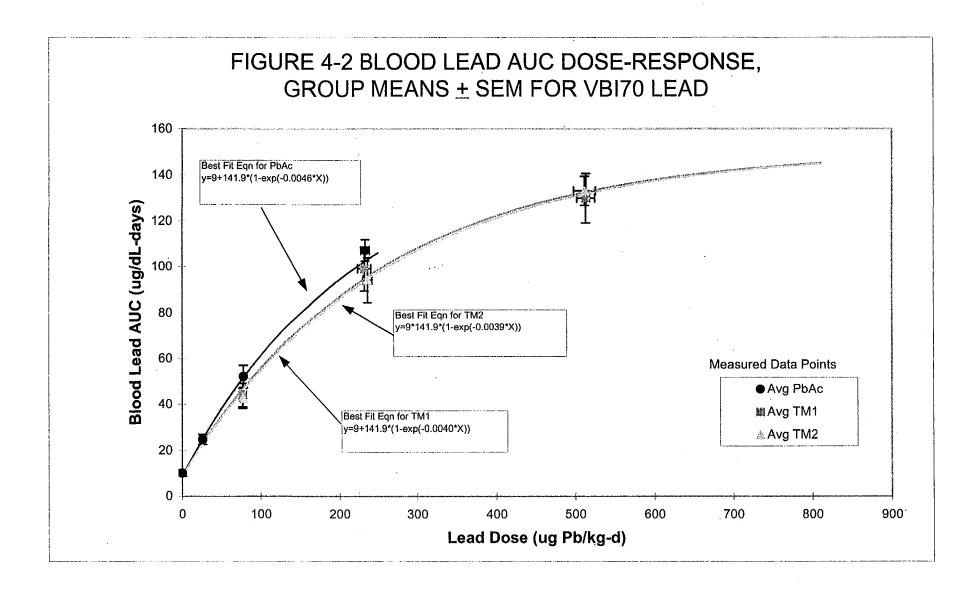
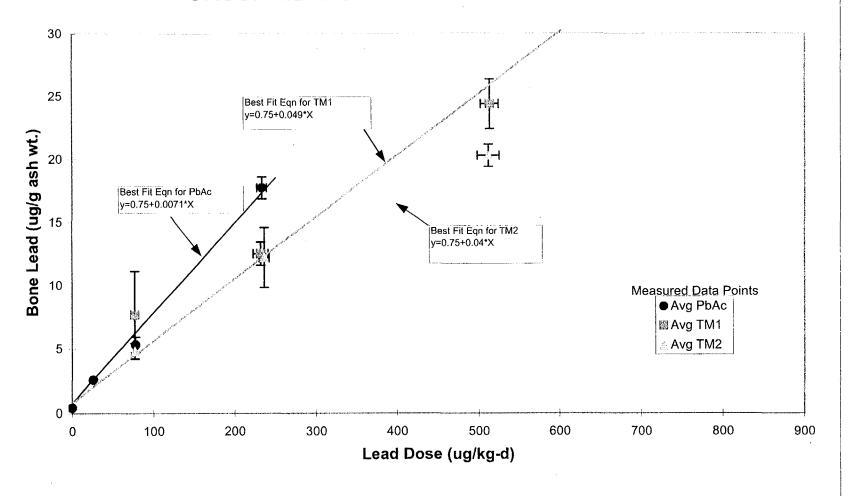
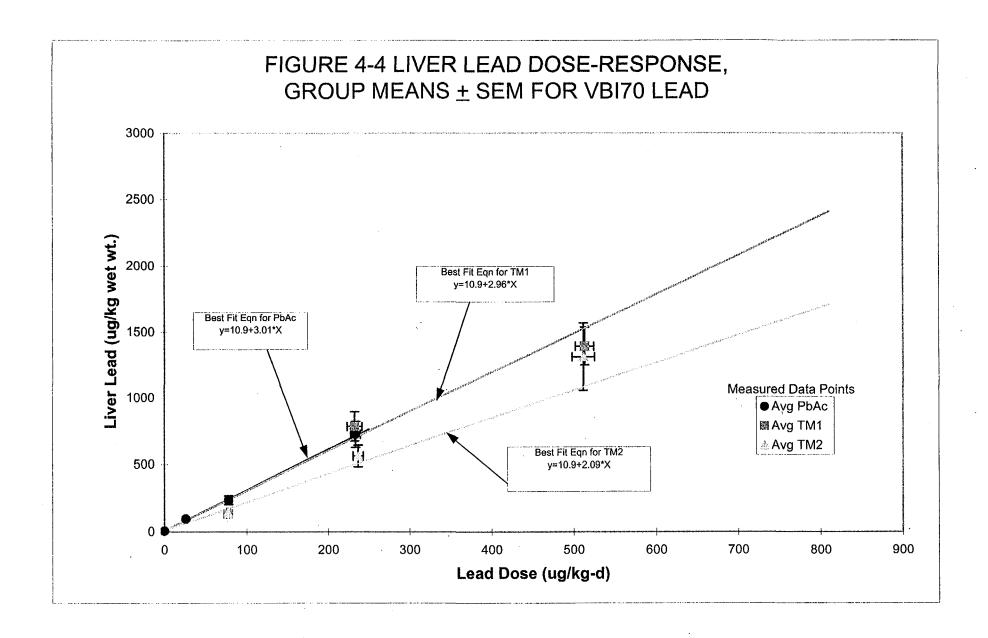
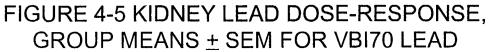
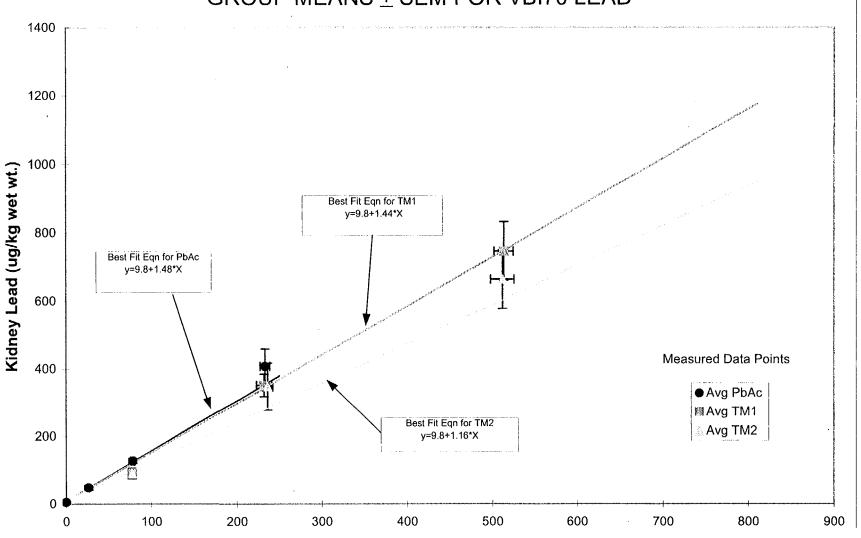


FIGURE 4-3 BONE LEAD DOSE-RESPONSE, GROUP MEANS + SEM FOR VBI70 LEAD









4.3 Calculated RBA Values

Relative bioavailability values were calculated for each test material for each measurement endpoint (blood AUC, bone, liver, kidney) using the method described in Section 3.0. The results are shown below:

	Estimated RBA		
Measurement Endpoint	Eastern Test Material #1	Western Test Material #2	
Blood Lead AUC	0.87	0.85	
Liver Lead	0.98	0.70	
Kidney Lead	0.97	0.78	
Bone Lead	0.69	0.56	

Recommended RBA Values

For each test soil, the estimates of RBA based on blood, liver, kidney, and bone are generally similar, but do not agree exactly in all cases. In general, we recommend greatest emphasis be placed on the RBA estimates derived from the blood lead data. There are several reasons for this recommendation, including the following:

- 1) Blood lead calculations are based on multiple measurements over time, and so are statistically more robust than the single measurements available for tissue concentrations. Further, blood is a homogeneous medium, and is easier to sample than complex tissues such as liver, kidney and bone. Consequently, the AUC endpoint is less susceptible to random measurement errors, and RBA values calculated from AUC data are less uncertain.
- 2. Blood is the central compartment and one of the first compartments to be affected by absorbed lead. In contrast, uptake of lead into peripheral compartments (liver, kidney, bone) depend on transfer from blood to the tissue, and may be subject to a variety of toxicokinetic factors that could make bioavailability determinations more complicated.
- 3. The dose-response curve for blood lead is non-linear, similar to the non-linear dose-response curve observed in children (e.g., see Sherlock and Quinn 1986). Thus, the response of this endpoint is known to behave similarly in swine as in children, and it is not known if the same is true for the tissue endpoints.

4. Blood lead is the classical measurement endpoint for evaluating exposure and health effects in humans, and the health effects of lead are believed to be proportional to blood lead levels.

However, data from the tissue endpoints (liver, kidney, bone) also provide valuable information. We consider the <u>plausible range</u> to extend from the RBA based on blood AUC to the mean of the other three tissues (liver, kidney, bone). The <u>preferred range</u> is the interval from the RBA based on blood to the mean of the blood RBA and the tissue mean RBA. Our <u>suggested point estimate</u> is the mid-point of the preferred range. These values are presented below:

Relative	Test Material		
Bioavailability of Lead	Eastern Test Material #1	Western Test Material #2	
Plausible Range	0.87-0.88	0.68-0.85	
Preferred Range	0.87-0.88	0.76-0.85	
Suggested Point Estimate	0.87	0.81	

4.4 Estimated Absolute Bioavailability in Children

These RBA estimates may be used to help assess lead risk at this site by refining the estimate of absolute bioavailability (ABA) of lead in soil, as follows:

$$ABA_{soil} = ABA_{soluble} * RBA_{soil}$$

Available data indicate that fully soluble forms of lead are about 50% absorbed by a child (USEPA 1991, 1994). Thus, the estimated absolute bioavailability of lead in the site samples is calculated as follows:

$$ABA_{Site} = 50\% * RBA_{Site}$$

Based on the RBA values shown above, the estimated absolute bioavailability in children is as follows:

Absolute	Test Material		
Bioavailability of Lead	Eastern Test Material #1	Western Test Material #2	
Plausible Range	0.43-0.44	0.34-0.42	
Preferred Range	0.43-0.44	0.38-0.42	
Suggested Point Estimate	0.44	0.40	

4.5 Uncertainty

The bioavailability estimates above are subject to uncertainty that arises from several different sources. First, differences in physiological and pharmacokinetic parameters between individual animals leads to variability in response, even when exposure is the same. Because of this inter-animal variability in the responses of different animals to lead exposure, there is mathematical uncertainty in the best fit dose-response curves for both lead acetate and test material. This in turn leads to uncertainty in the calculated values of RBA, because these are derived from the two best-fit equations. Second, there is uncertainty in how to weight the RBA values based on the different endpoints, and how to select a point estimate for RBA that is applicable to typical site-specific exposure levels. Third, there is uncertainty in the quantitative extrapolation of measured RBA values in swine to young children. Even though the immature swine is believed to be a useful and meaningful animal model for gastrointestinal absorption in children, it is possible that differences in stomach pH, stomach emptying time, and other physiological parameters may exist and that RBA values in swine may not be precisely equal to values in children. Finally, studies in humans reveal that lead absorption is not constant even within an individual, but varies as a function of many factors (mineral intake, health status, etc.). One factor that may be of special importance is time after the last meal, with the presence of food tending to reduce lead absorption. The values of RBA measured in this study are intended to estimate the maximum uptake that occurs when lead is ingested in the absence of food. Thus, these values may be somewhat conservative for children who ingest lead along with food. The magnitude of this bias is not known, although preliminary studies in swine suggest the factor may be relatively minor.

5.0 REFERENCES

Gibaldi, M. and Perrier, D. 1982. *Pharmacokinetics* (2nd edition) pp 294-297. Marcel Dekker, Inc, NY, NY.

Goodman, A.G., Rall, T.W., Nies, A.S., and Taylor, P. 1990. *The Pharmacological Basis of Therapeutics* (8th ed.) pp. 5-21. Pergamon Press, Inc. Elmsford, NY.

Klaassen, C.D., Amdur, M.O., and Doull, J. (eds). 1996. *Cassarett and Doull's Toxicology: The Basic Science of Poisons*. pp. 190. McGraw-Hill, Inc. NY,NY

LaVelle, J.M., Poppenga, R.H., Thacker, B.J., Giesy, J.P., Weis, C., Othoudt R, and Vandervoot C. 1991. Bioavailability of Lead in Mining Waste: An Oral Intubation Study in Young Swine. In: <u>The Proceedings of the International Symposium on the Bioavailability and Dietary Uptake of Lead.</u> Science and Technology Letters 3:105-111.

Mushak, P. 1991. Gastro-intestinal Absorption of Lead in Children and Adults: Overview of Biological and Biophysico-chemical Aspects. In: <u>The Proceedings of the International Symposium on the Bioavailability and Dietary Uptake of Lead.</u> Science and Technology Letters 3:87-104.

Sherlock, J.C., and Quinn, M.J. 1986. Relationship Between Blood Lead Concentration and Dietary Intake in Infants: the Glasgow Duplicate Diet Study 1979-1980. Food Additives and Contaminants 3:167-176.

USEPA 1991. Technical Support Document on Lead. United States Environmental Protection Agency, Environmental Criteria and Assessment Office. ECAO-CIN-757.

USEPA 1994. Guidance Manual for the Integrated Exposure Uptake Biokinetic Model for Lead in Children. United States Environmental Protection Agency, Office of Emergency and Remedial Response. Publication Number 9285.7-15-1. EPA/540/R-93/081.

USEPA 2000. Quality Assurance Project Plan for Vaquez BLVD-I70 Bioavailability of Lead in Site Soils Using Juvenile Swine as an Animal Model. Prepared by Syracuse Research Corporation.

Washington Group 2000. Technical Memorandum. VB/I-70 Bioavailability Soil Sample Preparation. From Kevin Williamson to Bonnie Lavelle.

Weis, C.P. and LaVelle, J.M. 1991. Characteristics to consider when choosing an animal model for the study of lead bioavailability. In: <u>The Proceedings of the International Symposium on the Bioavailability and Dietary Uptake of Lead.</u> Science and Technology Letters 3:113-119.

Weis, C.P., Henningsen, G.M., Poppenga, R.H., and Thacker, B.J. 1993. Pharmacokinetics of Lead in Blood of Immature Swine Following Acute Oral and Intravenous Exposure. The Toxicologist 13(1):175.

Weis, C.P., Poppenga, R.H., Thacker, B.J., Henningsen, G.M., and Curtis, A. 1995. "Design of Pharmacokinetic and Bioavailability Studies of Lead in an Immature Swine Model." In: LEAD IN PAINT, SOIL, AND DUST: HEALTH RISKS, EXPOSURE STUDIES, CONTROL MEASURES, MEASUREMENT METHODS, AND QUALITY ASSURANCE. ASTM STP 1226, Michael E. Beard and S. D. Allen Iske, Eds., American Society for Testing and Materials, Philadelphia, 1995.

Yang, J. and Blanchar, B. 1997a. Soil Characterization and Physiological-Based Extraction Test (PBET) of the Phosphorus Treated Soils. Department of Soil and Atmospheric Sciences. University of Missouri-Columbia. January 15, 1997.

Yang, J. and Blanchar, B. 1997b. Preliminary Report for Phosphoric Acid Treatment to Reduce Pb Bioavailability in Joplin Soils. Department of Soil and Atmospheric Sciences. University of Missouri-Columbia. February 1997.

APPENDIX A

DETAILED DATA AND CALCULATIONS

APPENDIX A

DETAILED DATA SUMMARY

1.0 OVERVIEW

Performance of this study involved collection and reduction of a large number of data items. All of these data items and all of the data reduction steps are contained in a Microsoft Excel spreadsheet named "VB_LEAD.XLS". This file is intended to allow detailed review and evaluation of all aspects of this study.

The following sections of this Appendix present printouts of selected tables and graphs from the XLS file. These tables and graphs provide a more detailed documentation of the individual animal data and the data reduction steps performed in this study than was presented in the main text. Any additional details of interest to a reader can be found in the XLS spreadsheet.

2.0 RAW DATA AND DATA REDUCTION STEPS

2.1 Body Weights and Dose Calculations

Animals were weighed on day -1 (one day before exposure) and every three days thereafter during the course of the study. Doses of lead for the three days following each weighing were based on the group mean body weight, adjusted by addition of 1 kg to account for the expected weight gain over the interval. After completion of the experiment, body weights were estimated by interpolation for those days when measurements were not collected, and the actual administered doses (ug Pb/kg) were calculated for each day and then averaged across all days. If an animal missed a dose or was given an incorrect dose, the calculation of average dose corrected for these factors. These data and data reduction steps are shown in Tables A-1 and A-2. During this study, one animal in Group 8, was inadvertently administered a Group 9 doughball in addition to its assigned doughball during the morning dosing on Day 3. This misdosing was accounted for in the spreadsheets for this experiment.

2.2 Blood Lead vs Time

Blood lead values were measured in each animal on days 0, 1, 2, 3, 5, 7, 9, 12, and 15. The raw laboratory data (reported as ug/L of diluted blood) are shown in Table A-3. These data were adjusted as follows: a) non-detects were evaluated by assuming a value equal to one-half the quantitation limit, and b) the concentrations in diluted blood were converted to units of ug/dL in whole blood by dividing by a factor of 1 dL of blood per L of diluted sample. The results are shown in the right-hand column of Table A-3. Figures A-1

to A-3 plot the results for individual animals organized by group and by day. Figure A-4 plots the mean for each dosing group by day.

After adjustment as above, values that were more than a factor of 1.5 above or below the group mean for any given day were "flagged" by computer as potential outliers. These values are shown in Table A-4 by cells that are shaded gray. Each data point identified in this way was reviewed and professional judgment was used to decide if the value should be retained or excluded. In order to avoid inappropriate biases, blood lead outlier designations were restricted to values that were clearly aberrant from a time-course and/or dose-response perspective. Values which were excluded are identified by a heavy black box outlining the values. Rationale for outlier exclusion is provided in Table A-5.

2.3 Blood Lead AUC

The area under the blood lead vs time curve for each animal was calculated by finding the area under the curve for each time step using the trapezoidal rule:

$$AUC(d_i \text{ to } d_j) = 0.5*(r_i + r_j)*(d_j - d_i)$$

where:

d = day number

r = response (blood lead value) on day i (r_i) or day j (r_i)

The areas were then summed for each of the time intervals to yield the final AUC for each animal. These calculations are shown in Table A-6. If a blood lead value was missing (either because of problems with sample preparation, or because the measured value was excluded as an outlier), the blood lead value for that day was estimated by linear interpolation.

2.4 Liver, Kidney, and Bone Lead Data

At sacrifice (day 15), samples of liver, kidney, and bone (femur) were removed and analyzed for lead. The raw data (expressed as ug Pb/L of prepared sample) are summarized in Table A-7. These data were adjusted as follows: a) non-detects were evaluated by assuming a value equal to one-half the quantitation limit, and b) the concentrations in prepared sample were converted to units of concentration in the original biological sample by dividing by the following factors:

Liver:

0.1 kg wet weight/L prepared sample

Kidney:

0.1 kg wet weight/L prepared sample

Bone:

2 gm ashed weight/L prepared sample

The resulting values are shown in the right-hand column of Table A-7.

3.0 CURVE FITTING

Basic Equations

A commercial curve-fitting program (Table Curve-2D™ Version 2.0 for Windows, available from Jandel Scientific) was used to derive best fit equations for each of the individual doseresponse data sets derived above. A least squares regression method was used for both linear and non-linear equations. As discussed in the text, three different user-defined equations were fit to each data set:

<u>Linear (LIN):</u> Response = $a + b \cdot Dose$

Exponential (EXP): Response = $a + c \cdot (1-exp(-d \cdot Dose))$

Combination (LIN+EXP): Response = $a + b \cdot Dose + c \cdot (1-exp(-d \cdot Dose))$

Constraints

In the process of finding the best-fits of these equations to the data, the values of the parameters (a, b, c, and d) were constrained as follows:

- Parameter "a" (the intercept, equal to the baseline or control value of the measurement endpoint) was constrained to be non-negative and was forced in all cases to be the same for the reference material (lead acetate) and the test materials. This is because, by definition, all dose-response curves for groups of animals exposed to different materials must arise from the same value at zero dose. In addition, for blood lead data, "a" was constrained to be equal to the mean of the control group ± 20% (typically 7.5 ± 1.5 AUC units).
- Parameter "b" (the slope of the linear dose-response line) was constrained to non-negative values, since all of the measurement endpoints evaluated are observed to increase, not decrease, as a function of lead exposure.
- Parameter "c" (the plateau value of the exponential curve) was constrained to be non-negative, and was forced to be the same for the reference material (lead acetate) and the test material. This is because: 1) it is expected on theoretical grounds that the plateau (saturation level) should be the same regardless of the source of lead, and 2) curve-fitting of individual curves tended to yield values of "c" that were close to each other and were not statistically different.
- Parameter "d" (which determines where the "bend" in the exponential equation occurs) was constrained to be greater than 0.0045 for the lead

acetate blood lead (AUC) dose-response curve. This constraint was judged to be necessary because the weight of evidence from all studies clearly showed the lead acetate blood lead dose response curve was non-linear and was best fit by an exponential equation, but in some studies there were only two low doses of lead acetate used to define the dose-response curve, and this narrow range data set could sometimes be fit nearly as well by a linear as an exponential curve. The choice of the constraint on "d" was selected to be slightly lower than the observed best-fit value of "d" (0.006) when data from all lead acetate AUC dose-response curves from all of the different studies in this program were used. This approach may tend to underestimate relative bioavailability slightly in some studies (especially at low dose), but use of the information gained from all studies is judged to be more robust than basing fits solely on the data from one study.

In general, one of these models (the linear, the exponential, or the combination) usually yielded a fit (as judged by the value of the adjusted correlation coefficient R² and by visual inspection of the fit of the line through the measured data points) that was clearly superior to the others. If two or more models fit the data approximately equally well, then the simplest model (that with the fewest parameters) was selected.

Outlier Identification

During the dose-response curve fitting process, all data were carefully reviewed to identify any anomalous values. Typically, the process used to identify outliers was as follows:

- Step 1 Any data points judged to be outliers based on information derived from analysis of data across multiple studies (as opposed to conclusions drawn from within the study) were excluded (a priori outliers).
- Step 2 The remaining raw data points were fit to the equation judged to be the most likely to be the best fit (linear, exponential, or mixed). Table Curve 2-D was then used to plot the 95% prediction limits around the best fit line. All data points that fell outside the 95% prediction limits were considered to be outliers and were excluded.
- Step 3 After excluding these points (if any), a new best-fit was obtained. In some cases, data points originally inside the 95% prediction limits were now outside the limits. However, further iterative cycles of data point exclusion were not performed, and the fit was considered final.

It should be noted that professional judgment can be imposed during any stage of the above outlier identification process. Table A-8 shows outliers selected using professional judgement.

Curve Fit Results

Table A-8 lists the data used to fit these curves, indicating which endpoints were excluded as outliers and why. Table A-9 shows the type of equation selected to fit each data set, and the best fit parameters. The resulting best-fit equations for the data sets are shown in Figures A-4 to A-15. Values excluded as outliers are represented in the figures by the symbol "+".

4.0 RESULTS -- CALCULATED RBA VALUES

The value of RBA for a test substance was calculated for a series of doses using the following procedure:

- 1. For each dose, calculate the expected response to test material, using the best fit equation through the dose-response data for that material.
- 2. For each expected response to test material, calculate the dose of lead acetate that is expected to yield an equivalent response. This is done by "inverting" the dose-response curve for lead acetate, solving for the dose that corresponds to a specified response.
- 3. Calculate RBA at that dose as the ratio of the dose of lead acetate to the dose of test material. For the situation where both curves are linear, the value of RBA is the ratio of the slopes (the "b" parameters). In the case where both curves are exponential and where both curves have the same values for parameters "a" and "c", the value of RBA is equal to the ratio of the "d" parameters.

The results are summarized in Table A-10.

5.0 QUALITY ASSURANCE DATA

A number of steps were taken throughout this study and the other studies in this project to ensure the quality of the results, including 5% duplicates, 5% standards, a program of interlaboratory comparison. These steps are detailed below.

Duplicates

Duplicate samples were prepared and analyzed for about 5% of all samples generated during the study. Table A-11 lists the first and second values for blood, liver, kidney, and bone. The results are shown in Figure 3-1 in the main text.

Standards

The Centers for Disease Control and Prevention (CDCP) provides a variety of blood lead "check samples" for use in quality assurance programs for blood lead studies. Each time a group of blood samples was prepared and sent to the laboratory for analysis, several CDCP check samples of different concentrations were included. Table A-12 lists the concentrations reported by the laboratory compared to the nominal concentrations indicated by CDCP for the samples submitted during this study, and the results are plotted in Figure 3-2 in the main text.

TABLE A-1 BODY WEIGHTS AND ADMINISTERED DOSES, BY DAY

Body weights were measured on days -1, 2, 5, 8, 11, 14. Weights for other days are estimated, based on linear interpolation between measured values.

Group	ID#	Da	y -1	1	Day 0	Τō	av 1	I D	ay 2	Da	y 3	Di	ay 4	Da	y 5	Di	y 6	Da	ij 7	Da	8 vs	Da	ıy 9	Da	v 10	Dav	v 11	Da	y 12	Da	y 13	Day	/ 14	Day	/ 15
1		BW	ug Pb	8W	ug Pb	BW	ug Pb		ug Pb	BW	ug Pb		ug Pb		ug Pb	BW	ug Pb	вw	ug Pb	8W	ug Pb	BW	ug Pb	BW	ug Pb	BW	ug Pb	BW	ug Pb	BW	ug Pb	BW	ug Pb	8W	ug Pb
L		(kg)	per day	(kg)	perday	(kg)	perday	(kg)	per day	(kg)	per day	(kg)	per day	(kg)	perday	(kg)	per day	(kg)	per day	(kg)	per day	(kg)	per day	(kg)	per day	(kg)	per day	(kg)	per day	(kg)	per day	(kg)	per day	(kg)	per day
1	311	9.5	0	10.0	254	10.5	254	11.06	254	11.4	285	11.7	285	11.95	285	12.0	308	12.1	308	12.23	308	12.8	325	13.4	325	13.98	325	13.9	362	13.9	362	13.88	362	13.8	0
1 1	326	9.04	0	9.6	254	10.1	254	10.66	254	10.9	285	11.1	285	11.36	285	11.5	308	11.7	308	11.85	308	12.4	325	13.0	325	13.62	325	13.7	362	13.7	362	13.8	362	13.9	0
1 1	338 -	9.76	0	10.1	254	10.4	254	10.75	254	11.4	285	12.1	285	12.84	285	13.1	308	13.3	308	13.53	308	13.9	325	14.3	325	14.66	325	14.7	362	14.8	362	14.65	362	14.9	0
1	343	8.34	0	8.6	254	8.8	254	9.07	254	9.2	285	9.4	285	9.5	285	9.8	308	10.1	308	10.35	308	10.8	325	11.3	325	11.77	325	12.0	362	12.2	362	12.35	362	12.5	0
1	349	9.17	. 0	9.6	254	10.1	254	10.54	254	10.7	285	10.8	285	10.96	285	11.3	308	11.7	308	12.1	308	12.5	325	13.0_	325	13.4	325	13.6	362	13.8	362	13.96	362	14.1	0
2	304	9.89	0	10.4	791	10.9	791	11.33	791	11.6	851	11.9	851	12.12	851	12.6	909	13.0	909	13.5	909	13.8	995	14.0	995	14.29	995	14.5	1069	14.8	1069	15,04	1069	15.3	0
2	307	9.47	0	9.6	791	9.8	791	9.9	791	10.0	851	10.1	851	10.26	851	10.8	909	11.3	909	11.81	909	12.2	995	12.6	995	13.05	995	13.1	1069	13.2	1069	13.3	1069	13.4	0
2	312	9.04	0	9.3	791	9.6	791	9.82	791	9,9	851	10.1	851	10.18	851	10.5	909	10.8	909	11.06	909	11.4	995	11.7	995	12.04	995	12.3	1069	12.7	1069	12.96	1069	13.3	0
2	314	9.57	0	9.7	791	9.9	791	10.08	791	10.4	851	10.6	851	11.16	851	11.5	909	11.9	909	12.2	909	12.4	995	12.6	995	12.85	995	13.2	1069	13.6	1069	13.9	1069	14.3	0
2	350	9.75	0	10.0		10.3	791	10.58		11.0	851	11.4	851	11.85	851	12.2	909	12.5	909	12.78	909	13.2	995	13.6	995	14.02	995	14.2	1069	14.4	1069	14.61	1069	14.8	0
3	302	8.77	0	9.0	2353	9.3	2353	9.53	2353	9.8	2509	10.1	2509	10.4	2509	10.8	2684	11.2	2684	11.64	2684	12.1	2977	12.5	2977	12.92	2977	13.1	3279	13.3	3279	13,5	3279	13.7	0
3	303	10.85	0	11.1	2353	11.4	2353	11.64		11.7	2509	11.7	2509	11.68	2509	12.1	2684	12.6	2684	13.08	2684	13.6	2977	14.1	2977	14.61	2977	14.5	3279	14,5	3279	14.38	3279	14.3	0
3	322	9.2	0	9.5	2353	9.9	2353	10.19		10.7	2509	11.2	2509	11.72	2509	12.0	2684	12.3	2684	12.53	2684	13.0	2977	13.5	2977	13:95	2977	14.1	3279	14.3	3279	14.45	3279	14.6	0
3	327	9.32	0	9.3	2353	9.3	2353	9.28	2353	9.6	2509	9.9	2509	10.23	2509	10.8	2684	11.3	2684	11.79	2684	12.2	2977	12.6	2977	12.95	2977	13.2	3279	13.5	3279	13.81	3278	14.1	0
3	342	9.14	0	9.5	2353	9.8	2353	10.12		10.3	2509	10.4	2509	10.61	2509	11.1	2684	11.6	2684	12.11	2684	12.6	2977	13.0	2977	13.44	2977	13.8	3279	14.2	3279	14.64	3279	15.0	0
4	313	9,55	0	10.1	762	10.6	762	11.19		11.4	843	11.6	843	11.73	843	11.9	920	12.1	920	12.3	920	12.9	992	13.5	992	14.03	992	14.3	1091	14.5	1091	14.7	1091	14.9	0
4	328	8.6	0	8.0	762	9.5	762	9.89	762	10.4	843	10.9	843	11.35	843	11.6	920	11.9	920	12.18	920	12.6	992	13.0	992	13.4	992	13.5	1091	13.7	1091	13.83	1091	14.0	0
4	329	8.8	0	9.0	762	9.3	762	9.54	762	9.9	843	10.2	843	10.57	843	11.0	920	11.5	920	11.9	920	12.4	992	13.0	992	13.48	992	13.7	1091	14.0	1091	14.21	1091	14.5	0
4	333	8.6	0	8.8	762	9.0	762	9.27	762	9.8	843	10.2	843	10.71	843	11.0	920	11.3	920	11.63	920	12.0	992	12.4	992	12.74	992	13.1	1091	13.4	1091	13.78	1091	14,1	0
4	348	10.26		10,6		11.0	762	11.34	762	11.5	843	11.6	843	11.96	843	12.4	920	12.7	920	13.14	920	13.5	992	13.8	992	14,11	992	14.5	1091	14.9	1091	15.23	1091	15.6	
5	317	8.42	0	8.6	2318	8.9	2318	9.09	2318	9.7	2485	10.3	2485	10.92	2465	11.5	2789	12.0	2789	12.52	2789	12.9	3103	13.3	3103	13.66	3103	14.2	3301	14.7	3301	15.22	3301	15.7	0
5	324	8.23	0	8.8	2318	9.3	2318	9.83	2318	10.0	2485	10.3	2485	10.46	2485	10.9	2789	11.4	2789	11.89	2789	12.2	3103	12.5	3103	12.75	3103	12.8	3301	12.9	3301	13.04	3301	13.1	0
5	340	9.5	0	9.6	2318	9.7	2318	9.86	2318	10.5	2485	11.1	2485	11.77	2485	12.0	2789	12.3	2789	12.52	2789	12.9	3103	13.3	3103	13.67	3103	14.1	3301	14.5	3301	14.97	3301	15.4	0
5	341	9.1	0	9.3	2318	9.4	2318	9.56	2318	9.9	2485	10.2	2485	10.53	2485	11.3	2789	12.1	2789	12.9	2789	12.7	3103	12.4	3103	12.2	3103	12.6	3301	13.1	3301	13.5	3301	13.9	0
5	354	11.25	0	11.5		11.7	2318	11.88		12.4	2485	12.8	2485	13.29	2485	13.6	2789	13,9	2789	14.13	2789	14.8	3103	15.4	3103	16.07	3103	16.4	3301	16.8	3301	17.17	3301	17.5	0
6	321	8.5	0	8.8	5024	9.1	5024	9.35	5024	9.8	5409	10.2	5409	10.61	5409	11.2	6005	11.7	6005	12.27	6005	12.5	6678	12.8	6678	13.01	6678	13.3	7441	13.5	7441	13,74	7441	14.0	0 1
8	332	10.1	0	10.1	5024	10.1	5024	10.11		10.3	5409	10.4	5409	10.6	5409	11.2	6005	11.8	6005	12.4	6005	13.0	6676	13.5	6678	14.11	6678	14.3	7441	14.4	7441	14.61	7441	14.8	0 1
6	335	10.03	ō	10.1	5024	10.3	5024	10.38		10.9	5409	11.4	5409	11.96	5409	12.3	6005	12.7	6005	13.05	6005	13.7	6678	14.4	6678	15.02	6678	15.4	7441	15.7	7441	16.06	7441	16.4	0 1
1 2	355	7.81	Ü	8.3	5024	8.9	5024	9.42		9.7	5409	10.1	5409	10.38	5409	10.9	6005	11.3	6005	11.81	6005	12.5	6678	13.1	6678	13.73	6678	13.8	7441	13.9	7441	13.93	7441	14.0	
	363 305	8.8	<u> </u>	9.1	5024	9.5	5024	9.83	5024	10.4	5409	10.9	5409	11.5	5409	11.8	6005	12.0	6005	12.25	6005	12.7	6678	13.1	6678	13.54	6678	13.9	7441	14.2	7441 1093	14.56	7441	14.9	_ <u>~</u>
4		9.15	v	9.3	772	9.4	772	9.46	772	9.9	840	10.3	840	10.66	840	11.1	909	11.5	909	11.97	909	12.4	1000	12.9	1000	13.32	1000	13.6	1093	13.9		14.21	1093	14.5	v
l ',	306	9.55	ŭ	10.0		10.4	772	10.85		11.1	840	11.3	840 840	11.54	840 840	11.9	909	12.3	909	12.67	909	13.0	1000	13.4	1000	13.74 13	1000	14.1 13.4	1093	14.5	1093	14.88	1093	15.3 14.4	V
',	316 320	9.4 8.67	0	9.6	772	9.9	772 772	10.08	772 772	10.3	840 840	10.4 10.7	840 840	10.6	840	11.1	909 909	11.5	909	11.98	909 909	12.3 12.7	1000 1000	12.7 13.2	1000	13,59	1000	13.4	1093	13.7 14.0	1093 1093	14.06	1093	14.4	0
'	347	9.7	,	10.0		10.4	772	10.71		10.3	840	11.4	840	11.68	840	12.0	909	12.4	909	12.31 12.71	909	13.2	1000	13.2	1000	14.21	1000	14.5	1093	14.8	1093	15.07	1093	15.4	ň
	301	10.17	<u> </u>	10.0		10.4	2405	10.53		10.7	2552	10.9	2552	11.08	2552	11.4	2819	11.8	2819	12.11	2819	12.4	3111	12.7	3111	13.03	3111	13.3	3239	13.5	3239	13.7	3239	13.9	
l å	318	10.12	ň	10.3		10.4	2405	10.37		10.7	2552	11.4	2552	11.85	2552	11.9	2819	12.0	2819	12.14	2819	12.7	3111	13.2	3111	13.8	3111	14.1	3239	14.5	3239	14.8	3239	15.1	ň
Ä	352	10.06	ň	10.2		10.3	2405	10.55		11.1	2552	11.6	2552	12.08	2552	13.3	2819	14.6	2819	15.84	2819	14.8	3111	13.2	3111	12.8	3111	13.6	3239	14.3	3239	15.11	3239	15.9	ň
l å	358	9.35	ñ	9.7	2405	10.4	2405	10.32		10.7	2552	11.0	2552	11.38	2552	11.5	2819	11.7	2819	11.84	2819	12.4	3111	13.0	3111	13.57	3111	13.9	3239	14.2	3239	14.51	3239	14.8	ň
l š	358	8.75	ň	9.1	2405	9.5	2405	9.94	2405	10.4	5453	10.8	2552	11.22	2552	11.6	2810	11.9	2819	12.21	2819	12.7	3111	13.3	3111	13.78	3111	13.9	3239	14.0	3239	14.15	3239	14.3	ň
 	309	11.25	- 0	11.5		11.8	5714	12.12		12.5	6107	12.8	6107	13.21	6107	13.5	6756	13.8	6756	14.1	6756	14.5	7312	14.9	7312	15.29	7312	15.9	7977	16.5	7977	17.04	7977	17.6	-
1 %	330	9.65	ň	10.0		10.4	5714	10.7	5714	11.3	6107	11.8	6107	12.35	6107	12.7	6756	13.0	6756	13,38	6756	14.0	7312	14.6	7312	15.28	7312	15.6	7977	16.0	7977	16.42	7977	18.8	ň
ĺ	351	11.6	ñ	11.6		11.5	5714	11.52		11.9	6107	12.3	6107	12.75	6107	13.1	6756	13.5	6756	13.84	6756	14.2	7312	14.6	7312	14.99	7312	15.4	7977	15.7	7977	16.1	7977	16.5	ñ
ű	353	9.04	ñ	9.5	5714	9.9	5714	10.27		10.6	6107	11.0	6107	11.29	6107	11.7	6756	12.2	6756	12.62	6756	12.9	7312	13.2	7312	13.49	7312	13.8	7977	14.2	7977	14.55	7977	14.9	ň
Å	359	10.6	ñ	10.9		11.2	5714	11.46		12.0	6107	12.5	6107	12.96	6107	13.4	6756	13.8	6756	14.18	6756	14.7	7312	15.2	7312	15.78	7312	16.0	7977	16.2	7977	16.41	7977	16.6	ň
10	325	9.09		9.5	0714	10.0	0	10.45		10.2	010/	10.0	V/V/	9.75	0107	10.4	0/30	11.1	0/30	11.77	0/30	12.3	0	12.9	7512	13.45	7312	13.8	-18/1	14.1	- '0'	14.45	- '8''	14.8	- ŏ
10	337	9.16	ň	9.8	õ	10.4	ŏ	10.96	ő	11.1	ŏ	11.2	ň	11.35	ŏ	11.7	ŏ	12.1	ŏ	12.42	Ď	13.3	·ŏ	14.1	ŏ	14.92	ŏ	15.1	ŏ	15.3	ň	15.52	ň	15.7	Ď
10	360	9.7	ŏ	9.8	õ	9.9	ŏ	10.03	Š	10.3	š	10.6	ŏ	10.86	ŏ	11.4	ŏ	12.0	ă	12.62	ő	12.9	. 0	13.2	ŏ	13.42	ŏ	13.9	ŏ	14.3	ŏ	14.71	ő	15.1	Ö
<u>v</u>	700	1 5.7		1 3.0	<u>~</u> _	, 5.0	<u>v</u> _	1 .0.00		1		,,,,,	<u> </u>	1.0.00	<u>~</u>		<u>-</u>				<u>`</u>		<u>`</u>				<u>`</u> _		<u> </u>	<u></u>	_ <u> </u>	14111	<u> </u>		

On 11/2/2000 (AM Dose), Pig #358 (group 8) received 100% of the Group 8 dose and 95% of the Group 9 dose

TABLE A-2
Body Weight Adjusted Doses
(Dose for Day/BW for Day)

Group	ID#	Day 0	Day 1	Day 2	Day 3	Day 4	Day 5	Day 6	Day 7	Day 8	Day 9	Day 10	Day 11	Day 12	Day 13	Day 14	Avg Dose Ta	arget Dose	% Target	Avg %
1	311	25.35	24.10	22.97	25.13	24.49	23.88	25.58	25.38	25.19	25.39	24.28	23.27	25.97	26.03	26.09	24.87	25	99	
1	326	26.52	25.10	23.83	26.20	25.65	25.12	26.73	26.36	26.00	26.15	24.97	23.88	26.47	26.36	26.24	25.71	25	103	
1 .	338	25.18	24.38	23.63	24.93	23.50	22.23	23.57	23.16	22.77	23.39	22.77	22.19	24.60	24.49	24.39	23.68	25	95	
1	343	29.60	28.78	28.01	30.98	30.50	30.04	31.49	30.60	29.76	30.06	28.80	27.64	30.27	29.79	29.32	29.71	25	119	
	349	26.39	25.20	24.10	26.72	26.38	26.04	27.16	26.28	25.46	25.95	25.09	24.28	26.65	26.29	25.94	25.86	25	103	
2	304	76.26	72.88	69.80	73.37	71.74	70.19	72.22	69.67	67.30	72.31	70.95	69.65	73.50	72.26	71.06	71.55	75	95	
2	307	82.26	81.05	79.88	84.90	83.89	82.91	84.31	80.45	76.93	81.42	78.76	76.26	81.38	80.86	80.36	81.04	75	108	
2	312	85.03	82.72	80.53	85.58	84.56	83.56	86.75	84.39	82.15 74.47	87.40	84.97 78.78	82.66 77.45	86.56	84.46	82.47	84.25	75	112	
2	314	81.19	79.80	78.45	81.48	78.76	76.22	78.96 74.72	76.65 72.86	74.47	80.15 75.44	78.78	70.99	80.97 75.18	78.87 74.15	76.89	78.61	75 75	105	
2	350	78.87	76.75 253.60	74.74	77.31 255.52	74.44 248.19	71.78		239.06			238.27	230.40			73.15 242.90	74.31 245.25	225	99 109	
3	302	260.72		246.86			241.27	248.19		230.57	246.69	238.27	203.75	250.06 225.63	246.43	228.04				
3	303	211.69	206.79	202.11	215.32	215.07	214.83	220.95	212.77	205.18	219.04				226.83		214.61	225	95	
3	322	246.86	238.60	230.87	234.50	223.84	214.10	223.84	218.91	214.19	228.92	220.88	213.39	232.29	229.58	226.93	226.51	225	101	
3 3	327	252.79	253.15	253.51	261.47	253.11	245.28	249.66	238.14	227.63	244.46	236.94 229.04	229.86	247.73	242.48 230.28	237.45 223.99	244.91	225	109	
	342	248.51	240.22	232.47	244.01 74.18	240.19	236.49 71.91	241,57 77,16	231.16 75.95	221.62 74.78	237.13 77.06	73.75	221.48 70.72	236.93 76.57	75.39	74.24	234.34 74.00	225 75	104 99	
4	313	75.49	71.61	68.11		73.03 77.64					78.83				79.74					
4	328	84.40	80.57 82.01	77.06 79.89	81.28 85.34	82.48	74.31 79.80	79.11 83.52	77.27 80.29	75.52 77.29	79.85	76.37 76.60	74.05 73.61	80.59 79.53	78.14	78.92 76.81	78.38 79.96	75 75	105	
4	329	84.25							81.23	79.09	82.69		77.88	83.40	81.25				107	
4	333	86.38	84.25	82.22	86.51	82.45	78.75	83.49				80.21				79.20	81.93	75	109	
	348	71.77 268.13	69.41	67.21 254.95	73.05 256.18	71.76 241.02	70.52 227.55	74,46 243,48	72.16 232.65	70.00 222.74	73.70 240.56	71.97 233.67	70.32 227.17	75.36 232.78	73.46 224.54	71.66 216.87	71.79 238.91	75 225	96	
5 5	317		261.37 249.28	235.76	247.50	242.43	237.56	254.98	244.33	234.54	254.85	248.99	243.39	256.93	255.02	253.13	248.21	225	106 110	
5 5	324 340	264.45 240.90	237.94	235.76	236.73	223.19	211.12	232.00	227.27	222.74	240.50	233.56	227.01	234.04	227.06	220.49	229.97	225	102	
5 5	341	250.45	246.37	242.42	251.42	243.46	235.98	246.35	230.28	216.17	244.99	249.59	254.36	261.27	252.61	244.50	244.68	225	102	
5 5	354	202.23	198.59	195.08	201.42	193.83	186.98	205.50	201.35	197.36	210.01	201.20	193.11	200.82	196.43	192.24	198.39	225	88	
6	321	571.99	554.12	537.33	553.63	530.81	509.80	537.92	512.52	489.41	533.53	523.22	513.30	561.44	551.32	541.56	534.79	500	107	103
6	332	497.26	497.10	496.93	526.51	518.27	510.28	536.16	508.90	484.27	514.88	493.21	473.28	521.20	515.19	509.31	506.85	500	101	
-			489.51	484.01	495.94	473.09	452.26	487.29	473.33	460.15	487.21	464.93	444.61	484.23	473.55	463.33		500	95	
6 6	335 355	495.14 601.92	565.55	533.33	555.34	537.67	521.10	553.12	529.85	508.47	536.39	510.16	486.38	539.33	536.74	534.17	475.24 536.63	500	107	
6	363	549.47	529.59	511.09	520.76	494.27	470.35	511.06	500.42	490.20	526.66	509.38	493.21	536.10	523.28	511.06	511.79	500	107	103
7	305	83.43	82.51	81.61	85.24	81.92	78.84	81.94	78.84	75.96	80.48	77.67	75.05	80.26	78.55	76.91	79.95	75	107	103
7	306	77.33	74.12	71.16	75.85	74.31	72.83	76.30	73.97	71.77	76.73	74.69	72.75	77.40	75.37	73.45	74.54	75	99	
7	316	80.20	78.35	76.59	81.97	80.61	79.29	82.22	78.93	75.90	81.14	78.96	76.89	81.84	79.73	77.73	79.36	75	106	
7	320	84.93	81.18	77.75	81.33	78.28	75.44	78.86	76.28	73.87	78.48	75.94	73.55	79.23	78.08	76.96	78.01	75	104	
7	347	76.92	74.43	72.09	76.17	74.00	71.96	75.63	73.53	71.54	75.67	72.91	70.34	75.39	73.93	72.52	73.80	75	98	
8	301	233.75	231.05	228.42	237.98	233.77	229.70	246.33	239.35	232.77	250.57	244.53	238.78	244.40	240.35	236.43	237.88	225	106	
8	318	235.73	233.82	231.94	234,91	224.71	215.35	235.95	234.05	232.19	245.11	234.87	225.46	229.18	223.90	218.86	230.40	225	102	
8	352	235.27	231.57	227.99	230.74	220,57	211.25	211.41	193.24	177.95	209.84	225.24	243.07	238.70	225.88	214.37	219.81	225	98	
8	356	248.65	240.61	233.07	239.10	231.43	224.25	244.40	241.20	238.07	250.57	239.45	229.28	233.31	228.16	223.23	236.32	225	105	
8	358	262.96	252.03	241.98	525.99	236,44	227.45	244.05	237.27	230.86	244.34	234.70	225.78	232.97	230.92	228.91	257.11	225	114	
9	309	495.15	483.01	471.45	489.21	475.38	462.30	500.20	489.45	479.15	504.39	490.96	478.22	502.54	484.73	468.13	484.95	500	97	
9	330	571.40	552.08	534.02	542.84	517.54	494.49	532.25	518.23	504.93	522.53	500.59	480.42	510.69	497.94	485.81	517.72	500	104	
9	351	493.72	494.86	496.01	511.90	494.89	478.98	515.20	501.31	488.15	514.08	500.59	487.79	519.34	507.12	495.47	499.96	500	100	
9	353	604.66	579.51	556.38	575.59	557.72	540.92	575.80	554.83	535.34	566.38	553.94	542.03	576.23	561.89	548.25	561.96	500	112	
9	359	524.86	511.40	498.60	510.62	490.13	471.22	505.44	490.51	476.45	496.96	479.58	463.37	498.87	492.41	486.11	493.10	500	99	102
10	325	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0		
10	337	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0		
10	360	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0.00	0		

TABLE A - 3 RAW AND ADJUSTED BLOOD LEAD DATA

pig number	sample	group	material administered	dosage	qualifier	result	day	MATRIX	Adjusted Value ()	Notes
311	VB-03-0173	1	PbAc .	25	~	1.0	0	Blood	0.5	
326	VB-03-0221	1	PbAc	25		2.0	Ð.	Blood	2.0	
338	VB-03-0214	1	PbAc .	25	<	1.0	0	Blood	0.5	
343	VB-03-0212	1	PbAc	25	<	1.0	0	Blood	0.5	
349	VB-03-0205 VB-03-0217	1	PhAc	25 75	< <	1.0	0	Blood	0.5	
304 307	VB-03-0223	2 2	PbAc PbAc	75	~	1.0 1.0	0	Blood Blood	0.5 0.5	
312	VB-03-0194	2	PbAc	75	~	1.0	ő	Blood	0.5	
314	VB-03-0188	2	PbAc	75	<	1.0	ŏ	Blood	0.5	
350	VB-03-0196	. 2	PbAc	75	<	1.0	ŏ	Blood	0.5	
302	VB-03-0186	3	PbAc	225	<	1.0	0	Blood	0.5	
303	VB-03-0203	3	PbAc	225	<	1.0	0	Blood	0.5	
322	VB-03-0219	3	PbAc	225	<	1.0	0.	Blood	0.5	
327	VB-03-0198	3	PbAc	225	<	1.0	0	Blood	0.5	
342	VB-03-0197	3	PbAc	225	<	1.0	0	Blood	0.5	
313	VB-03-0180	4	Site 1	75 75	<	1.0	0	Blood	0.5	
328 329	VB-03-0183 VB-03-0213	4	Site 1 Site 1	75 75	< <	1.0	0	Blood Blood	0.5	
333	VB-03-0195	. 4	Site 1	75 75	<	1.0 1.0	ő	Blood	0.5 0.5	
348	VB-03-0199	4	Site 1	75		1.0	ŏ	Blood	0.5	
317	VB-03-0175	5	Site 1	225	· <	1.0	ŏ	Blood	0.5	
324	VB-03-0182	5	Site 1	225	< ·	1.0	ō	Blood	0.5	
340	VB-03-0176	5	Site 1	225	<	1.0	0	Blood	0.5	
341	VB-03-0200	5	Site 1	225	<	1.0	0	Blood	0.5	•
354	VB-03-0206	5	Site 1	225	<	1.0	0	Blood	0.5	
321	VB-03-0174	6	Site 1	500	<	1.0	0	Blood	0.5	
332	VB-03-0216	6	Site 1	500		8.1	0	Blood	8.1	
335	VB-03-0207	6	Site 1	500		1.0	0	Blood	0.5	
355	VB-03-0187	6	Site 1	500	<	1.0	0	Blood	0.5	
363	VB-03-0208	6 7	Site 1	500	< <	1.0	0	Blood	0.5	
305 306	VB-03-0201 VB-03-0220	7	Site 2 Site 2	75 75	~	1.0 1.0	Ö	Blood Blood	0.5 0.5	
316	VB-03-0210	7	Site 2	75 75	<	1.0	Ö	Blood	0.5	•
320	VB-03-0209	7	Site 2	75	<	1.0	ŏ	Blood	0.5	
347	VB-03-0218	7	Site 2	75	<	1.0	ŏ	Blood	0.5	
301	VB-03-0190	8	Site 2	225	<	1.0	ō	Blood	0.5	
318	VB-03-0189	8	Site 2	225	<	1.0	0	Blood	0.5	
352	VB-03-0224	8	Site 2	225	<	1.0	0	Blood	0.5	
356	VB-03-0204	8	Site 2	225	<	1.0	0	Blood	0.5	
358	VB-03-0193	8	Site 2	225	<	1.0	0	Blood	0.5	
309	VB-03-0185	9	Site 2	500	<	1.0	0	Blood	0.5	
330	VB-03-0181	9	Site 2	500	<	1.0	0	Blood	0.5	
351 353	VB-03-0179 VB-03-0215	9 9	Site 2 Site 2	500 500	< ·	1.0	0	Blood	0.5	
359	VB-03-0215 VB-03-0177	9	Site 2	500	` `	1.0 1.0	0	Blood Blood	0.5 0.5	
325	VB-03-0184	10	Control	0	ζ.	1.0	ŏ	Blood	0.5	
337	VB-03-0225	10	Control	Ŏ	<	1.0	ŏ	Blood	0.5	
360	VB-03-0192	10	Control	Ō	<	1.0	ō	Blood	0.5	
311	VB-03-0244	1	PbAc	25	<u> </u>	1.0	1	Blood	0.5	
326	VB-03-0228	1	PbAc	25	<	1.0	1	Blood	0.5	
338	VB-03-0237	1	PbAc	25	<	1.0	1	Blood	0.5	
343	VB-03-0257	1	PbAc	25	<	1.0	1	Blood	0.5	
349 304	VB-03-0269 VB-03-0277	1 2	PbAc PbAc	25 75	<	1.0 2.0	1	Blood Blood	0.5 2.0	
307	VB-03-0247	2	PbAc	75		2.0	i	Blood	2.0	
312	VB-03-0253	2	PbAc	75		1.0	i	Blood	1.0	
314	VB-03-0272	2	PbAc	75		2.0	1	Blood	2.0	
350	VB-03-0240	2	PbAc	75		2.0	1	Blood	2.0	
302	VB-03-0230	3	PbAc	225		2.0	1	Blood	2.0	
303	VB-03-0260	3	PbAc	225		2.0	1	Blood	2.0	
322	VB-03-0258	3	PbAc	225		4.7	1	Blood	4.7	
327 342	VB-03-0243 VB-03-0266	3	PbAc PbAc	225 225		3.0	1	Blood	3.0	
313	VB-03-0262	3 4	Site 1	75	<	1.0 1.0	į	Blood Blood	1.0 0.5	
328	VB-03-0242	4	Site 1	75	•	1.0	· 1	Blood	1.0	
329	VB-03-0278	4	Site 1	75	<	1.0	i	Blood	0.5	
333	VB-03-0229	4	Site 1	75		2.0	i	Blood	2.0	
348	VB-03-0254	4	Site 1	75		1.0	1	Blood	1.0	
317	VB-03-0232	5	Site 1	225		3.0	1	Blood	3.0	
324	VB-03-0239	5	Site 1	225		3.6	1	Blood	3.6	
340	VB-03-0263	5	Site 1	225		2.0	1	Blood	2.0	
341	VB-03-0234	5	Site 1	225		5.2	1	Blood	5.2	
354	VB-03-0274	5	Site 1	225		2.0	1	Blood	2.0	·
321 332	VB-03-0265 VB-03-0236	6 6	Site 1	500 500	>~ .	4.0	1	Blood	4.0	
335	VB-03-0235	6	Site 1	500		3.1 3.2	i	Blood Blood	3.1 3.2	
355	VB-03-0256		Site 1	500		6.9	1	Blood	6.9	
363	VB-03-0264		Site 1	500		4.0	i	Blood	4.0	
305	VB-03-0255		Site 2	75	<	1.0	i	Blood	0.5	
306	VB-03-0273	7	Site 2	75		1.0	i	Blood	1.0	
316	VB-03-0259	7	Site 2	75		1.0	1	Blood	1.0	
320	VB-03-0261	7	Site 2	75		1.0	1	Blood	1.0	
347	VB-03-0249		Site 2	75		1.0	1	Blood	1.0	
301	VB-03-0268		Site 2	225		3.0	1	Blood	3.0	
318	VB-03-0241		Site 2	225		2.0	1	Blood	2.0	
352 356	VB-03-0227 VB-03-0245	8 8	Site 2 Site 2	225 225		3.9	1	Blood	3.9	
356 358	VB-03-0245 VB-03-0251		Site 2	225		2.0 3.5	1	Blood Blood	2.0 3.5	
309	VB-03-0251 VB-03-0276		Site 2	500		3.5 4.5	1	Blood	3.5 4.5	
330	VB-03-0250		Site 2	500		5.7	i	Blood	5.7	
351	VB-03-0233		Site 2	500		2.0	1	Blood	2.0	

pig number	sample	group	material administered	dosage qualifier	result	day MATRIX	Adjusted Value ()*	Notes
353	VB-03-0246	9	Site 2	500	4.1	1 Blood	4,1	
359	VB-03-0271	9	Site 2	500	3.7	1 Blood	3.7	
325 337	VB-03-0252 VB-03-0248	10 10	Control Control	0 < 0 <	1.0 1.0	1 Blood 1 Blood	0.5 0.5	
360	VB-03-0246 VB-03-0267	10	Control	. 0	4.0	1 Blood	4.0	
311	VB-03-0289	1	PbAc	25 <	1.0	2 Blood	0.5	
326	VB-03-0313	1	PbAc	25 <	1.0	2 Blood	0.5	
338 343	VB-03-0311 VB-03-0319	1	PbAc PbAc	25 < 25 <	1.0 1.0	2 Blood 2 Blood	0.5 0.5	
343 349	VB-03-0319 VB-03-0302	1	PbAc	25 ~ 25	17.0	2 Blood	17.0	
304	VB-03-0295	2	PbAc	75	2.0	2 Blood	2.0	
307	VB-03-0323	2	PbAc	75	2.0	2 Blood	2.0	
312	VB-03-0306	2	PbAc	75 	3.0	2 Blood	3.0	
314	VB-03-0304 VB-03-0280	2	PbAc PbAc	75 75	2.0 2.0	2 Blood 2 Blood	2.0 2.0	
350 302	VB-03-0288	3	PbAc	225	4.0	2 Blood	4.0	
303	VB-03-0305	3	PbAc	225	4.9	2 Blood	4.9	
322	VB-03-0309	3	PhAc	225	6.3	2 Blood	6.3	
327	VB-03-0307	3	PbAc	225	4.2	2 Blood	4.2	
342 313	VB-03-0330 VB-03-0301	3 4	PbAc Site 1	225 75	1.0 1.0	2 Blood 2 Blood	1.0 1.0	
328	VB-03-0283	4	Site 1	75	1.0	2 Blood	1.0	
329	VB-03-0298	4	Site 1	75	1.0	2 Blood	1.0	
333	VB-03-0326	4	Site 1	75	2.0	2 Blood	2.0	
348	VB-03-0320	4	Site 1	75 205	1.0	2 Blood	1.0	
317 324	VB-03-0279 VB-03-0285	5 5	Site 1 Site 1	225 225	3.6 4.9	2 Blood 2 Blood	3.6 4.9	
340	VB-03-0324	5	Site 1	225	3.1	2 Blood	4.5 3.1	
341	VB-03-0312	5	Site 1	225	4.7	2 Blood	4.7	
354	VB-03-0292	5	Site 1	225	3.0	2 Blood	3.0	
321	VB-03-0325	6	Site 1	500	5.7	2 Blood	5.7	
332 335	VB-03-0314 VB-03-0322	6 6	Site 1 Site 1	500 500	4.6 3.8	2 Blood . 2 Blood	4.6 3.8	
355	VB-03-0286	6	Site 1	500	7.8	2 Blood	7.8	
363	VB-03-0310	6	Site 1	500	5.2	2 Blood	5.2	
305	VB-03-0300	7	Site 2	75	2.0	2 Blood	2.0	
306	VB-03-0331	7 7	Site 2	75 75	29.0 2.0	2 Blood 2 Blood	29.0 2.0	
316 320	VB-03-0294 VB-03-0293	7	Site 2 Site 2	75 75	2.0	2 Blood 2 Blood	2.0	
347	VB-03-0318	7	Site 2	75 <	1.0	2 Blood	0.5	
301	VB-03-0299	8	Site 2	225	3.0	2 Blood	3.0	
318	VB-03-0321	8	Site 2	225	2.0	2 Blood	2.0	
352	VB-03-0281	8	Site 2	225 225	3.7	2 Blood 2 Blood	3.7	
356 358	VB-03-0282 VB-03-0317	8 8	Site 2 Site 2	225	2.0 5.0	2 Blood 2 Blood	2.0 5.0	
309	VB-03-0287	. 9	Site 2	500	6.7	2 Blood	6.7	
330	VB-03-0303	9	Site 2	500	6.3	2 Blood	6.3	
351	VB-03-0327	9	Site 2	500	3.9	2 Blood	3.9	
353	VB-03-0296 VB-03-0315	9	Site 2 Site 2	500 500	5.9 4.4	2 Blood 2 Blood	5.9 4.4	
359 325	VB-03-0316	10	Control	0 <	1.0	2 Blood	0.5	
337	VB-03-0308	10	Control	0 <	1.0	2 Blood	0.5	
360	VB-03-0290	10	Control	0	2.0	2 Blood	2.0	
311	VB-03-0356	1	PbAc	25 < 25	1.0 2.8	3 Blood 3 Blood	0.5 2.8	
326 338	VB-03-0337 VB-03-0342	1	PbAc PbAc	25 <	1.0	3 Blood	0.5	
343	VB-03-0371	1	PbAc	25	0.9	3 Blood	0.9	
349	VB-03-0380	. 1	PbAc	25	1.0	3 Blood	1.0	
304	VB-03-0381	2	PbAc	75	2.6	3 Blood	2.6	
307	VB-03-0361 VB-03-0370	2 2	PbAc PbAc	75 75	3.2 3.8	3 Blood 3 Blood	3.2 3.8	
312 314	VB-03-0343	2	PbAc	75 75	2.0	3 Blood	2.0	
350	VB-03-0357	2	PbAc	75	2.4	3 Blood	2.4	
302	VB-03-0339	3	PbAc	225	4.5	3 Blood	4.5	
303	VB-03-0369	3 3	PhAc	225 225	6.7 6.4	3 Blood 3 Blood	6.7 6.4	
322 327	VB-03-0352 VB-03-0374	3	PbAc PbAc	225	5.0	3 Blood	5.0	
342	VB-03-0349	3	PbAc	225	4.0	3 Blood	4.0	
313	VB-03-0340	4	Site 1	75	1.6	3 Blood	1.6	
328	VB-03-0363	4	Site 1	75 75	1.3	3 Blood	1.3	•
329 333	VB-03-0344 VB-03-0338	4	Site 1 Site 1	75 75	4.7 6.7	3 Blood 3 Blood	4.7 6.7	
333 348	VB-03-0338 VB-03-0379	4	Site 1	75 75	2.2	3 Blood	2.2	
317	VB-03-0378	5	Site 1	225	3.6	3 Blood	3.6	
324	VB-03-0372	5	Site 1	225	5.2	3 Blood	5.2	•
340	VB-03-0351	5	Site 1	225	4.1	3 Blood	4.1	
341 354	VB-03-0353 VB-03-0341	5 5	Site 1 Site 1	225 225	4.7 3.3	3 Blood 3 Blood	4.7 3.3	
354 321	VB-03-0368	6	Site 1	500	7.7	3 Blood	7.7	
332	VB-03-0355	6	Site 1	500	3.8	3 Blood	3.8	
335	VB-03-0364	6	Site 1	500	3.7	3 Blood	3.7	
355	VB-03-0367	6	Site 1	500	10.1	3 Blood	10.1	
363 305	VB-03-0350 VB-03-0336	6 7	Site 1 Site 2	500 75	6.4 1.5	3 Blood 3 Blood	6.4 1.5	
305 306	VB-03-0336 VB-03-0332	7	Site 2	75 75	2.6	3 Blood	2.6	
316	VB-03-0333	7	Site 2	75	2.5	3 Blood	2.5	
320	VB-03-0348	7	Site 2	75	1.3	3 Blood	1.3	
347	VB-03-0347	7	Site 2	75 <	1.0	3 Blood	0.5	
301	VB-03-0354	8	Site 2	225 225	3.8 3.5	3 Blood 3 Blood	3.8 3.5	
318 352	VB-03-0376 VB-03-0362	8 8	Site 2 Site 2	225 225	3.5 3.8	3 Blood 3 Blood	3.5 3.8	
352 356	VB-03-0366	8	Site 2	225	3.1	3 Blood	3.1	
358	VB-03-0345	8	Site 2	225	4.6	3 Blood	4.6	
309	VB-03-0360	9	Site 2	500	7.1	3 Blood	7.1	

.

							•	
pig number	sample	group material administered	dosage qualifier	result	day MATRIX	Adjusted Value ()2	Notes	
330	VB-03-0335	9 Site 2	500	7.3	3 Blood	7.3	•	
351	VB-03-0358	9 Site 2	500	4.6	3 Blood	4.6		
353 359	VB-03-0359 VB-03-0365	9 Site 2 9 Site 2	500 500	11.0 5.6	3 Blood 3 Blood	11.0 5.6	•	
325	VB-03-0377	10 Control	0 <	1.0	3 Blood	0.5		•
337	VB-03-0375	10 Control	0 <	1.0	3 Blood	0.5		
360 311	VB-03-0383	10 Control	0 < 25 <	1.0	3 Blood 5 Blood	0.5 0.5		
326	VB-03-0406 VB-03-0388	1 PbAc 1 PbAc	25 \	1.3	5 Blood 5 Blood	1.3		
338	VB-03-0400	1 PbAc	25 <	1.0	5 Blood	0.5	*	
343	VB-03-0391	1 PbAc	25	1.4	5 Blood	1.4		
349 304	VB-03-0428 VB-03-0424	1 PbAc 2 PbAc	25 75	1.2 2.8	5 Blood 5 Blood	1.2 2.8		
307	VB-03-0390	2 PbAc	75	1.9	5 Blood	1.9		
312	VB-03-0437	2 PbAc	75	4.5	5 Blood	4.5		
314	VB-03-0416	2 PbAc	75 75	2.0	5 Blood	2.0		
350 302	VB-03-0385 VB-03-0417	2 PbAc 3 PbAc	75 225	2.8 7.1	5 Blood 5 Blood	2.8 7.1		
303	VB-03-0399	3 PbAc	225	6.3	5 Blood	6.3		
322	VB-03-0407	3 PbAc	225	7.3	5 Blood	7.3		
327 342	VB-03-0420 VB-03-0413	3 PbAc 3 PbAc	225 225	6.3 3.6	5 Blood 5 Blood	6.3 3.6		
313	VB-03-0395	4 Site 1	75	1.7	5 Blood	1.7		
328	VB-03-0401	4 Site 1	75	1.0	5 Blood	1.0		
329	VB-03-0412	4 Site 1	75 76	2.6	5 Blood	2.6		•
333 348	VB-03-0415 VB-03-0436	4 Site 1 4 Site 1	75 75	2.9 2.2	5 Blood 5 Blood	2.9 2.2		
317	VB-03-0403	5 Site 1	225	4.6	5 Blood	4.6		
324	VB-03-0411	5 Site 1	225	4.9	5 Blood	4.9		
340 341	VB-03-0394 VB-03-0410	5 Site 1 5 Site 1	225 225	3.5 6.5	5 Blood 5 Blood	3.5 6.5		
354	VB-03-0434	5 Site 1	225	5.4	5 Blood	5.4		
321	VB-03-0392	6 Site 1	500	12.0	5 Blood	12.0		•
332	VB-03-0409	6 Site 1	500	4.7	5 Blood	4.7		
335 355	VB-03-0418 VB-03-0405	6 Site 1 6 Site 1	500 500	5.1 9.2	5 Blood 5 Blood	5.1 9.2		
363	VB-03-0404	6 Site 1	500	6.8	5 Blood	6.8		
305	VB-03-0398	7 Site 2	75	2.6	5 Blood	2.6		
306	VB-03-0422	7 Site 2 7 Site 2	75 75	1.7	5 Blood	1.7		
316 320	VB-03-0389 VB-03-0408	7 Site 2 7 Site 2	75 75	2.6 1.2	5 Blood 5 Blood	2.6 1.2		
347	VB-03-0397	7 Site 2	75	1.1	5 Blood	1.1		
301	VB-03-0414	8 Site 2	225	5.2	5 Blood	5.2		
318 352	VB-03-0431 VB-03-0430	8 Site 2 8 Site 2	225 225	5.4 6.7	5 Blood 5 Blood	5.4 6.7		
356	VB-03-0425	8 Site 2	225	3.7	5 Blood	3.7		
358	VB-03-0393	8 Site 2	225	5.8	5 Blood	5.8		
309	VB-03-0387	9 Site 2	500	7.7	5 Blood	7.7		
330 351	VB-03-0433 VB-03-0423	9 Site 2 9 Site 2	500 500	7.9 5.9	5 Blood 5 Blood	7.9 5.9		
353	VB-03-0396	9 Site 2	500	6.6	5 Blood	6.6		
359	VB-03-0402	9 Site 2	500	7.4	5 Blood	7.4		
325 337	VB-03-0386 VB-03-0419	10 Control 10 Control	0 < 0 <	1.0 1.0	5 Blood 5 Blood	0.5 0.5		
360	VB-03-0426	10 Control	0 <	1.0	5 Blood	0.5		*
311	VB-03-0484	1 PbAc	25 <	1.0	7 Blood	0.5		
326 338	VB-03-0460 VB-03-0451	1 PbAc 1 PbAc	25 25	1.0 1.3	7 Blood 7 Blood	1.0		•
343	VB-03-0448	1 PbAc	25 <	1.0	7 Blood	1.3 0.5		
349	VB-03-0439	1 PbAc	25	1.9	7 Blood	1.9		
304 307	VB-03-0465 VB-03-0447	2 PbAc 2 PbAc	75 75	3.8 1.7	7 Blood 7 Blood	3.8 1.7		
312	VB-03-0467	2 PbAc	75 75	5.2	7 Blood	5.2		
314	VB-03-0446	2 PbAc	75	2.7	7 Blood	2.7		
350	VB-03-0478	2 PbAc	75	2.4	7 Blood	2.4		
302 303	VB-03-0472 VB-03-0462	3 PbAc 3 PbAc	225 225	6.9 6.3	7 Blood 7 Blood	6.9 6.3		
322	VB-03-0441	3 PbAc	225	8.1	7 Blood	8.1		
327	VB-03-0468	3 PbAc	225	6.8	7 Blood	6.8		
342 313	VB-03-0469 VB-03-0485	3 PbAc 4 Site 1	225 75	7.9 2.2	7 Blood 7 Blood	7.9 2.2		
328	VB-03-0471	4 Site 1	75	1.8	7 Blood	1.8		
329	VB-03-0487	4 Site 1	75	3.5	7 Blood	3.5		
333 348	VB-03-0457 VB-03-0480	4 Site 1 4 Site 1	75 75	2.9 3.0	7 Blood 7 Blood	2.9 3.0		
346 317	VB-03-0456	5 Site 1	75 225	5.7	7 Blood	3.0 5.7		
324	VB-03-0476	5 Site 1	225	7.0	7 Blood	7.0		
340	VB-03-0450	5 Site 1	225	4.5	7 Blood	4.5		
341 354	VB-03-0466 VB-03-0461	5 Site 1 5 Site 1	225 225	9.6 7.8	7 Blood 7 Blood	9.6 7.8		
321	V8-03-0473	6 Site 1	500	8.2	7 Blood	8.2		
332	VB-03-0490	6 Site 1	500	7.0	7 Blood	7.0		
335 355	VB-03-0442 VB-03-0443	6 Site 1 6 Site 1	500 500	6.6 9.8	7 Blood 7 Blood	6.6 9.8		
363	VB-03-0486	6 Site 1	500	6.5	7 Blood	9.8 6.5		
305	VB-03-0489	7 Site 2	75	3.3	7 Blood	3.3		
306	VB-03-0459	7 Site 2	75	3.7	7 Blood	3.7		
316 320	VB-03-0440 VB-03-0455	7 Site 2 7 Site 2	75 75	2.8 2.5	7 Blood 7 Blood	2.8 2.5		
347	VB-03-0495 VB-03-0481	7 Site 2	75 75	2.5 1.5	7 Blood	2.5 1.5		
301	VB-03-0454	8 Site 2	225	8.0	7 Blood	8.0		
318	VB-03-0458	8 Site 2	225	6.3	7 Blood	6.3		
352 356	VB-03-0477 VB-03-0453	8 Site 2 8 Site 2	225 225	7.4 3.6	7 Blood 7 Blood	7.4 3.6		
		-			. 2.500			

pig number	sample	group	material administered	dosage qualifier	result	day	MATRIX	Adjusted Value ()*	Notes
358	VB-03-0470	8	Site 2	225	7.4	7	Blood	7.4	
309	VB-03-0475	9	Site 2	500	8.2	7	Blood	8.2	
330	VB-03-0482	. 9	Site 2	500	9.7	7	Blood	9.7	
351	VB-03-0474	9	Site 2	500	6.5	7	Blood	6.5	
353	VB-03-0445	9	Site 2	500	8.5	7	Blood	8.5	
359 325	VB-03-0438 VB-03-0463	9 10	Site 2 Control	500 0 <	9.5 1.0	7 7	Blood Blood	9.5 0.5	
337	VB-03-0464	10	Control	0 <	1.0	7	Blood	0.5	
360	VB-03-0479	10	Control	0 <	1.0	7	Blood	0.5	
311	VB-03-0518	1	PbAc	25	1.9	9	Blood	1.9	
326	VB-03-0501	1	PbAc	25	. 2.0	9	Blood	2.0	
338	VB-03-0511	1	PbAc	25	2.2	9	Blood	2.2	
343	VB-03-0521	1	PbAc	25	2.2	9	Blood	2.2	
349 304	VB-03-0529 VB-03-0509	1 2	PbAc PbAc	25 75	2.4 4.3	9	Blood Blood	2.4 4.3	
307	VB-03-0535	2	PbAc	75 75	3.7	9	Blood	4.3 3.7	
312	VB-03-0531	2	PbAc	75	5.4	9	Blood	5.4	
314	VB-03-0499	2	PbAc	75	3.5	9	Blood	3.5	
350	VB-03-0502	2	PbAc	75	3.5	9	Blood	3.5	
302	VB-03-0495	3	PbAc	225	7.8	9	Blood	7.8	
303	VB-03-0512	3	PbAc	225	7.6	9	Blood	7.6	
322	VB-03-0514	3 3	PbAc	225	9.9 7.2	9	Blood Blood	9.9 7.2	
327 342	VB-03-0540 VB-03-0526	3	PbAc PbAc	225 225	9.4	9	Blood	9.4	
313	VB-03-0510	4	Site 1	75	2.9	9	Blood	2.9	•
328	VB-03-0494	4	Site 1	75	2.6	9	Blood	2.6	
329	VB-03-0500	4	Site 1	75	4.0	9	Blood	4.0	
333	VB-03-0528	4	Site 1	75	3.8	9	Blood	3.8	
348	VB-03-0534	4	Site 1	75	5.3	9	Blood	5.3	
317	VB-03-0496	5	Site 1	225	7.3	9	Blood	7.3	
324 340	VB-03-0530	5	Site 1	225	8.0	9	Blood	8.0 5.1	
340 341	VB-03-0538 VB-03-0492	5 5	Site 1 Site 1	225 225	5.1 10.5	9 -	Blood Blood	5.1 10.5	
354	VB-03-0522	5	Site 1	225	10.5	9	Blood	10.1	
321	VB-03-0541	ě	Site 1	500	11.3	9	Blood	11.3	
332	VB-03-0523	6	Site 1	500	10.0	9	Blood	10.0	
335	VB-03-0515	6	Site 1	500	8.6	9	Blood	8.6	
355	VB-03-0537	6	Site 1	500	11.7	9	Blood	11.7	
363 305	VB-03-0505 VB-03-0507	6 7	Site 1 Site 2	500 75	9.5 4.4	9	Blood Blood	9.5 4.4	
306	VB-03-0520	7	Site 2	75 75	4.4	9	Blood	4.4	
316	VB-03-0539	7	Site 2	75	3.1	9	Blood	3.1	
320	VB-03-0543	7	Site 2	75	2.8	9	Blood	2.8	
347	VB-03-0542	7	Site 2	75	1.7	9	Blood	1.7	
301	VB-03-0498	8	Site 2	225	6.2	9	Blood	6.2	
318	VB-03-0504	8	Site 2	225	7.3	9	Blood	7.3	
352 356	VB-03-0491 VB-03-0503	8 8	Site 2 Site 2	225 225	11.4 4.7	9	Blood Blood	11.4 4.7	
358	VB-03-0532	8	Site 2	225	9.2	9	Blood	9.2	
309	VB-03-0493	9	Site 2	500	11.3	9	Blood	11.3	
330	VB-03-0536	9	Site 2	500	10.4	9	Blood	10.4	
351	VB-03-0513	9	Site 2	500	9.4	9	Blood	9.4	
353	VB-03-0497	9	Site 2	500	10.9	9	Blood	10.9	
359	VB-03-0527	9	Site 2	500	11.4	9	Blood	11.4	
325 337	VB-03-0506 VB-03-0525	10 10	Control Control	0 < 0 <	1.0 1.0	9	Blood Blood	0.5 0.5	
360	VB-03-0519	10	Control	0 <	1.0	9	Blood	0.5	
311	VB-03-0590	1	PbAc	25	2.0	12	Blood	2.0	
326	VB-03-0552	1	PbAc	25	2.9	12	Blood	2.9	
338	VB-03-0545	1	PbAc	25	3.5	12	Blood	3.5	
343	VB-03-0584	1	PbAc	25	2.2	12	Blood	2.2	•
349	VB-03-0589	1	PbAc	25	2.9	12	Blood	2.9	
304 307	VB-03-0564	2 2	PhAc	75 75	5.5 4.7	12 12	Blood Blood	5.5 4.7	
307 312	VB-03-0555 VB-03-0549	2	PbAc PbAc	75 75	5.9	12	Blood	4.7 5.9	
314	VB-03-0559	2	PbAc	75	3.2	12	Blood	3.2	
350	VB-03-0585	2	PbAc	75	4.8	12	Blood	4.8	
302	VB-03-0591	3	PbAc	225	9.7	12	Blood	9.7	
303	VB-03-0550	3	PbAc	225	9.1	12	Blood	9.1	
322	VB-03-0568	3	PbAc	225	11.1	12 12	Blood Blood	11.1 9.4	
327 342	VB-03-0556 VB-03-0573	3 3	PbAc PbAc	225 225	9.4 12.0	12	Blood	9.4 12.0 ^	
342 313	VB-03-0566	4	Site 1	75	3.8	12	Blood	3.8	
328	VB-03-0572	4	Site 1	75	9.9	12	Blood	9.9	
329	VB-03-0582	4	Site 1	75	4.4	12	Blood	4.4	
333	VB-03-0596	4	Site 1	75	5.0	12	Blood	5.0	
348	VB-03-0593	4	Site 1	75	4.7	12	Blood	4.7	
317	VB-03-0579	5	Site 1	225	8.8	12	Blood	8.8 9.6	
324 340	VB-03-0558 VB-03-0586	5 5	Site 1 Site 1	225 225	9.6 6.2	12 12	Blood Blood	9.6 6.2	
340 341	VB-03-0557	5	Site 1	225	9.7	12	Blood	9.7	
354	VB-03-0595	5	Site 1	225	10.2	12	Blood	10.2	
321	VB-03-0588	6	Site 1	500	17.0	12	Blood	17.0	
332	VB-03-0594	6	Site 1	500	11.4	12	Blood	11.4	
335	VB-03-0569	6	Site 1	500	9.9	12	Blood	9.9	
355	VB-03-0561	6	Site 1	500	13.6	12	Blood	13.6	
363	VB-03-0563	6	Site 1	500	10.0	12	Blood Blood	10.0 4.5	
305 306	VB-03-0548 VB-03-0576	7 7	Site 2	75 75	4.5 4.6	12 12	Blood	4.5 4.6	
306 316	VB-03-05/6 VB-03-0562	7	Site 2	75 75	4.0	12	Blood	4.0	
320	VB-03-0581	7	Site 2	75 75	5.2	12	Blood	5.2	
		7	Site 2	75	3.3	12	Blood	3.3	
347	VB-03-0587	r							
	VB-03-0575 VB-03-0551	8 8	Site 2 Site 2	225 225	7.1 9.0	12 12	Blood	7.1 9.0	

pig number	sample	group	material administered	dosage	qualifier	result	day	MATRIX	Adjusted Value ()	Notes
352	VB-03-0592	8	Site 2	225		12.0	12	Blood	12.0	
356	VB-03-0554	8	Site 2	225		6.4	12	Blood	6.4	
358	VB-03-0547	8	Site 2	225		8.9	12	Blood	8.9	
309	VB-03-0571	9,	Site 2	500		12.2	12	Blood	12.2	
330	VB-03-0570	9	Site 2	500		12.0	12	Blood	12.0	
351	VB-03-0553	9	Site 2	500		10.2	. 12	Blood	10.2	
353	VB-03-0565	9	Site 2	500		11.1	12	Blood	11.1	
359	VB-03-0574	9	Site 2	500		13.3	12	Blood	13.3	
325	VB-03-0560	10	Control	0		1.5	12	Blood	1.5	
337	VB-03-0577	10	Control	0	<	1.0	12	Blood	0.5	
360	VB-03-0544	10	Control	0	<	1.0	12	Blood	0.5	
311	VB-03-0635	1	PbAc	25		2.0	15	Blood	2.0	
326	VB-03-0616	1	PbAc	25		3.2	15	Blood	3.2	
338	VB-03-0649	1	PbAc	25		2.8	15	Blood	2.8	
343	VB-03-0622	1	PbAc	25		2.5	15	Blood	2.5	
349	VB-03-0605	1	PbAc	25		3.1	15	Blood	3.1	
304	VB-03-0618	2	PbAc	75		6.3	15	Blood	6.3	
307	VB-03-0610	2	PbAc	75		4.4	15	Blood	4.4	
312	VB-03-0625	2	PbAc	75		4.9	15	Blood	4.9	
314	VB-03-0601	2	PbAc	75		3.6	15	Blood	3.6	
350	VB-03-0608	2	PbAc	75		4.5	15	Blood	4.5	
302	VB-03-0604	3	PbAc	225		9.7	15	Blood	9:7	
303	VB-03-0613	3	PbAc	225		9.1	15	Blood	9.1	
322	VB-03-0629	3	PbAc	225		10.7	15	Blood	10.7	
327	VB-03-0645	3	PbAc	225		7.8	15	Blood	7.8	
342	VB-03-0602	3	PbAc	225		9.0	15	Blood	9.0	
313	VB-03-0626	4	Site 1	75 75		2.5	15	Blood	2.5	
328 329	VB-03-0598 VB-03-0640	4	Site 1	75 75		. 3.0	15 15	Blood Blood	3.0 4.0	
329 333	VB-03-0623	4	Site 1	75 75		4.0 3.6	15	Blood	3.6	
348	VB-03-0634	4	Site 1	75 75		4.8	15	Blood	4.8	
317	VB-03-0630	5	Site 1	225		9.4	15	Blood	9.4	
324	VB-03-0614	5	Site 1	225		9.5	15	Blood	9.5	•
340	VB-03-0597	5	Site 1	225		5.5	15	Blood	5.5	
341	VB-03-0621	5	Site 1	225		13.0	15	Blood	13.0	
354	VB-03-0644	5	Site 1	225		10.9	15	Blood	10.9	
321	VB-03-0600	6	Site 1	500		13.0	15	Blood	13.0	
332	VB-03-0647	6	Site 1	500		14.6	15	Blood	14.6	
335	VB-03-0641	6	Site 1	500		11.1	15	Blood	11.1	
355	VB-03-0603	6	Site 1	500		13.5	15	Blood	13.5	
363	VB-03-0611	6	Site 1	500		11.2	15	Blood	11.2	
305	VB-03-0620	7	Site 2	75		5.7	15	Blood	5.7	
306	VB-03-0606	7	Site 2	75		5.2	15	Blood	5.2	
316	VB-03-0642	7	Site 2	75		4.0	15	Blood	4.0	
320	VB-03-0619	7	Site 2	75		5.3	15	Blood	5.3	
347	VB-03-0643	7	Site 2	75		2.8	15	Blood	2.8	
301	VB-03-0607	8	Site 2	225		7.9	15	Blood	7.9	
318	VB-03-0646	8	Site 2	225		10.2	15	Blood	10.2	
352	VB-03-0612		Site 2	225		11.0	15	Blood	11.0 .	
356	VB-03-0631	8	Site 2	225		6.3	15	Blood	6.3	
358	VB-03-0636		Site 2	225		8.0	15	Blood	8.0	
309	VB-03-0599		Site 2	500		13.1	15	Blood	13.1	
330	VB-03-0617		Site 2	500		16.2	15	Blood	16.2	
351	VB-03-0627		Site 2	500		10.8	15	Blood	10.8	
353	VB-03-0615		Site 2	500		12.1	15	Blood	12.1	
359	VB-03-0624	9	Site 2	500		12.7	15	Blood	12.7	
325	VB-03-0637	10	Control	0	<	1.0	15	Blood	0.5	
337 360	VB-03-0632 VB-03-0639	10 10	Control Control	0	<	1.0 1.0	15 15	Blood Blood	0.5 0.5	

Non-detects evaluated using 1/2 the quantitation limit, laboratory results (ug/L) converted to concentration in blood (ug/dL) by dividing by dilution factor of 1 dL/L

TABLE A-4 BLOOD LEAD OUTLIERS

Flagged Data Points
Outliers

test	target	Actual						BLOOD LEA	AD (ug/dL)	BY DAY			
material	dosage	Dose*	group	pig#	0	1	2	3	5	7	9	12	15
PbAc	25	24.87	1 .	311	0.5	0.5	0.5	0.5	0.5	0.5	1.9	2.0	2.0
PbAc	25	25.71	1	326	2.0	0.5	0.5	2.8	1.3	1.0	2.0	2.9	3.2
PbAc	25	23.68	1	338	0.5	0.5	0.5	0.5	0.5	1.3	, 2.2	3.5	2.8
PbAc	25	29.71	1	343	0.5	0.5	0.5	0.9	1.4	0.5	2.2	2.2	2.5
PbAc	25	25.86	1	349	0.5	0.5	17.0	1.0	1.2	1.9	2.4	2.9	3.1
PbAc	75	71.55	2	304	0.5	2.0	2.0	2.6	2.8	3.8	4.3	5.5	6.3
PbAc	75	81.04	2	307	0.5	2.0	2.0	3.2	1.9	1.7	3.7	4.7	4.4
PbAc	75	84.25	2	312	0.5	1.0	3.0	3.8	4.5	5.2	5.4	5.9	4.9
PbAc	75	78.61	2	314	0.5	2.0	2.0	2.0	2.0	2.7	3.5	3.2	3.6
PbAc	75	74.31	2	350	0.5	2.0	2.0	2.4	2.8	2.4	3.5	4.8	4.5
PbAc	225	245.25	3	302	0.5	2.0	4.0	4.5	7.1	6.9	7.8	9.7	9.7
PbAc	225	214.61	3	303	0.5	2.0	4.9	6.7	6.3	6.3	7.6	9.1	9.1
PbAc	225	226.51	3	322	1	4.7	6.3	6.4	7.3	8.1	9.9	11.1	10.7
PbAc	225	244.91	3	327	0.5	3.0	4.2	5.0	6.3	6.8	7.2	9.4	7.8
PbAc	225	234.34	3	342	0.5	1.0	1.0	4.0	3.6	7.9	9.4	12.0	9.0
TM1	75	74.00	4	313	0.5	0.5	1.0	1.6	1.7	2.2	2.9	3.8	2.5
TM1	75	78.38	4	328	0.5	1.0	1.0	1.3	1.0	1.8	2.6	9.9	3.0
TM1	75	79.96	4	329	0.5	0.5	1.0	4.7	2.6	3.5	4.0	4.4	4.0
TM1	75	81.93	4	333	0.5	2.0	2.0	6.7	2.9	2.9	3.8	5.0	3.6
TM1	75	71.79	4	348	0.5	1.0	1.0	2.2	2.2	3.0	5.3	4.7	4.8
TM1	225	238.91	5	317	0.5	3.0	3.6	3.6	4.6	5.7	7.3	8.8	9.4
TM1	225	248.21	5	324	0.5	3.6	4.9	5.2	4.9	7.0	8.0	9.6	9.5
TM1	225 225	229.97	5	340	0.5	2.0	3.1	4.1	3.5	4.5	5.1	6.2	5.5
TM1 TM1	225 225	244.68 198.39	5 5	341 354	0.5 0.5	5. 2 2.0	4.7 3.0	4.7 3.3	6.5 5.4	9.6 7.8	10.5 10.1	9.7 10.2	13.0 10.9
TM1	500	534.79	6	321	0.5	4.0	5.7	7.7	12.0	8.2	11.3	17.0	13.0
				332	8.1	3.1		The state of the s	4.7				
TM1	500	506.85	6				4.6	3.8	Charles South Control	7.0	10.0	11.4	14.6
TM1	500 500	475.24	6	335	0.5	3.2	3.8	3.7	5.1	6.6	8.6	9.9	11.1.
TM1 TM1	500	536.63 511.79	6 6	355	0.5	6.9	7.8	10.1	9.2	9.8	11.7	13.6	13.5
TM2	75	79.95	7	363 305	0.5 0.5	4.0 0.5	5.2 2.0	6.4 1.5	6.8 2.6	6.5 3.3	9.5	10.0 4.5	11.2 5.7
TM2	75 75	74.54	7	306	0.5	1.0	29.0	2.6	1.7	3.7	4.4	4.6	5.2
TM2	75 75	79.36	7		1	<u>L</u>	23.0				3.1	-	
TM2	75 75	79.36 78.01	7	316 320	0.5 0.5	1.0 1.0	2.0	2.5 1.3	2.6 1.2	2.8 2.5	3.1 2.8	4.0 5.2	4.0 5.3
TM2	75 75	73.80	7	347	1	1.0	2.0 0.5		1.1		1.7		
TM2	225	237.88	8	301	0.5	3.0	3.0	0.5 3.8	5.2	1.5 8.0	6.2	3.3 7.1	2.8 7.9
TM2	225	230.40	8	318	0.5	2.0	2.0	3.5	5.4	6.3	7.3	9.0	10.2
TM2	225	219.81	8	352	0.5	3.9	3.7	3.8	6.7	7.4	11.4	12.0	11.0
TM2	225	236.32	8	35E	0.5	2.0	2.0	3.1	3.7	3.6	4.7	6.4	6.3
TM2	225	257.11	8	358	0.5	3.5	5.0	4.6	5.8	7.4	9.2	8.9	8.0
TM2	500	484.95	9	309	0.5	4.5	6.7	7.1	7.7	8.2	11.3	12.2	13.1
TM2	500	517.72	9	330	0.5	5.7	6.3	7.3	7.9	9.7	10.4	12.0	16.2
TM2	500	499.96	9	351	0.5	2.0	3.9	4.6	5.9	6.5	9.4	10.2	10.8
TM2	500	561.96	9	353	0.5	4.1	5.9	11.0	6.6	8.5	10.9	11.1	12.1
TM2	500	493.10	9	359	0.5	3.7	4.4	5.6	7.4	9.5	11.4	13.3	12.7
Control	0	0.00	10	325	0.5	0.5	0,5	0.5	0.5	0.5	0.5	1.5	0.5
Control	0	0.00	10	337	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5	0.5
Control	0	0.00	10	360	0.5	4.0	2.0	0.5	0.5	0.5	0.5	0.5	0.5

^{*} Average Time and Weight-Adjusted Dose for Each Pig

TABLE A-6 Area Under Curve Determinations

Calculated using interpolated values for excluded data as noted in Table A-5

				AUC (ı	ig/dL-days)	For Time S	oan Shown			AUC Lotal
group	pig#	0-1	1-2	2-3	3-5	5-7	7-9	9-12	12-15	(ug/dL-days
1	311	0.50	0.50	0.50	1.00	1.00	2.40	5.85	6.00	17.75
1	326	1.25	0.50	1.65	4.10	2.30	3.00	7.35	9.15	29.30
1	338	0.50	0.50	0.50	1.00	1.80	3.50	8.55	9.45	25.80
1	343	0.50	0.50	0.70	2.30	1.90	2.70	6.60	7.05	22.25
1	349	0.50	0.63	0.88	2.20	3.10	4.30	7.95	9.00	28.55
2	304	1.25	2.00	2.30	5.40	6.60	8.10	14.70	17.70	58.05
2	307	1.25	2.00	2.60	5.10	3.60	5.40	12.60	13.65	46.20
2	312	0.75	2.00	3.40	8.30	9.70	10.60	16.95	16.20	67.90
2	314	1.25	2.00	2.00	4.00	4.70	6.20	10.05	10.20	40.40
2	350	1.25	2.00	2.20	5.20	5.20	5.90	12.45	13.95	48.15
3	302	1.25	3.00	4.25	11.60	14.00	14.70	26.25	29.10	104.15
3	303	1.25	3.45	5.80	13.00	12.60	13.90	25.05	27.30	102.35
3	322	2.60	5.50	6.35	13.70	15.40	18.00	31.50	32.70	125.75
3	327	1.75	3.60	4.60	11.30	13.10	14.00	24.90	25.80	99.05
3	342	0.75	1.00	2.50	7.60	11.50	17.30	32.10	31.50	104.25
4	313	0.50	0.75	1.30	3.30	3.90	5.10	10.05	9.45	34.35
4	328	0.75	1.00	1.15	2.30	2.80	4.40	8.10	8.70	29.20
4	329	0.50	0.75	2.85	7.30	6.10	7.50	12.60	12.60	50.20
4	333	1.25	2.00	4.35	9.60	5.80	6.70	13.20	12.90	55.80
4	348	0.75	1.00	1.60	4.40	5.20	8.30	15.00	14.25	50.50
5	317	1.75	3.30	3.60	8.20	10.30	13.00	24.15	27.30	91.60
5	324	2.05	4.25	5.05	10.10	11.90	15.00	26.40	28.65	103.40
5	340	1.25	2.55	3.60	7.60	8.00	9.60	16.95	17.55	67.10
5	341	2.85	4.95	4.70	11.20	16.10	20.10	30.30	34.05	124.25
5	354	1.25	2.50	3.15	8.70	13.20	17.90	30.45	31.65	108.80
6	321	2.25	4.85	6.70	15.65	16.15	19.50	42.45	45.00	152.55
6	332	1.80	3.85	4.20	8.50	11.70	17.00	32.10	39.00	118.15
6	335	1.85	3.50	3.75	8.80	11.70	15.20	27.75	31.50	104.05
6	355	3.70	7.35	8.95	19.30	19.00	21.50	37.95	40.65	158.40
6	363	2.25	4.60	5.80	13.20	13.30	16.00	29.25	31.80	116.20
7	305	0.50	1.25	1.75	4.10	5.90	7.70	13.35	15.30	49.85
7	306	0.75	1.40	2.20	4.30	5.40	8.10	13.50	14.70	50.35
7	316	0.75	1.50	2.25	5.10	5.40	5.90	10.65	12.00	43.55
7	320	0.75	1.50	1.65	2.50	3.70	5.30	12.00	15.75	43.15
7	347	0.75	0.75	0.50	1.60	2.60	3.20	7.50	9.15	26.05
8	301	1.75	3.00	3.40	9.00	13.20	14.20	19.95	22.50	87.00
8	318	1.25	2.00	2.75	8.90	11.70	13.60	24.45	28.80	93.45
8	352	2.20	3.80	3.75	10.50	14.10	18.80	35.10	34.50	122.75
8	356	1.25	2.00	2.55	6.80	7.30	8.30	16.65	19.05	63.90
8	358	2.00	4.25	4.80	10.40	13.20	16.60	27.15	25.35	103.75
9	309	2.50	5.60	6.90	14.80	15.20	19.50	35.25	37.95	138.40
9	330	3.10	6.00	6.80	15.20	17.60	20.10	33.60	42.30	144.70
9	351	1.25	2.95	4.25	10.50	12.40	15.90	29.40	31.50	108.15
9	353	2.30	5.00	4.25 8.45	17.60	15.10	19.40	33.00	34.80	135.65
9	359	2.10	4.05	5.00	13.00	16.90	20.90	33.00 37.05	39.00	138.00
10	325	0.50	4.05 0.50	0.50	1.00	1.00	1.00	3.00	3.00	10.50
10	337	0.50	0.50	0.50	1.00	1.00	1.00	3.00 1.50	3.00 1.50	7.50
10	360	2.25	3.00	1.25	1.00	1.00	1.00	1.50	1.50	7.50 12.50
-10	300	<u> </u>	3.00	1.40	1.00	1.00	1.00	1.50	1,30	12.50

TABLE A - 7 TISSUE LEAD DATA

pig number	sample	group	material administered	dosage qualifier	result	day	MATRIX	Adjusted Value ()	Notes
311	VB-03-0689	1	PbAc	25	4.8	15	femur	2.4	
326	VB-03-0669	1	PbAc	25	5.4	15	femur	2.7	
338	VB-03-0667	1	PbAc	25	5.0	15	femur	2.5	
343	VB-03-0672	1	PbAc	25	5.0	15	femur	2.5	
349	VB-03-0674	1	PbAc	25	5.4	15	femur	2.7	
304	VB-03-0680	2	PbAc	75 ~~	8.5	15	femur	4.3	
307	VB-03-0693	2	PbAc	75 76	13.0	15	femur	6.5	
312	VB-03-0687	2	PbAc .	75 75	9.5	15	femur	4.8	
314	VB-03-0653	2 2	PbAc PbAc	75 75	14.0	15	femur	7.0	
350 302	VB-03-0683 VB-03-0679	3	PbAc	225	8.2 39.0	15 15	femur femur	4.1 19.5	
303	VB-03-0665	3	PbAc	225	36.0	15	femur	18.0	
322	VB-03-0695	3	PbAc	225	33.0	15	femur	16.5	
327	VB-03-0657	3	PbAc	225	39.0	15	femur	19.5	•
342	VB-03-0686	3	PbAc	225	30.0	15	femur	15.0	
313	VB-03-0676	4	Site 1	75	7.5	15	femur	3.8	•
328	VB-03-0694	4	Site 1	75	7.3	15	femur	3.7	
329	VB-03-0685	4	Site 1	75	11.0	15	femur	5.5	
333	VB-03-0655	4	Site 1	75	7.9	15	femur	4.0	
348	VB-03-0677	4	Site 1	75	43.0	15	femur	21.5	
317	VB-03-0688	5	Site 1	225	29.0	15	femur	14.5	
324	VB-03-0656	5	Site 1	225	25.0	15	femur	12.5	
340	VB-03-0659	5	Site 1	225	22.0	15	femur	11.0	
341 354	VB-03-0692 VB-03-0700	5 5	Site 1	225 225	29.0 20.0	15 15	femur femur	14.5 10.0	
354 321	VB-03-0681	6	Site 1	500	20.0 56.0	15	femur femur	28.0	
332	VB-03-0697	6	Site 1	500	48.0	15	femur	24.0	
335	VB-03-0652	6	Site 1	500	35.0	15	femur	17.5	
355	VB-03-0664	6	Site 1	500	48.0	15	femur	24.0	
363	VB-03-0661	6	Site 1	500	57.0	15	femur	28.5	
305	VB-03-0658	7	Site 2	75	13.0	15	femur	6.5	
306	VB-03-0698	7	Site 2	75	9.5	15	femur	4.8	
316	VB-03-0650	7	Site 2	75 75	11.0	15	femur	5.5	
320	VB-03-0670	7	Site 2	75 76	9.2	15	femur	4.6	
347 301	VB-03-0654 VB-03-0662	7 8	Site 2 Site 2	75 225	5.6 25.0	15 15	femur femur	2.8 12.5	
318	VB-03-0682	8	Site 2	225	8.0	15	femur	4.0	
352	VB-03-0666	8	Site 2	225	37.0	15	femur	18.5	
356	VB-03-0671	8	Site 2	225	23.0	15	femur	11.5	
358	VB-03-0663	8	Site 2	225	29.0	15	femur	14.5	
309	VB-03-0696	9	Site 2	500	38.0	15	femur	19.0	
330	VB-03-0668	9	Site 2	500	46.0	15	femur	23.0	
351	VB-03-0675	9	Site 2	500	36.0	15	femur	18.0	
353	VB-03-0690	9	Site 2	500	43.0	15	femur	21.5	
359	VB-03-0660	9	Site 2	500	40.0	15	femur	20.0	
325	VB-03-0678	10	Control	0 <	1.0	15	femur	0.3	
337 360	VB-03-0699 VB-03-0684	10 10	Control Control	0 0	1.0 1.0	15 15	femur femur	0.5 0.5	
311	VB-03-0776	1	PbAc	25	4.8	15	kidney	48.0	
326	VB-03-0774	1	PbAc	25	7.3	15	kidney	73.0	
338	VB-03-0791	1	PbAc	25	3.0	15	kidney	30.0	
343	VB-03-0758	1	PbAc	25	4.4	15	kidney	44.0	
349	VB-03-0760	1	PbAc	25	4.9	15	kidney	49.0	
304	VB-03-0769	2	PbAc	75	12.0	15	kidney	120.0	
307	VB-03-0765	2	PbAc	75	15.0	15	kidney	150.0	
312	VB-03-0800	2	PbAc	75	12.0	15	kidney	120.0	
314	VB-03-0762	2	PbAc	75 75	11.0	15	kidney	110.0	
350 302	VB-03-0785 VB-03-0764	2 3	PbAc PbAc	75 225	14.0 34.0	15 15	kidney kidney	140.0 340.0	
303	VB-03-0802	3	PbAc	225	37.0	15	kidnéy	370.0	
322	VB-03-0777	3	PbAc	225	56.0	15	kidney	560.0	
327	VB-03-0753	3	PbAc	225	36.0	15	kidney	360.0	
342	VB-03-0784	3	PbAc	225		15	kidney	#VALUE!	Missing
313	VB-03-0768	4	Site 1	75	7.4	15	kidney	74.0	
328	VB-03-0797	4	Site 1	75	11.0	15	kidney	110.0	
329	VB-03-0788	4	Site 1	75	10.0	15	kidney	100.0	
333	VB-03-0793	4	Site 1	75 75	8.9	15	kidney	89.0	Minda
348	VB-03-0759	4 5	Site 1	75 225	37.0	15 15	kidney	#VALUE! 370.0	Missing
317 324	VB-03-0773 VB-03-0752	5	Site 1	225	34.0	15	kidney kidney	340.0	
340	VB-03-0786	5	Site 1	225	23.0	15	kidney	230.0	
341	VB-03-0790	5	Site 1	225	41.0	15	kidney	410.0	
354	VB-03-0787	5	Site 1	225	41.0	15	kidney	410.0	
321	VB-03-0778	6	Site 1	500	95.0	15	kidney	950.0	
332	VB-03-0775	6	Site 1	500	92.0	15	kidney	920.0	
335	VB-03-0761	6	Site 1	500	53.0	15	kidney	530.0	
355	VB-03-0771	6	Site 1	500	58.0	15	kidney	580.0	
363	VB-03-0780	6	Site 1	500 75	75.0 14.0	15 15	kidney	750.0 140.0	
305	VB-03-0757	7 7	Site 2 Site 2	75 75	14.0 8.4	15 15	kidney kidney	84.0	
306 316	VB-03-0789 VB-03-0796	7	Site 2	75 75	8.8	15	kidney	88.0	
320	VB-03-0770	7	Site 2	75	10.0	15	kidney	100.0	
347	VB-03-0798	7	Site 2	75	4.3	15	kidney	43.0	
	-						•		

pig number	sample	group	material administered	ALCOHOLD TO THE REAL PROPERTY AND ADDRESS OF THE PERTY ADDRESS OF THE PE	qualifier	result	day	MATRIX	Adjusted Value ()*	Notes
301	VB-03-0781	8	Site 2	225		35.0	15	kidney	350.0 ·	
318	VB-03-0779	8	Site 2	225			15	kidney	#VALUE!	Missing
352	VB-03-0794	8	Site 2	225		54.0	15	kidney	540.0	
356	VB-03-0767	8	Site 2	225		22.0	15	kidney	220.0	
358	VB-03-0772	8	Site 2	225		28.0	15	kidney	280.0	
309	VB-03-0792	9	Site 2	500		58.0	15	kidney	580.0	
330	VB-03-0754	9	Site 2	500		100.0	15	kidney	1000.0	
351	VB-03-0766	9 -	Site 2	500		54.0	15	kidney	540.0	
353	VB-03-0799	9	Site 2	500		56.0	15	kidney	560.0	
359	VB-03-0801	9	Site 2	500		64.0	15	kidney	640.0	
325	VB-03-0782	10	Control	0	<	1.0	15	kidney	5.0	
337	VB-03-0756	10	Control	0	<	1.0	15	kidney	5.0	
360	VB-03-0795	10	_Control	0	<	1.0	15	kidney	5.0	_
311	VB-03-0702	1	PbAc	25		8.7	15	liver	87.0	
326	VB-03-0717	1	PbAc	25		12.0	15	liver	120.0	
338	VB-03-0728	1	PbAc	25		9.7	15	liver	97.0	
343	VB-03-0701	1	PbAc	25		10.0	15	liver	100.0	
349	VB-03-0713	1	PbAc	25		8.6	15	liver	86.0	
304	VB-03-0708	2	PbAc	75		31.0	15	liver	310.0	
307	VB-03-0746	2	PbAc	75		20.0	15	liver	200.0	
312	VB-03-0744	2	PbAc	75		20.0	15	liver	200.0	
314	VB-03-0722	2	PbAc	75		16.0	15	liver	160.0	
350	VB-03-0706	2	PbAc	75		32.0	15	liver	320.0	
302	VB-03-0720	3	PbAc	225		54.0	15	liver	540.0	
303	VB-03-0743	3	PbAc	225		89.0	15	liver	890.0	
322	VB-03-0735	3	PbAc	225		70.0	15	liver	700.0	
327	VB-03-0724	3	PbAc	225		49.0	15	liver	490.0	
342	VB-03-0710	3	PbAc	225		100.0	15	liver	1000.0	
313	VB-03-0727	4	Site 1	75		9.0	15	liver	90.0	
328	VB-03-0703	4	Site 1	75		18.0	15	liver	180.0	
329	VB-03-0742	4	Site 1	75		13.0	15	liver	130.0	
333	VB-03-0721	4	Site 1	75		14.0	15	liver	140.0	
348	VB-03-0734	4	Site 1	75		14.0	15	liver	140.0	
317	VB-03-0736	5	Site 1	225		88.0	15	liver	0.088	
324	VB-03-0718	5	Site 1	225		86.0	15	liver	860.0	
340	VB-03-0715	5	Site 1	225		47.0	15	liver	470.0	
341	VB-03-0719	5	Site 1	225		110.0	15	liver	1100.0	
354	VB-03-0723	5	Site 1	225		61.0	15	liver	610.0	
321	VB-03-0749	6	Site 1	500		180.0	15	liver	1800.0	
332	VB-03-0704	6	Site 1	500		160.0	15	liver	1600.0	
335	VB-03-0716	6	Site 1	500		130.0	15	liver	1300.0	
355	VB-03-0740	6	Site 1	500		130.0	15	liver	1300.0	
363	VB-03-0726	6	Site 1	500		96.0	15	liver	960.0	
305	VB-03-0733	7	Site 2	75		18.0	15	liver	180.0	
306	VB-03-0748	7	Site 2	75		14.0	15	liver	140.0	
316	VB-03-0730	7	Site 2	75		15.0	15	liver	150.0	
320	VB-03-0741	7	Site 2	75		15.0	15	liver	150.0	
347	VB-03-0711	7.	Site 2	75		11.0	15	liver	110.0	
301	VB-03-0709	8	Site 2	225		40.0	15	liver	400.0	
318	VB-03-0729	8	Site 2	225		76.0	15	liver	760.0	
352	VB-03-0731	8 -	Site 2	225		70.0	15	liver	700.0	
356	VB-03-0751	8	Site 2	225		35.0	15	liver	350.0	
358	VB-03-0747	8	Site 2	225		60.0	15	liver	600.0	
309	VB-03-0738	9	Site 2	500		120.0	15	liver	1200.0	
330	VB-03-0737	9	Site 2	500		230.0	15	liver	2300.0	
351	VB-03-0725	9	Site 2	500		97.0	15	liver	970.0	
353	VB-03-0732	9	Site 2	500		89.0	15	liver	890.0	
359	VB-03-0712	9	Site 2	500		120.0	15	liver	1200.0	
325	VB-03-0705	10	Control	0	<	1.0	15	liver	5.0	
337	VB-03-0739	10	Control	0		1.0	15	liver	10.0	
360	VB-03-0750	10	Control	0	<	1.0	15	liver	5.0	

Non-detects evaluated using 1/2 the quantitation limit. Laboratory results (ug/L) converted to tissue concentrations by dividing by sample dilution factors of 0.1 kg/L (liver, kidney) or 2 g/L (ashed bone). Final units are ug Pb/kg wet weight (liver, kidney) or ug Pb/g ashed bone (femur)

TABLE A-8 SUMMARY OF ENDPOINT OUTLIERS

Selected Outliers

test	target	Actual				MEASUREN	MENT ENDPOINT	
material	dosage	Dose*	group	pig#	Blood	Femur	Liver	Kidney
PbAc	25	24.87	1	311	17.8	2.4	87 ·	48
PbAc	25	25.71	1	326	29.3	2.7	120	73
PbAc	25	23.68	1	338	25.8	2.5	97	30
PbAc	25	29.71	1	343	22.3	2.5	100	44
PbAc	25	25.86	1	349	28.6	2.7	86	49
PbAc	75	71.55	2	304	58.1	4.25	310	120
PbAc	75	81.04	2	307	46.2	6.5	200	150
PbAc	75	84.25	2	312	67.9	4.75	200	120
PbAc	75	78.61	2	314	40.4	7	160	110
PbAc	75	74.31	2	350	48.2	4.1	320	140
PbAc	225	245.25	3	302	104.2	19.5	540	340
PbAc	225	214.61	3	303	102.4	18	890	370
PbAc	225	226.51	3	322	125.8 b	16.5	700	560 b
PbAc	225	244.91	3	327	99.1	19.5	490	360
PbAc	225	234.34	3	342	104.3	15	1000	Missing
TM1	75	74.00	4	313	34.4	3.75	90	74
TM1	75	78.38	4	328	29.2	3.65	180	110
TM1	75	79.96	4	329	50.2	5.5	130	100
TM1	75	81.93	4	333	55.8	3.95	140	89
TM1	75	71.79	4	348	50.5	21.5 b	140	Missing
TM1	225	238.91	5	317	91.6	14.5	880	370
TM1	225	248.21	5	324	103.4	12.5	860	340
TM1	225	229.97	5	340	67.1	11	470	230
TM1	225	244.68	5	341	124.3	14.5	1100	410
TM1	225	198.39	5	354	108.8	10	610	410
TM1	500	534.79	6	321	152.6	28	1800	950
TM1	500	506.85	6	332	118.2	24	1600	920
TM1	500	475.24	6	335	104.1	17.5 b	1300	530
TM1	500	536.63	6	355	158.4	24	1300	580
TM1	500	511.79	6	363	116.2	28.5	960 b	750
TM2	75	79.95	7	305	49.9	6.5	180	140
TM2	75	74.54	7	306	50.4	4.75	140	84
TM2	75	79.36	7	316	43.6	5.5	150	88
TM2	75	78.01	7	320	43.2	4.6	150	100
TM2	75	73.80	7	347	26.1	2,8	110	43.
TM2	225	237.88	8	301	87.0	12.5	400	350
TM2	225	230.40	8	318	93.5	4 b	760	Missing
TM2	225	219.81	8	352	122.8	18.5 b	700	540
TM2	225	236.32	8	356	63.9	11.5	350	220
TM2	225	257.11	8	358	103.8	14.5	600	280
TM2	500	484.95	9	309	138.4	19	1200	580
TM2	500	517.72	9	330	144.7	23	2300 b	1000 b
TM2	500	499.96	9	351	108.2	18	970	540
TM2	500	561.96	9	353	135.7	21.5	890	560
TM2	500	493.10	9	359	138.0	20	1200	640
Control	0	0.00	10	325	10.5	0.25	5	5
Control	0	0.00	10	337	7.5	0.5	10	5
Control	0	0.00	10	360	12.5	0.5	5	5

a a priori outlier determinations (none selected in this study)

b Outside 95% Prediction Intervals
 c Professional judgement - Response was lower than that seen in next lowest dose group

TABLE A-9 Best Curve Fit Parameters

BLOOD		BONE		LIVER		KIDNEY	
PbAc Curve -	Ехр	PbAc Curve -	Linear	PbAc Curve	- Linear	PbAc Curve -	Linear
а	. 9	a	0.75	a	10.9	a	9.8
b		b	0.071	b	3.01	b	1.48
C	141.9	C		C		C	
d	0.0046	d		d		d	
R2	0.971	R2	0.973	R2	0.842	R2	0.979
					•		
Untreated Curv	/e - Exp	Untreated Cu	rve - Exp	Untreated Cu	ırve - Exp	Untreated Curve	- Exp
а	9	a	0.75	а	10.9	а	9.8
b	ū	ĥ	0.049	b	2.96	b	1.44
c .	141.9	č		· c		C	
ď	0.004	d		d		ď	
R2	0.905	R2	0.981	R2	0.935	R2	0.909
P-treated Curv	е- Ехр	P-treated Cur	ve - Linear	P-treated Cu	rve - Linear	P-treated Curve	- Linear
a	9	a	0.75	а	10.9	я	9.8
<u>.</u>	J	, h	0.04	b	2.096	b	1.16
č	141.9	ć	3104	c	2.000	č	
ď	0.0039	ď		ď		d	
R2	0.931	R2	0.961	Ř2	0.901	R2	0.89
						• —	

Equatio	ns Used	
EXP	Y=a+c*(1-exp(-d*dose))	
LIN	Y=a+b*dose	

TABLE A-10 Relative Bioavailability of Lead in Test Materials

	Test Material				
Endpoint	TM1	TM2			
Blood	0.87	0.85			
Liver	0.98	0.70			
Kidney	0.97	0.78			
Bone	0.69	0.56			

Definitions

Plausible Range:

RBA(Blood) to mean RBA for Tissues

Preferred Range:

RBA(Blood) to (RBA(Blood) + RBA(Tissues))/2

Suggested Point Est:

1/2(RBA(Blood) + (RBA(Blood)+RBA(Tissues))/2)

Relative Bioavailability

	TM	1	T	И 2
Plausible Range	0.87	0.88	0.85	0.68
Preferred Range	0.87	0.88	0.85	0.76
Point Estimate	0.8	7	0.	81

Absolute Bioavailability

	TM	1	TI	M2
Plausible Range	43%	44%	42%	34%
Preferred Range	43%	44%	42%	38%
Point Estimate	449	%	40)%

TABLE A-11 INTRALABORATORY DUPLICATES

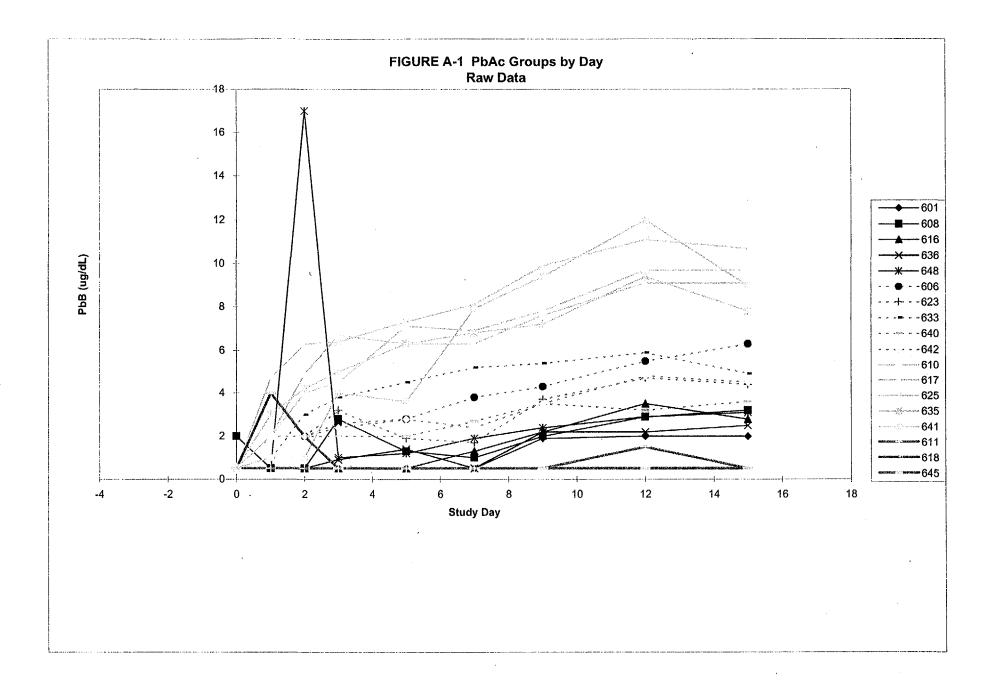
RPD = Relative Percent Difference RPD = 100*[Orig-Dup]/((Orig+Dup)/2 * Non detects evaluated at 1/2 DL

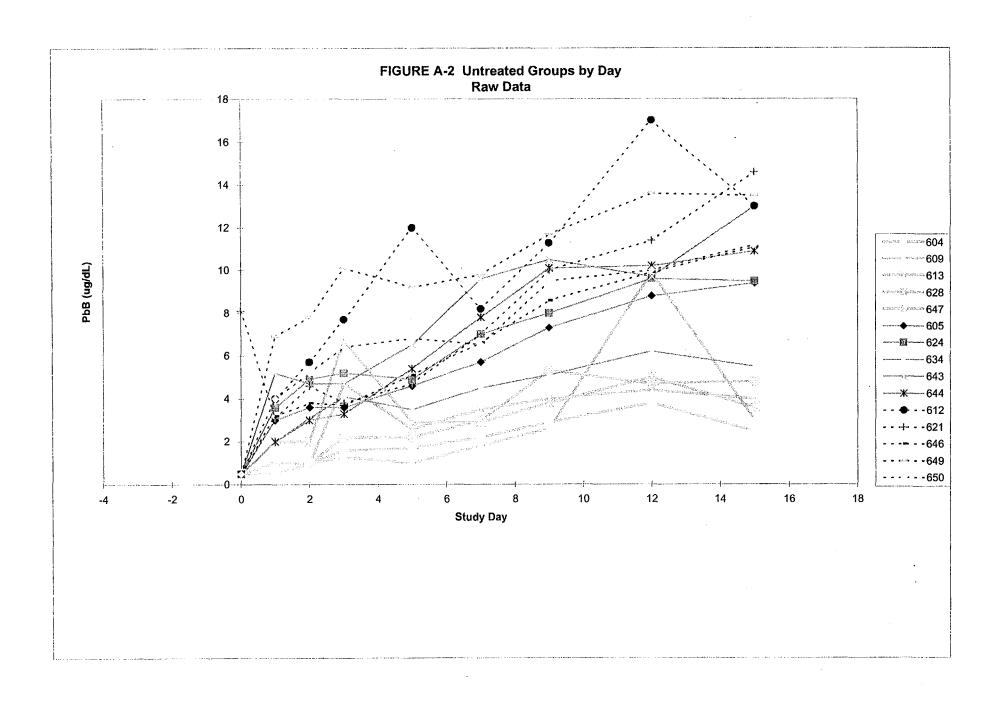
Pig number	group	material administered	dosage	day	matrix	Duplicate Value*	Original Value*	Average	RPD	Avg RPD	
349	1	PbAc	25	0	Blood	0.5	0.5	0.5	0%		
335	6	Site 1	500	0	Blood	0.5	0.5	0.5	0%		
330	9	Site 2	500	0	Blood	0.5	0.5	0.5	0%		
350	2	PbAc	75	1	Blood	2.0	2.0	2.0	0%	•	
342	3	PbAc	225	1	Blood	2.0	1.0	1.5	-67%		
320	7	Site 2	75	1	Blood	2.0	1.0	1.5	-67%	•	
358	8	Site 2	225	2	Blood	5.5	5.0	5.3	-10%		
351	9	Site 2	500	2	Blood	3.9	3.9	3.9	0%		
325	10	Control	0	2	Blood	0.5	0.5	0.5	0%		
318	8	Site 2	225	3	Blood	3.9	3.5	3.7	-11%		
337	10	Control	0	3	Blood	1.0	0.5	0.8	-67%		
360	10	Control	0	3	Blood	1.0	0.5	0.8	-67%		
348	4	Site 1	75	5	Blood	2.0	2.2	2.1	10%		
330	9	Site 2	500	5	Blood	8.3	7.9	8.1	-5%		
359	9	Site 2	500	5	Blood	8.0	7.4	7.7	-8%		
322	3	PbAc	225	7	Blood	7.8	8.1	8.0	4%		
320	7	Site 2	75	7	Blood	1.9	2.5	2.2	27%	•	
351	9	Site 2	500	7	Blood					1 ·	
307	2	PbAc	75	9	Blood	4.5	3.7	4.1	-20%	4	
302	3	PbAc	225	9	Blood	8.3	7.8	8.1	-6%		
347	7	Site 2	75	9	Blood	2.2	1.7	2.0	-26%	•	
349	1	PbAc	25	12	Blood	3.8	2.9	3.4	-27%		
342	3	PbAc	225	12	Blood	12.3	12.0	12.2	-2%		
355	6	Site 1	500	12	Blood	15.2	13.6	14.4	-11%		
332	6	Site 1	500	15	Blood	14.1	14.6	14.4	3%	*	
305	7	Site 2	75	15	Blood	4.4	5.7	5.1	26%		
309	9	Site 2	500	15	Blood	13.0	13.1	13.1	1%	-12%	BLOOD
305	7	Site 2	75	15	femur	6.0	6.5	6.3	8%		
309	9	Site 2	500	15	femur	20.0	19.0	19.5	-5%		
332	6	Site 1	500	15	femur	23.0	24.0	23.5	4%	2%	FEMUR
321	6	Site 1	500	15	kidney	280.0	950.0	615.0	109%		
340	5	Site 1	225	15	kidney	240.0	230.0	235.0	-4%		
347	7	Site 2	75	15	kidney	52.0	43.0	47.5	-19%	29%	KIDNEY
342	3	PbAc	225	15	liver	750.0	1000.0	875.0	29%		•
358	8	Site 2	225	15	liver	560.0	600.0	580.0	7%		
359	9	Site 2	500	15	liver	1400.0	1200.0	1300.0	-15%	7%	LIVER

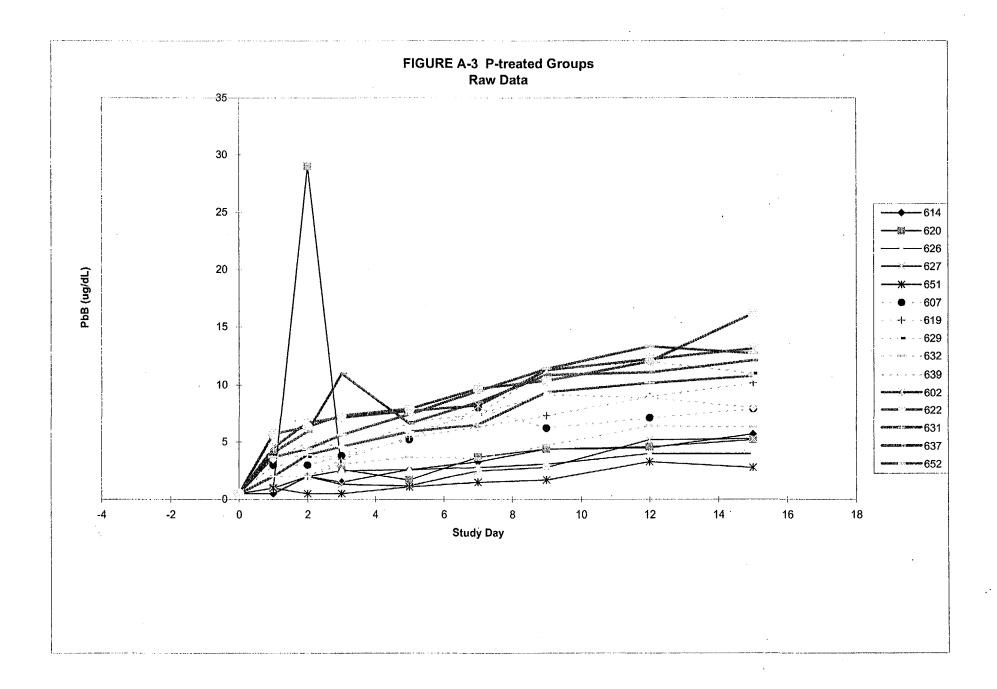
Values determined to be outliers (Original = 6.5; Duplicate = 28.7)

TABLE A-12 CDC STANDARDS

			-	Measured*			Nominal	
Sample ID	Day	Q	Low Std	Med Std	High Std	Low Std	Med Std	High Std
3.1	0		2.0			1.7	4.8	14.9
3.1	1		2.0			1.7	4.8	14.9
3.1	3		1.0	1		1.7	4.8	14.9
3.1	5		2.1			1.7	4.8	14.9
3.1	12		2.7			1.7	4.8	14.9
3.2	1			3.9		1.7	4.8	14.9
3.2	2			3.6		1.7	4.8	14.9
3.2	15			3.5	'	1.7	4.8	14.9
3.3	0				14.0	1.7	4.8	14.9
3.3	2	•		ŀ	14.0	1.7	4.8	14.9
· 3.3	3				14.9	1.7	4.8	14.9
3.3	5				8.2	1.7	4.8	14.9
3.3	7				13.7	1.7	4.8	14.9
3.3	7		•		13.4	1.7	4.8	14.9
3.3	9				14.7	1.7	4.8	14.9
3.3	9				14.3	1.7	4.8	14.9
3.3	12				14.4	1.7	4.8	14.9
3.3	15				14.1	1.7	4.8	14.9
		Avg	2.0	3.7	13.6			







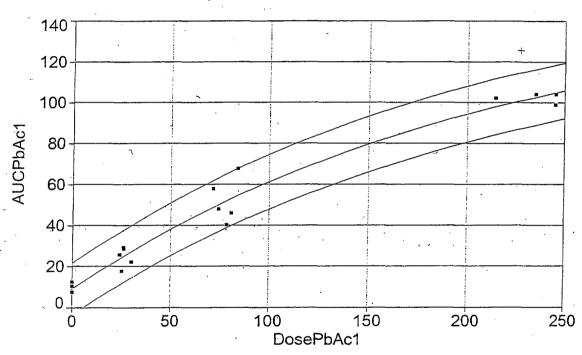
for the fix of the first of the first of the fix

FIGURE A-4 BEST FIT CURVE WITH 95% PREDICTION INTERVALS (generated using Table Curve 2D v. 3.0. Outliers represented by "+"

PbAc - AUC

Rank 2 Eqn 8001 [UDF 1] y=AUC(a)

r²=0.971143 DF Adj r²=0.9692192 FitStdErr=5.9820592 Fstat=538.45812



Rank 2 Eqn 8001 [UDF 1] y=AUC(a)

r ² Coef Det	DF Adj r ²	Fit Std Err	F-value
0.9711429962	-0.9692191960	5.9820591731	538.45811828

Parm	Value	Std Error	t-value	95% Confide	nce Limits
а	0.004602252	0.000215463	21.35986735	0.004144553	0.005059950

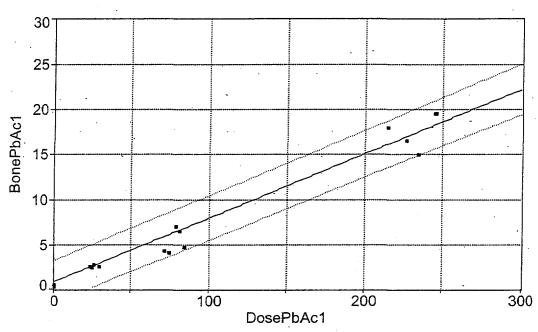
Date	Time	File Source
Jan 9, 2001	3:01:16 PM	CLIPBRD.WK1

FIGURE A-5 BEST FIT CURVE WITH 95% PREDICTION INTERVALS (generated using Table Curve 2D v. 3.0. Outliers represented by "+"

PbAc - Bone

Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

r²=0.9729649 DF Adj r²=0.9712752 FitStdErr=1.1470933 Fstat=611.81208



Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

r² Coef Det DF Adj r² Fit Std Err F-value 0.9729648960 0.9712752020 1.1470933295 611.81208007

Parm Value Std Error t-value 95% Confidence Limits a 0.071267046 0.002185359 32.61113966 0.066650512 0.075883581

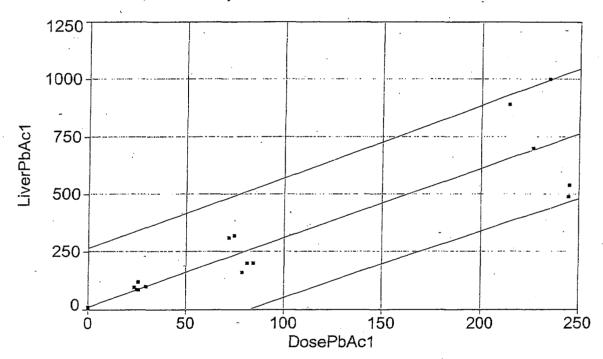
Date Time File Source Feb 9, 2001 12:58:45 PM CLIPBRD.WK1

FIGURE A-6 BEST FIT CURVE WITH 95% PREDICTION INTERVALS (generated using Table Curve 2D v. 3.0. Outliers represented by "+"

PbAc - Liver

Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

 r^2 =0.84202407 DF Adj r^2 =0.83215058 FitStdErr=120.78928 Fstat=90.611333



Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

0.842024		6321505779	120.78927910	90.611332677	
	Value .011262498	Std Error 3 0.230118969	t-value 13.08567700	95% Confider 2.525140049	

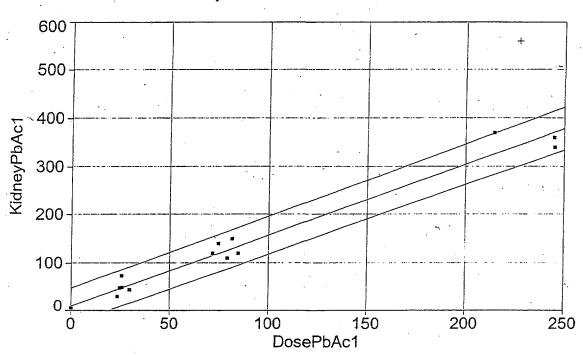
Date Time File Source
Jan 15, 2001 9:58:31 AM CLIPBRD.WK1

FIGURE A-7 BEST FIT CURVE WITH 95% PREDICTION INTERVALS (generated using Table Curve 2D v. 3.0. Outliers represented by "+"

PbAc - Kidney

Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

r²=0.97924659 DF Adj r²=0.9777642 FitStdErr=17.898238 Fstat=707.77282



Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

Parm Value Std Error t-value 95% Confidence Limits a 1.475620344 0.042454579 34.75762545 1.385388352 1.565852335

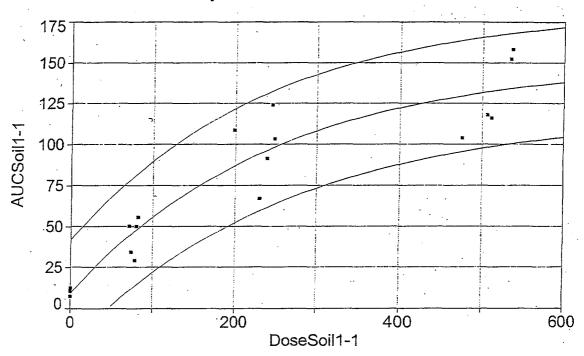
Date Time File Source
Jan 15, 2001 10:00:26 AM CLIPBRD.WK1

FIGURE A-8 BEST FIT CURVE WITH 95% PREDICTION INTERVALS (generated using Table Curve 2D v. 3.0. Outliers represented by "+"

TM#1 - AUC

Rank 1 Eqn 8009 [UDF 2] y=AUC NC(a)

r²=0.90451184 DF Adj r²=0.89854383 FitStdErr=15.595457 Fstat=161.03255



Rank 1 Eqn 8009 [UDF 2] y=AUC NC(a)

r ² Coef Det	DF Adj r ²	Fit Std Err	F-value
0.9045118438	0.8985438341	15.595456570	161.03255068

Parm	value '	Std Error	t-value	95% Confide	nce Limits
а	0.004003848	0.000411061	9.740274613	0.003135488	0.004872208

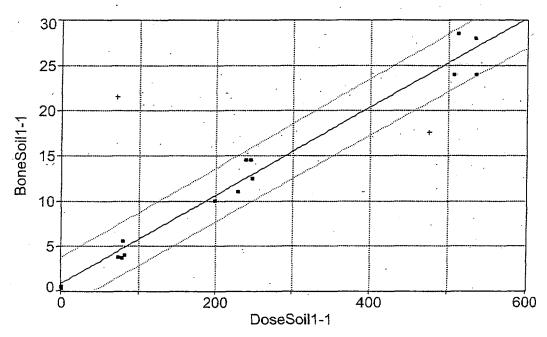
Date	Time	File Source
Jan 9, 2001	3:01:41 PM	CLIPBRD.WK1

FIGURE A-9 BEST FIT CURVE WITH 95% PREDICTION INTERVALS (generated using Table Curve 2D v. 3.0. Outliers represented by "+"

TM#1 - Bone

Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

r²=0.98120086 DF Adj r²=0.97985807 FitStdErr=1.3760025 Fstat=782.90896



Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

r ² Coef Det	DF Adj r ²	Fit Std Err	F-yalue
0.9812008628	. 0.9798580673	1.3760025073	782.90895873

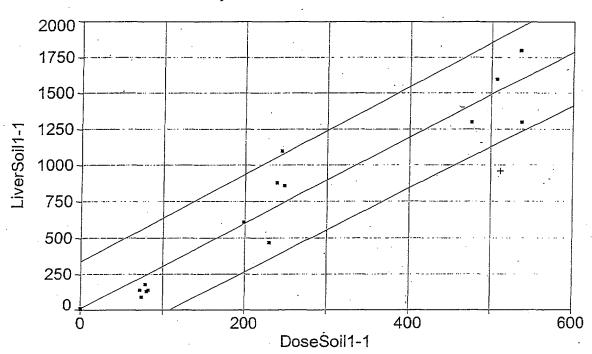
Parm	ı Value	Std Error	t-value	95% Confide	nce Limits
a	0.048897266	0.001238993	39.46533933	0.046263939	0.051530592

Date	Time	File Source
Feb 9, 2001	1:00:34 PM	CLIPBRD.WK1

FIGURE A-10 BEST FIT CURVE WITH 95% PREDICTION INTERVALS (generated using Table Curve 2D v. 3.0. Outliers represented by "+"

TM#1 – Liver Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

r²=0.93488562 DF Adj r²=0.93054466 FitStdErr=155.52173 Fstat=229.72145



Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

•			
r ² Coef Det	DF Adj r ²	Fit Std Err	F-value
0.9348856184	0.9305446597	155.52173092	229.72144606

Parm	Value	Std Error	t-value	95% Confide	nce Limits
a ·	2.964784622	0.141156976	21.00345800	2.664930509	3.264638735

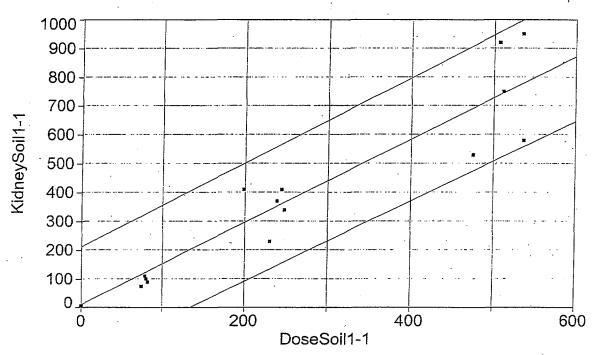
Date	Time	File Source
Jan 15, 2001	9:59:06 AM	CLIPBRD.WK1

FIGURE A-11 BEST FIT CURVE WITH 95% PREDICTION INTERVALS (generated using Table Curve 2D v. 3.0. Outliers represented by "+"

· TM#1 - Kidney

Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

 $r^2 = 0.9089 \overset{\cdot}{1}658 \ \ \mathsf{DF} \ \mathsf{Adj} \ r^2 = 0.90284435 \ \ \mathsf{FitStdErr} = 95.251142 \ \ \mathsf{Fstat} = 159.66314$



Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

r ² Coef Det	DF Adj r ²	Fit Std Err	F-value
0.9089165776	0.9028443495	95.251142383	159.66314030

Parm	Value	Std Error	t-value	95% Confide	nce Limits
а	1.436303152	0.079279311	18.11699849	1.267893286	1.604713017

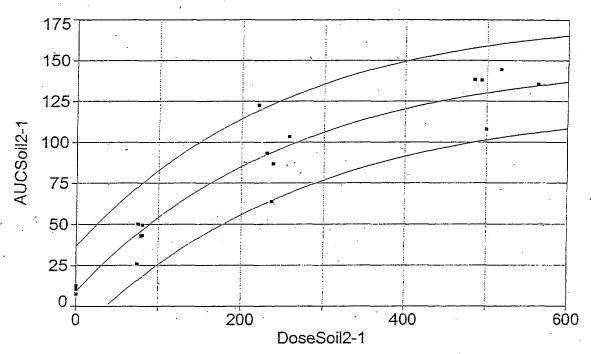
Date	Time	File Source
Jan 15, 2001	10:00:51 AM	CLIPBRD.WK1

FIGURE A-12 BEST FIT CURVE WITH 95% PREDICTION INTERVALS (generated using Table Curve 2D v. 3.0. Outliers represented by "+"

TM#2 - AUC

Rank 1 Eqn 8009 [UDF 2] y=AUC NC(a)

r²=0.93103012 DF Adj r²=0.9267195 FitStdErr=13.17285 Fstat=229.48439



Rank 1 Eqn 8009 [UDF 2] y=AUC NC(a)

r² Coef Det DF Adj r² Fit Std Err F-value 0.9310301163 0.9267194985 13.172849831 229.48439407

Parm Value Std Error t-value 95% Confidence Limits a 0.003858687 0.000332455 11.60663730 0.003156381 0.004560993

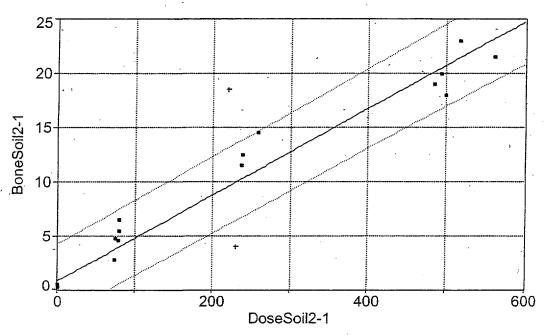
Date Time File Source
Jan 9, 2001 3:02:04 PM CLIPBRD.WK1

FIGURE A-13 BEST FIT CURVE WITH 95% PREDICTION INTERVALS (generated using Table Curve 2D v. 3.0. Outliers represented by "+"

TM#2 - Bone

Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

 r^2 =0.96128686 DF Adj r^2 =0.95852163 FitStdErr=1.6347754 Fstat=372.4653



Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

r ² Coef Det	DF Adj r ²	Fit Std Err	F-value
0.9612868555	0.9585216309	1.6347754208	372.46529658

Parm	Value	Std Error	t-value	95% Confide	nce Limits
а	0.039859551	0.001406598	28.33755021	0.036869999	0.042849103

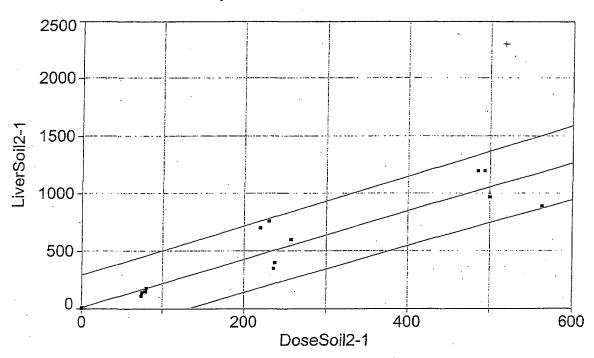
Date	Time	File Source
Feb 9, 2001	1:01:55 PM	CLIPBRD.WK1

FIGURE A-14 BEST FIT CURVE WITH 95% PREDICTION INTERVALS (generated using Table Curve 2D v. 3.0. Outliers represented by "+"

TM#2 - Liver

Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

r²=0.90065167 DF Adj r²=0.89402845 FitStdErr=133.39454 Fstat=145.04951



Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

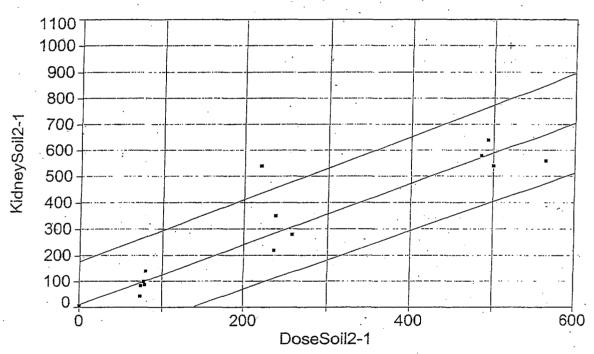
r² Coef Det DF Adi r² F-value Fit Std Err 0.9006516700 0.8940284480 133.39453976 145.04951135 95% Confidence Limits Std Error t-value 2.095646700 0.121234398 17.28590847 1.838113323 2.353180076 File Source Date Time. 9:59:40 AM CLIPBRD.WK1 Jan 15, 2001

FIGURE A-15 BEST FIT CURVE WITH 95% PREDICTION INTERVALS - (generated using Table Curve 2D v. 3.0. Outliers represented by "+"

TM#2 - Kidney

Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

 r^2 =0.89039679 DF Adj r^2 =0.88256799 FitStdErr=79.091465 Fstat=121.85731



Rank 1 Eqn 8017 [UDF 3] y=LINFORCE(a)

r² Coef Det DF Adj r² Fit Std Err F-value 0.8903967929 0.8825679924 79.091465332 121.85730922

Parm Value Std Error t-value 95% Confidence Limits a 1.160730634 0.073575623 15.77602182 1.004354686 1.317106581

Date Time File Source
Jan 15, 2001 10:01:29 AM CLIPBRD.WK1

APPENDIX B

WASHINGTON GROUP MEMORANDUM



Technical Memorandum

To:

Bonnie Lavelle

From: Kevin Williamson

REF:

RAC No. 68-W7-0039, WA. No. 004-RICO-089R

Subject: VB/I-70 Bio-Availability Soil Sample Preparation

Cc:

M. Green, B. Meyers, T. Hammonds

In support of the Vasquez Bivd./ I-70 (VB/I-70) risk assessment program, Washington Group International (WGI) was tasked with preparing soil samples for a Bio-Availability study to be performed by EPA Region VIII toxicologists. Soil samples from various residential properties were selected that represent specific concentrations of lead, and arsenic. These soil samples were collected during Phase IIIA and Phase IIIB of the remedial investigation within the neighborhoods of Swansca, Elyria, Cole, Clayton, and the southwest portion of Globeville. Bonnie Lavelle (EPA) instructed WGI to prepare two soil samples for this study representating the Western and Eastern neighborhoods. Soil samples were selected based on previously determined lead (Pb) and arsenic (As) concentrations, as determined using an EDXRF Quanx, and composited in accordance with the attached procedure (Attachment 1). On October 19, 200 twelve soil samples were selected for the East sample, and eight soil samples were selected for the West sample. Based on remaining sample weights and previously measured concentrations, six soil samples were combined to make the East sample, and five soil samples were combined to make the West sample. The following sample IDs were used to make the East sample: 3-03583-B, 3-03588-B, 3-02387-B, 3-08444-B, 3-08978-B, and 3-08979-B. Upon composite of those samples the East sample was designated 3-15621-B. The following sample IDs were used to make the West sample: 3-10740-B, 3-10318-B, 3-03910-B, 3-10734-B, and 3-10319. Upon composite of those samples the West sample was designated 3-15628-B.

On October 20, 2000, soil samples were dried in a laboratory oven at 105 C, bulk sieved with a 2-mm screen, and fine sieved with a 250-mm screen, Bulk and fine fraction samples were split for EDXRF, and TAL metals analysis via Inductively Coupled Plasma Atomic Emission Spectroscopy (ICP) analysis. Two bio-availability test substances were produced consisting of 1.2 and 1.6 kilograms of the fine fraction composite soils. Test substance samples were relinquished under chain of custody to Syracuse Research Center (SRC).

All arsenic and lead soil sample results and corresponding quality control sample results are summarized in the attached tables. Table 1 displays initial XRF screening results to insure study mandated concentration levels. Table 2 displays the bulk and fine sieved fractions also analyzed using XRF. Table 3 displays off-site laboratory

ICP results, with laboratory data sheets included as Attachment 2. Analytical results indicate that the average lead concentrations of the East test substance are 723 mg/kg (ICP) and 788 mg/kg (XRF). Average lead concentrations for the West test substance are 1050 mg/kg (ICP) and 987 mg/kg (XRF). Intra-sample variability is low with the exception of the bulk fraction ICP results.

Table 1

Location	Fraction	Sample ID	Parent	Results (mg/Kg)		ICP Result	s (mg/Kg)	Date/Time Analyzed
LOCATION	rraction		Sample ID	Arsenic	Lead	Arsenic	Lead	Cate Time Milaryzeu
West	Bulk (Screen)	3-15628-B	3-15628-B	30	908	T		10/19/2000 @ 1207
West	Bulk (Screen)	3-15628-B	3-15628-B	23	764			10/19/2000 @ 1207
West	Bulk (Screen)	3-15628-B	3-15628-B	30	823			10/19/2000 @ 1207
			Average	28	832			
East	Bulk (Screen)	3-15621-B	3-15621-B	27	639			10/19/2000 @ 1207
Ęast	Bulk (Screen)	3-15621-B	3-15621-B	10	658		,	10/19/2000 @ 1207
			Average	19	649			
		NI\$T2711		88	1150			10/19/2000 @ 1207

Table 2

	Euro (1 * - :	A	Parent	Results (m	g/Kg)	ICP Result	s (mg/Kg)	Date/Time Analyzed
Location	Fraction	Sample ID	Sample	Arsenic	Lead	Arsenic	Lead	
West	Bulk		3-15628-B	lancours a said to	983			10/19/2000 @ 1851
West	Bulk	3-15628-B2	3-15628-B		1001			10/19/2000 @ 1851
West	Bulk	3-15628-B3	3-15628-B	31	1038			10/19/2000 @ 1851
West	Bulk	3-15628-B4	3-15628-B	13	978			10/19/2000 @ 1851
West	Bulk	3-15628-B5	3-15628-B	46	960			10/19/2000 @ 1851
West	Bulk	3-15628-B6	3-15628-B	26	898		. ,	10/19/2000 @ 1851
West	Bulk	3-15628-B7	3-15628-B	22	998			10/19/2000 @ 1851
West	Bulk	3-15628-B8	3-15628-B	31	1001	1.		10/19/2000 @ 1851
West	Bulk	3-15628-B9	3-15628-B	25	857	1		10/19/2000 @ 1851
			Average	27	968	1		
West	Fine	3-15628-F1	3-15628-F	35	1062	.]		10/19/2000 @ 1851
West	Fine	3-15628-F2		27	1008			10/19/2000 @ 1851
West	Fine	3-15628-F3		27	1059			10/19/2000 @ 1851
West	Fine	3-15628-F4		19	1052			10/19/2000 @ 1851
West	Fine	3-15628-F5		32	1078			10/19/2000 @ 1851
West	Fine	3-15628-F6		41	1095			10/19/2000 @ 1851
West	Fine	3-15628-F7	3-15628-F	22	1008			10/19/2000 @ 1851
West	Fine	3-15628-F8		17	1022		.,	10/19/2000 @ 1851
West	Fine	3-15628-F9	3-15628-F	23	1062			10/19/2000 @ 1851
			Average	27	1050	<u> </u>		
		NIST2711		99	1169			10/19/2000 @ 1851
East	Bulk	3-15621-B1			722			10/19/2000 @ 1549
East	Bulk	3-15621-B2	or service and the service of	26	721	<u> </u>		10/19/2000 @ 1549
East	Bulk	3-15621-B3		26	685	[10/19/2000 @ 1549
East	Bulk	3-15621-B4		24	746			10/19/2000 @ 1549
East	Bulk	3-15621-B5		25	746	<u> </u>		10/19/2000 @ 1549
East	Bulk	3-15621-B6		21	764	<u> </u>		10/19/2000 @ 1549
East	Bulk	3-15621-B7		. 27	840			10/19/2000 @ 1549
East	Bulk	3-15621-B8		29	702			10/19/2000 @ 1549
East	Bulk	3-15621-B9		10	694]		10/19/2000 @ 1549
	<u>.</u>	0 40004	Average	24	736			
East	Fine	3-15621-F1		9	839			10/19/2000 @ 1549
East	Fine	3-15621-F2		. 17	780			10/19/2000 @ 1549
East	Fine	3-15621-F3		14	774			10/19/2000 @ 1549
East	Fine	3-15621-F4		18	765			10/19/2000 @ 1549
East	Fine	3-15621-F5		17	770			10/19/2000 @ 1549
East	Fine	3-15621-F6		16	750	<u> </u>		10/19/2000 @ 1549
East	Fine	3-15621-F7		33	827	<u> </u>		10/19/2000 @ 1549
East	Fine	3-15621-F8		22	804			10/19/2000 @ 1549
East	Fine	3-15621-F9		16	7.87			10/19/2000 @ 1549
ļ <u>-</u>			Average	18	788			,
		NIST2711	<u></u>	96	1168			10/19/2000 @ 1549

Table 3

Location	Fraction	Sample	Parent	Results (m	g/Kg)	ICP Resul	ts (mg/Kg)	Date/Time Analyzed
Location	Fraction	ID Sample		Arsenic	Lead	Arsenic	Lead	Date/ Time Analyzou
West	Bulk	3-15703-B	3-15628-B			10	900	10/23/00
West	Bulk	3-15704-B	3-15628-B			9	370	10/23/00
West	Bulk	3-15705-B	3-15628-B			10	400	10/23/00
					Average	10	557	
West	Fine	3-15700-F	3-15628-F	<u></u>	_	26	970	10/23/00
West	Fine	3-15701-F	3-15628-F		1	25	1000	10/23/00
West	Fine	3-15702-F	3-15628-F			24	990	10/23/00
					Average	25	987	
East	Bulk	3-15622-B	3-15621-B			10	610	10/23/00
Éast	Bulk	3-15623-B	3-15621-B			9	1100	10/23/00
East	Bulk	3-15624-B	3-15621-B			10	620	10/23/00
					Average	10	777	
East	Fine	3-15627-F	3-15621-F			19	700	10/23/00
East	Fine	3-15629-F	3-15621-F			19	710	10/23/00
East	Fine	3-15630-F	3-15621-F			20	760	10/23/00
					Average	19	723	

ATTACHMENT 1 BIO-AVAILABILITY SAMPLE PREPARATION PROCEDURE

BIO-AVAILABILITY SOIL SAMPLE PREPARATION PROCEDURE OCTOBER 2000

RE: RAC No. 68-W7-0039, WA No. 004-RICO-089R (VB/1-70 OU1)

OBJECTIVES:

- 1. Produce two fine sieved, composite soil samples for use EPA's use in a pig study containing relatively high levels of lead and low levels of arsenic. If possible, one sample should be representative of COLE (and FIVE POINTS) neighborhood soils and one representative of CLAYTON (and SWANSEA) soils.
- 2. Measure concentrations and variability of both the fine fraction and bulk fraction of the composite using both XRF and ICP.

CONTACTS:

Bonnie Lavelle 303/312-6579, 303/898-8465

Chris Weis

303/312-6671, 720/320-6254 (cell), 800/759-8888 #1083306 (pager)

Tracy Hammond, SRC 303/713-9549

PROCEDURE

- I. Identify Samples for Compositing
- 1. Evaluate list of candidate samples and minimum quantity provided by Tracy along with neighborhood data. Group by 2 neighborhoods WEST and EAST (and other may not want to include Globeville or Elyria for this study). Calculate average concentration consult Bonnie if concentration is too low based on Tracy's mass requirements (target 1100-1200 ppm lead); or eliminate samples from consideration that bring the average down.
- 2. Note that mass of each composite needed is for Test Substance per Tracy's table, PLUS 50 grams, plus 18 XRF samples, plus 6 ICP samples, and plus some if possible for archive.
- 3. Retrieve samples from archive for each group.
- 4. Weigh raw samples; Re-evaluate relative contribution (composite design) of samples (maximize volume of higher Pb concentration samples) in order to achieve target/high Pb concentration in composite. If mass is inadequate to generate WEST and EAST composites, consult EPA for guidance on alternative approach targeting 2 different concentration levels.
- II. Prepare Composite
- 1. Weigh out portions/all of the raw samples per composite design and composite into the two Raw samples. Label each uniquely 3-XXXXX-R. Homogenize very well,

- 2. Thoroughly dry each raw sample. Bulk sieve each entire composite; Label with sample IDs (-B).
- 3. Prep one XRF cup for each -B and screen by XRF confirm adequate sample mass for each composite. If mass is low, dry and bulk sieve additional soil (from appropriate EAST or WEST group), re-homogenize, and re-screen -B.
- 4. Reserve -B sample quantities adequate for 9 XRF samples and 3 ICP samples.
- 5. Fine sieve (60 mesh) remaining -B sample mass to produce the two composite -F samples.

III. Split Samples - Bulk

- 1. Split each -B composite into 3 uniquely identified samples and ship for ICP analysis of all EPA Target Analyte List metals (std or 48-hour turnaround **BONNIE: may be easier to submit all for quick turnaround to enable completion of this task next week, rather than get a separate data package in 3+ weeks).
- 2. Grind / cup 9 aliquots (original sample ID 3-XXXXX-B1 --> B9 *** BONNIE: does this have to be blind to the XRF analyst? If so, assign unique sample IDs) for XRF analysis
- IV. Split Samples Fine
- 1. Split each -F composite into 3 uniquely identified samples and ship for ICP analysis of all EPA Target Analyte List metals (48-hour turnaround)
- 2. Grind / cup 9 aliquots for each (original sample ID 3-XXXXX-F1 --> F9 *** BONNIE: does this have to be blind to the XRF analyst? If so, assign unique sample IDs) for XRF analysis
- 3. Split each remaining -F composite into a 50-gram sample (for physical testing), a required mass of Test Substance, and any remaining quantity for archive; ALL with same original -F sample ID.
- 4. Transfer Sample consisting of 50-gram and Test Substance portions to Tracy Hammond (phone her for a pick-up) under COC.
- VI. XRF Analysis / Reporting
- 1. Analyze 9 B and 9 -F samples from each composite together along with NIST standard and blank.

VII. Documentation

- 1. Produce a Technical Memorandum summarizing task, include: prep log/XRF instrument log/COCs/ICP data received; three copies to Bonnie.
- FAX ICP data to Bonnie, Chris and Tracy upon receipt.

BIO-AVAILABILITY SOIL SAMPLE PREPARATION PROCEDURE OCTOBER 2000

RE: RAC No. 68-W7-0039, WA No. 004-RICO-089R (VB/I-70 OUI)

OBJECTIVES:

- 1. Produce two fine sieved, composite soil samples for use EPA's use in a pig study containing relatively high levels of lead and low levels of arsenic. If possible, one sample should be representative of COLE (and FIVE POINTS) neighborhood soils and one representative of CLAYTON (and SWANSEA) soils.
- 2. Measure concentrations and variability of both the fine fraction and bulk fraction of the composite using both XRF and ICP.

CONTACTS:

Bonnie Lavelle 303/312-6579, 303/898-8465

Chris Weis

303/312-6671, 720/320-6254 (ceil), 800/759-8888 #1083306 (pager)

Tracy Hammond, SRC 303/713-9549

PROCEDURE

- I. Identify Samples for Compositing
- 1. Evaluate list of candidate samples and minimum quantity provided by Tracy along with neighborhood data. Group by 2 neighborhoods WEST and EAST (and other may not want to include Globeville or Elyria for this study). Calculate average concentration consult Bonnie if concentration is too low based on Tracy's mass requirements (target 1100-1200 ppm lead); or eliminate samples from consideration that bring the average down.
- Note that mass of each composite needed is for Test Substance per Tracy's table, PLUS grams, plus 18 XRF samples, plus 6 ICP samples, and plus some if possible for archive.
- 3. Retrieve samples from archive for each group.
- 4. Weigh raw samples; Re-evaluate relative contribution (composite design) of samples (maximize volume of higher Pb concentration samples) in order to achieve target/high Pb concentration in composite. If mass is inadequate to generate WEST and EAST composites, consult EPA for guidance on alternative approach targeting 2 different concentration levels.
- II Prepare Composite

-

1. Weigh out portions/all of the raw samples per composite design and composite into the two Raw samples. Label each uniquely 3-XXXXX-R. Homogenize very well.

- 2. Thoroughly dry each raw sample. Bulk sieve each entire composite; Label with sample IDs (-B).
- 3. Prep one XRF cup for each -B and screen by XRF confirm adequate sample mass for each composite. If mass is low, dry and bulk sieve additional soil (from appropriate EAST or WEST group), re-homogenize, and re-screen -B.
- 4. Reserve -B sample quantities adequate for 9 XRF samples and 3 ICP samples.
- 5. Fine sieve (60 mesh) remaining -B sample mass to produce the two composite -F samples.
- III. Split Samples Bulk
- 1. Split each -B composite into 3 uniquely identified samples and ship for ICP analysis of all EPA Target Analyte List metals (std or 48-hour turnaround **BONNIE: may be easier to submit all for quick turnaround to enable completion of this task next week, rather than get a separate data package in 3+ weeks).
- 2. Grind / cup 9 aliquots (original sample ID 3-XXXXX-B1 --> B9 *** BONNIE: does this have to be blind to the XRF analyst? If so, assign unique sample IDs) for XRF analysis
- IV. Split Samples Fine
- 1. Split each -F composite into 3 uniquely identified samples and ship for ICP analysis of all EPA Target Analyte List metals (48-hour turnaround)
- 2. Grind / cup 9 aliquots for each (original sample ID 3-XXXXX-F1 --> F9 *** BONNIE: does this have to be blind to the XRF analyst? If so, assign unique sample IDs) for XRF analysis
- 3. Split each remaining -F composite into a 50-gram sample (for physical testing), a required mass of Test Substance, and any remaining quantity for archive; ALL with same original -F sample ID.
- 4. Transfer Sample consisting of 50-gram and Test Substance portions to Tracy Hammond (phone her for a pick-up) under COC.
- VI. XRF Analysis / Reporting
- 1. Analyze 9 B and 9 -F samples from each composite together along with NIST standard and blank.
- VII. Documentation
- 1. Produce a Technical Memorandum summarizing task, include: prep log/XRF instrument log/COCs/ICP data received; three copies to Bonnie.
- FAX ICP data to Bonnie, Chris and Tracy upon receipt.

ATTACHMENT 2

LABORATORY DATA SHEETS FOR TARGET ANALYTE LIST METALS BY ICP

Method SW6010 Sample Results

PRELIMINARY RESULTE

Lab Name: Paragon Analytics, Inc.

Work Order Number: 0010156

Client Name: Monison Knudsen Corporation

ClientProject ID: V8/I-70 IIIB 4994

ாய்ப் ID; 3-16622-8 `ட்ஷ் |்ற்; 0010156-1 Sample Matrix: SO/L

% Moisture: 0.2

Date Collegier: 19-Oct-06

Date Collected: 19-Oct-00 Date Extracted: 23-Oct-00 Date Analyzed: 23-Oct-00 Prep Batch: IP001023-1 GCBatchi0: IP001023-1-1

Run ID: IT001023-1A4 Clèanup: NONE Basis: Dry Weight Sample Allquot: 1 G Final Volume: 100 ML Result Units: MG/KG

File Name: TS01023

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	MDL	Result Qualifier	EPA Qualifier
7429-00-5	ALUMINUM	1	2900	20	0.69		
7440-36-0	ANTIMONY	1	1.6	2	0,26	В	
7440-38-2	ARSENIC	1	96	1)	0.28		
7440-39-3	BARIUM	1	130	10	0.018	· · · · · · · · · · · · · · · · · · ·	
7440-41-7	BERYLLIUM	1	0.33	0,5	0.015	В	
74-10-43-9	CADMUM	1	1.8	0,5	0.017		-
7440-70-2	CALCIUM	1	3400	100	0.54		
7410-47-3	CHROMIUM	1	15	1	0.047		
7440-48-4	COBALT	1	3.1	1	0.05		
7440-50-8	COPPER	1	35	1	0.032		
7439-89-6	IRON	1	15000	10	0.8		
7439-92-1	LEAD	1	610	03	0,14	.,	
7430-99-4	MACHESIUM	^	1100	100	0.79		
7-130-95-5	MANGANESE	1	230	1	0.025		,
7440-02-0	NICKEL	1	0,5	2	0,078		
74-10-09-7	POTAESIUM	1	840	100	5.3		
7762-49-2	SELENIUM	1	1	0.5	0 27		
7440-22-4	SILVER	1	1	1	0.063	U	
7440-23-5	SOOIUM	1	140	100	0.25		
7440-28-0	THALLIUM	1	1	1	0,39	U	""",
7440-62-2	VANADIUM	1	9.9	1	0.033		
7440-66-6	ZINC	1	300	2	0.29		

Method SW6010 Sample Results

PRELIMINARY RESULTS

Lab Name: Paragon Analytics, Inc.

Work Order Number: 0010156

Client Name: Morrison Knudsen Corporation

ClientProject ID: V8/1-70 IIIB 4994

Fjeld JD123-15623-8 Lab JD2 (0010156-2 Sample Matrix; SOJI, % Moisture: 0.2

Date Collected: 19-Oct-00 Date Extracted: 23-Oct-00

Date Analyzed: 23-Oct-00

Prep Batch: IP001023-1-1 QCBatch(D: IP001023-1-1

Run ID: IT001023-1A4 Cleanup: NONE Basis: Dry Welght Sample Aliquot: 1 G
Final Volume: 100 ML
Result Units: MG/KG

File Name: TS01023

CASNO	Tärget Analyte	Dilution Factor	Result	Reporting Limit	MDL	Result Qualifier	EPA Qualifier
7429-90-5	ALUMINUM	1	2500	20	0.69		
7440-36-0	ANTIMONY	1	1.2	2	0.26	В	
7440-58-2	ARSENIC	1	8.7	1	0.28		
7440-39-3	BARIUM	1 :	760	10	0.018		
7440 41-7	BERYLLIUM	1 !	0,28	0,5	0.015	В	
7440-43-9	CADMIUM	1 !	22	0,5	0.017		
74-10-70-2	CALCIUM	1	4000	100	0.54		
7440-47-3	CHRONIUM	1	7,1	1	0.047		
7440-48-4	COBALT	1	2.6	1	0.05		···
74-10-50 8	COPPER	1	24,	1	0,032		
7439-89-6	IRON	1	10000	10	0.6		
7439-92-1	LEAD	10	1100	3	1.4		<u></u>
7439-95-4	MAGNESIUM	7	1200	100	0.79		**
7439-96-5	MANGANESE	1	260	1	0.025		
7410-02-0	NICKEL	1	4.5	2	0.078		
7440-09-7	POTASSIUM		780	100	5.3		
7782-49-2	SELENIUM	-	0,88	0.5	0,27		
7440-22-4	SILVER		0.52	1	630.0	В	
7440-23-5	SODIUM	1	140	100	0.25		
440-28-D	THALLIUM	1	1	1	0.39	U	· · · · · · · · · · · · · · · · · · ·
4-10-62-2	VANADIUM		9.8	1	0.033		
440 66-6	ZINC		540	2	0,29		

Method SW6010 Sample Results

PRELIMINARY RESULTS

Lab Name: Paragon Analytics, Inc.

Work Order Number: 0010156

Client Name: Morrison Knudsen Corporation

ClientProject ID: VB/I-70 IIIB 4994

Fight ID: 3-15624-8 - Lab IO(L) 0010155-3 Sample Matrix; SOIL % Moisture: 0.5

Date Collected: 19-Oct-00

Date Extracted: 23-Oct-00 Date Analyzed: 23-Oct-00 Prep Batch: (P001023-1 QCBatchID: (P001023-1-1

Run ID: IT001023-1A4

Cleanup: NONE Basis; Dry Weight Sample Afiquat: 1 G Final Volume: 100 ML

Result Units: MG/KG

File Name: TS01023

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	MDL	Rosult Qualifier	EPA Qualitier
7429-90-5	ALUMINUM	1	2800	20	0,69		
7410-38-0	ANTIMONY	1	0,67	2	0,27	В	
7440-30-2	ARSENIC	1	9.5	1	0.28		
7440-39-3	BARIUM	1	130	10	0.018		
7440-41-7	BERYLLIUM	1	0.34	0.5	0.015	В	
7440-43-9	CADMIUM	7	2.2	0,5	0.017		
7440-70-2	CAL CRUM	1	4100	100	0.54		
7440-47-3	CHROMIUM	1	7.3	1	0.048		
7440-48-4	COBALT	1	2.9	1	0.05		
7440-50-8	COPPER	1	21	1	0.032		
7439.69.5	IRON	1	7700	10	0.8		
7439-02-1	LEAD	1	620	0.3	0,14		
7439-95-4	MAGNESKIM	1	1200	100	0.79		
7459-86-6	MANGANESE	1	150	t	0.025	······································	
7440-02-0	NICKEL	7	52	2	0.078		
7440-09-7	POTASSIUM	1	920	100	5.3	• • • • •	
7782-49-2	SELENIUM	1	0.38	0.5	0.27	В	
7440-22-4	SILVER	1	0.16	1	0.063	B	
7440-23-5	SODIUM	5	160	100	0.25	<u>-</u>	
7410-28-0	THALLIUM	1	1:	1	0.39	U	·
7-140-02-2	VANADIUM		10	1	0.033		
7440-65-6	ZINC	1	300	2	0,29		

Method SW6010 Sample Results



Lab Name: Paragon Analytics, Inc.

Work Order Number: 0010156

Client Name: Mordson Knudsen Corporation

ClientProject ID: VB/I-70 IIIB 4994

Field ID: 2-15627-F Lab ID: 50010156-4 Sample Matrix: SOIL

% Moisture: 0.5

Date Collected: 19-Oct-00

Date Extracted: 23-Oct 00 Date Analyzed: 23-Oct-00 Prep Batch: 1P001023-1

QCBatchID: IP001023-1-1 Run ID: IT001023-1A4

Cleanup: NONE
Basis: Dry Weight

Sample Aliquot;

1 G 100 ML

Final Volume: 100 Result Units: MG/KG

File Name: TS01023

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	MDL	Regult Qualifier	EPA Qualifier
7429-90-5	ALUMINUM	1	7000	20	0 69		
7440-36-0	ANTIMONY	1	1.6	2	0.27	8	
7440-36-2	ARSENIC	1	19	1	0.28		
7440-39-3	BARIUM	1	290	10	0.018		
7440-41-7	BERYLLIUM	1	0,71	0.5	0.015		
74-10-43-9	CADMIUM	1	5.4	0.5	0.017		
7440-70-2	CALCIUM	1	6500	100	0.54		
7440-47-3	CHROMIUM	1	21	1	0.048	*****	
7440-48-4	COBALT	1	6.4	1	0,05		
7440-50 S	COPPER	1	71	1	0.032		
7430-80-6	IRON	1	16000	10	6,8	*	· · · · · · · · · · · · · · · · · · ·
7439-92-1	LEAD	1	700	0.3	0.14		
7439-95-4	MAGNESIUM	1	2400	100	0.79		
7439-36-5	MANGANESE	1	380	1	0.025		*****
7440 02-0	NICKEL	1	12	2	0.078		
7440-09-7	POTASSIUM	1	2500	100	5,3		****
7732-49-2	SELENIUM	1	1	0,5	0.27		
7440-22-4	SILVER	1	0 68	1	0.063	В	
440-23-5	SODIUM		260	100	0.25		
/440 28 0	THALLIUM	1	1 ₁	1	0.39	U	
140-62-2	VANADIUM	1	24	1	0.033		
440-56-6	ZING	1	620	2	0.29		·

Method SW6010 Sample Results

PRELIMINARY RESULTS

Sample Aliquot:

Lab Name: Paragon Analytics, Inc.

Work Order Number: 0010156

Client Name: Morrison Knudsen Corporation

ClientProject ID: VB/I-70 III8 4994

Field (D): 3-15629-F Lab (D): 0010156-5 Somple Matrix: SOIL % Moisture: 0.4

Date Collected: 19-Oct-00

Date Extracted: 23-Oct-00 Date Analyzed: 23-Oct-00 Prep Batch: IP001023-1 QCBatchID: IP001023-1-1

Run ID: IT001023-1A4 Cleanup: NONE Basis: Dry Weight Final Volume: 100 ML Result Units: MG/KG

File Name: TS01023

1 G

CASNO	Yarget Analyte	Dilution Factor	Result	Reporting Limit	MDL	Result Qualifier	EPA Qualifier
7423-00-5	ALUMINUM	1	7500	20	0,69		
7440-36-0	MIMONY	1	1,6	2	0.27	8	
7440-38-2	ARSENIC	1	19	1	0.28		
7440 39-3	BARIUM	1	310	10	0,018		
7440-41-7	BERYLUUM	1	0 73	0.5	0.015		
7440-43-9	CADMILM	1	5.5	0,5	9.017		
7440-70-2	CALCIUM	1	6600	100	0.54		
7440 47-3	CHROMIUM		21	1	0.048		
7440-48-4	COBALY	1	6.5	1	0.05	-	
7-1-10-50-8	COPPER	1	63	1	0,032		
7439-89-6	IRON	1	17000	10	0.8		
7439 92-1	LEAD	1	710	0.3	0.14		
7439-05-4	MACNESIUM	1	2500	100	0.79		······································
7439-96-5	MANGANESE	1	400	1	0.025		
7440 02-0	NICKEL	1	12	2	0,078		
7440 09-7	POTASSIUM	1	2700	100	5.3		
7782-49-2	SELENIUM	1	1.2	0.5	0.27	··· · · · ·	
7440-22-4	SILVER	1	0,69	1	0.063	В	
7440-23-5	SODIUM	1	270	100	0.25		
7440-28-0	THALLIUM	1	1	1	0.39	- u	
7-140-52-2	VANADIUM	1	24	1	0.033	_	
7440-66-6	ZINC	1	520	2	0.29		

Method SW6010 Sample Results

PRELIMINARY RESULTS

Lab Name: Paragon Analytics, Inc.

Work Order Number: 0010156

Client Name: Morrison Knudsen Corporation

ClientProject ID: VB/I-70 IIIB 4994

Field ID: 3-15630 F

Sample Matrix: SOIL

% Moisture: 0.2

Date Collected: 19-Oct-00

Date Extracted: 23-Oct-00 Date Analyzed: 23-Oct-00 Prep Batch: IP001023-1

QCBatchib: IP001023-1-1 Run ID: IT001023-1A4

Cleanup: NONE

Basis: Dry Weight

Sample Aliquet:

1 G 100 ML

Final Volume: 100 Result Units: MG/KG

File Name: TS01023

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	MDL	Result Qualitier	EPA Qualifier
7429-90-5	ALUMINUM	1	8000	20	0.69		
7440-36-0	ANTIMONY	1	1.8	2	0,26	В	
7440-38-2	ARSENIC	1	20	1	0.28		
7440.39.3	BARIUM	i	310	10	0.018		
7440-41-7	BERYLLIUM	1	0.77	0,5	0.015		·
7440-43-9	CADMIUM	1	5.7	0.5	0.017		
7440-70-2	CALCIUM	1	7000	100	0.54		
74-10-47-3	CHROMIUM	1	21	1	0.047		
7440-45-4	COBALT	1	6.9	1	0.05	717-74	
7440-50 8	COPPER	1	64	1	0.032		Y MARIA LA
7430-89-6	IRON	1	18000	10	0.8		
7439-92-1	LKAD	1	760	0.3	0,14		
7439 95-4	MAGNESIUM	7	2700	100	0,79		
7439-90-5	MANGANESE	i	410	1	0.025		ENAME TO A STREET
7440-02-0	NICKEL	1	13	2	0.078		
7440-09-7	POTASSIUM	1	2900	100	5.3		
7792-49-2	SELENIUM	1	0.97	0.5	0.27		
7440-22-4	SILVER	1	0.77	1	0,083	B B	
7-1-10-23-5	SODIUM	1	300	100	0.25		
7440-28-0	THALLIUM	1		1	0.39	IJ	•
7440 62-2	VANALIUM	1	26	1	0.033		
7440-68-6	ZINC		680	2	0.29		

Method SW6010 Sample Results

PRELIMINARY RESULTS

Lab Name: Paragon Analytics, Inc.

Work Order Number: 0010156

Client Name: Morrison Knudsen Corporation

ClientProject ID: V8/I-70 IIIB 4994

Fig.fr (D: : 3-15703-B Lab ID: 0010156-7 Sample Matrix: SOIL % Moisture: 0.2

Date Collected: 19-Oct-00

Date Extracted: 23-Oct-00 Date Analyzed: 23-Oct-00 Prep Baich: 1P001023-1 QCBatchiD: IP001023-1-1

Run ID: IT001023-1A4 Cleanup: NONE

Basis: Dry Weight

Sample Aliquot: Final Volume:

1 G 100 ML

Result Units: MG/KG

File Name: TS01023

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	MDL	Result Qualifier	EPA Qualifier
7429-90-5	ALUMINUM	1	2600	20	D,69		
7440-36-0	ANTIMONY	ı	1.5	2	0.26	8	
7440-38-2	ARSENIC	1	B,C	1	0.28		
7440-33-3	BARIUM	1	160	10	0.018	· · · · · · · · · · · · · · · · · · ·	· · · · · · · · · · · · · · · · · · ·
7-140-41-7	BERYLLIUM	1,	0.31	0.5	0.015	8	
7440-43-9	CADMIUM	1	2,1	0.5	0.017		
7440-70-2	CALCIUM	1	3000	100	0.54		
7440-47-3	CHROMIUM	7	9.1	1	0.047		
7440-48-1	COBALT	1	2.9	1	0.05		
7440-50-8	COPPER	7	22	1	0.032		
7439 69 6	IRON	1	0083	10	0.8		
7439 92-1	LEAD	1	900	0.3	0.14		
7430-95-4	MAGNESIUM	1	790	100	0.75	1	
7459 96 5	MANGANESE	1	270	1	0.025		
7440-02-0	NICKEL	1	4,4,	2	0.076		
7-140-09-7	POTASSIUM	1	860	100	5,3		
7782-40-2	SELENIUM	1	0.39	0.5	0 27	В	-
7440-22-4	SILVER	1	0.17	1	0 063	В	
744D-23-5	SODIUM	1	120	100	0.25		
7440-28-0	THALLIUM	1	1	1	0.39	ν	•
7440-63-2	VANADIUM	1	11	1	0.033		
7440-66-6	ZINC		400	2	0,29		

Method SW6010 Sample Results

PRELIMINARY RESULTS

Lab Name: Paragon Analytics, Inc.

Work Order Number: 0010156

Client Name: Morrison Knudsen Corporation

ClientProject ID: V8/1-70 III8 4994

Field 10; 13-15704-8 CAB ID: 0000156-8 Sample Matrix: SOIL % Moisture: 0.3

Dale Collected: 19-Oct-00

Date Extracted: 23-Oct-00 Date Analyzed: 23-Oct-00 Prop Batch: IP001023-1 QCBatchip: IP001023-1-1

Run ID: IT001023-1A4 Cleanup: NONE

Basis: Dry Weight

Sample Aliquot:

1 G 100 ML

Final Volume: Result Units: MG/KG

File Name: TS01023

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	MDL	Result Qualifier	EPA Qualifies
7429-90-5	ALUMINUM	1	2200	20	0.69		
7440-36-0	ANTIMONY	1	0.67	2	0,26	В	THE PERSON AND ADDRESS OF THE PERSON ADDRESS OF THE PERSON AND ADDRESS OF THE PERSON AND ADDRESS OF THE PERSON ADDRESS OF THE PERSON AND ADDRESS OF THE PERSON AND ADDRESS OF THE PERSON ADDRESS OF THE PERSON ADDRESS OF THE PERSON ADDRESS OF THE PERSON AND ADDRESS OF THE PERSON AND ADDRESS OF THE PERSON ADDRESS
74-10-38-2	ARSENIC	1	D.1	1	0.28		
7440-39-3	DARIUM	1	100	10	0.018	<u> </u>	
7-440-41-7	BERYLLIUM	1	0,3	0.5	0,015	В	
7440 43 9	CADMIUM	1	1.8	0,5	0.017		
7440-70-2	CALCIUM	1	2900	100	0,54		
7440-47-3	CHROMIUM	1	7.9	1	0.048		777
7440-48 4	COBALT	1	2.3	1	0.05		
7440-50-8	COPPER	1	20	1	0.032		
7430-90-6	IRON	1	6400	10	8.0		
7430-52-1	LEAD	1	370	E,0	0.14		TERROR !
7459-95-4	MAGNESIUM	7	750	100	0.79		·
7450-96-5	MANGANESE	1	190	1	0.025		
7440-02-0	NICKEL	1	3.7	2	0,078		
7440-09-7	POTASSIUM	1	750	100	5,3		
7782-49-2	SELENIUM	1	0.5	0.5	0.27	u	
7440-22-4	SILVER	1	0.17	1	0.063	В	
7440-23-5	SOOIUM	1	76	100	0.25	В	.
7440-28-0	THALLIUM	771-		1	0.39	U	-
7440-62-2	VANADIUM	1	8.2	1	0,033		
7440-66-6	ZINC	1	240	2	0.29		

Method SW6010 Sample Results

PRELIMINARY RESULTS

Lab Name: Paragon Analytics, Inc.

Work Order Number: 0010155

Client Name: Morrison Knudsen Corporation

ClientFroject ID: VB/I-70 IIIB 4994

. Field | D. : 3-15705-8 - Lab | D. : 70010156-9 Sample Matrix: SOIL % Moisture: 0.1

Date Collected: 19-Oct-00 Date Extracted: 23-Oct-00

Date Extracted: 23-Oct-00
Date Analyzed: 23-Oct-00

Prep Batch: IP001023-1 QCBatchID: IP001023-1-1

Run ID: IT001023-1-1 Cleanur: NONE

Cleanup: NONE
Basis: Dry Weight

Sample Aliquot: Final Volume;

1 G 100 ML

Result Units: MG/KG

File Name: TS01023

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	MOL	Re∉ult Qualifier	EPA Qualifier
7429-00-5	ALUMINUM	1	2500	20	0.69		
7440-36 0	ANTIMONY	1	1	2	0,26	8	
7440-38-2	ARSENIC	1	9.8	1	0.28		
7440-39 3	BARIUM	1	130	10	0.018		
74-10-41-7	BERYLLIUM	1	0.31	0,5	0.015	8	
7440-43-9	CADMIUM	1	1.9	0,5	0.017		
7440-70-2	CALCIUM	1	3100	100	0.53		
7440.47-3	CHROMIUM	1	9.7	í	0.047		
7440-18-4	COBALT	1	4	1	0.05		
7440-50-8	COPPER	1	28	4	0,032	•	
7439-89 6	IRON		8000	10	0.8		
7439-92-1	LEAD	1	400	0.3	0.14		75
7439-95-4	MAGNESIUM	1	780	100	0.79	***************************************	
7430 06-5	MANGANESE	1	260	1	0.025	•	
7440-02-0	NICKEL	ī	1.2	2	0.078		· · · · · · · · · · · · · · · · · · ·
7440 09 7	POTASSIUM	1	950	100	5.3		
7782-49-2	SELENIUM	7	0.31	0,5	0.27	B)
7440 22-4	SILVER	1	0.38	1	0.063	B	
7440-23-5	NUIDOS	1	88	100	0.25	В	
7-1-10-28-0	THALLIUM	7	1	1	0,39	-0	
7440-62-2	VANACIUM	1	93	1	0,033		
7.440-66-6	ZING	1	320	2	0.29		

Method SW6010 Sample Results

PRELIMINARY RESULTS

Lab Name: Paragon Analytics, Inc.

Work Order Number: 0010156

Client Name: Morrison Knudsen Corporation

ClientProject ID: VB/I-70 IIIB 4994

Field (0) 3-15700-F

Sample Matrix: SOIL
'S Moisture: 0.3

Date Collected: 19-Oct-00 Date Extracted: 23-Oct-00

Date Analyzed: 23-Oct-00

Prep Batch: IP001023-1

QCBatchID: 19001023-1-1 Run ID: (T001023-1A4 Cleanup: NONE

Hasis: Dry Weight

Sample Aliquot:

1 G 100 ML

Final Volume: 100 | Result Units: MG/KG

File Name: T\$01023

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	MDL	Result Qualifier	EPA Qualifier
7429-80-5	ALUMINUM	1	5900	20	0.69		
7440-36-0	ANTIMONY	1	2.3	2	0.26		
7440-33-2	ARSENIC	. 1	25	1	0 28		·
7440-39 3	BARIUM	1	200	10	810.0		-
7440-41-7	BERYLUM	1	0.66	5.5	0.015		
7440-43 9	CACINIUM	1	4.2	0.5	0.017		
7440-70-2	CALCIUM	1	6200	100	0,54		
7440-47-3	CHROMIUM	7.	21	1	0.048		· · · · ·
7440-48 4	COBALT	1	5.5	1	0.05		
7440-50-8	OOPPER		54	1	0.032		
7439 89 6	IRON	1	16000	10	0,8		
7439-92-1	LEAD	1	970	0.3	0,14		
7430 95-4	MAGNESIUM	3	1800	100	0.79		
7439-96-5	MANGANESE	1	410	1	0,025		
7440-02-0	NICKEL	7	9,5	2	0.078		
7410 09 7	POTASSIUM	1	2000	100	5.3		
7782-49-2	SELENIUM	1	1	0.5	0.27		
7440-22-4	SILVER	1	0.81	1	0.063	8	
140-23-5	SOCIUM	1	160	100	0.25		
440-28-0	T'HALLIUM		1	1	0.39	U V	
440-62-2	VANADIUM		23	1	0.033	-	
140-56-6	ZINC		540	2	0.29		······································

Method SW6010 Sample Results



Lab Name: Paragon Analytics, Inc.

Work Order Number: 5010156

Client Name: Morrison Knudsen Corporation

ClientProject ID: VII/I-70 IIIB 4994

Field in: 3-15701-F Lub in: 20010156-11 Sample Matrix: SOIL

% Moisture: 0.2

Date Collected: 19-Oct-00 Date Extracted: 23-Oct-00

Date Analyzed: 23-Oct-00

Prep Batch: IP001023-1

QCBatchID: IP001023-1-1 Run ID: IT001023-1A4

Cleanup: NONE Basis: Dry Weight Sample Aliquot:

1 G 100 ML

Final Volume: 100 Result Units: MG/KG

File Name: TS01023

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	MDL	Result Qualifier	EPA Qualifier
7429-90-5	ALUMINUM	1	6100	20	0.69		-
7440-35-0	ANTIMONY	1	2	2	0.26	В	N
7-140-38-2	ARSENIC		25	1	0.28		
7440-39-3	BARIUM	, 1	300	10	0,018		
7440 41-7	BERYLLIUM	1 ;	0 67	0.5	0.015		
7440-43-9	CADMIUM	7 1	4.3	0.5	0,017		
7440-70-2	CALCIUM	1	6200	100	0.54		
7440-47-3	CHROMIUM	7	21	1	0,047		
7440-48-4	COBALT	ī	5.5	1	0.05		······································
7410-50-8	COPPER	1	54	1	0 032		
7439-69-6	IRON		16000	10	8.0		
7439-92-1	LEAD	10	1000	3	1.4		-
7439-95-4	MAGNESIUM	i	1800	100	0.79		
7439 96-5	MANGANESE	1	430	1	0.025		
7440-03-0	NICKEL	1	9.5	2	0.078		
7440-09-7	POTASSIUM	1	2000	100	5.3		
7/52-49-2	SELENIUM	1	1.1	0.5	0 27		
7440-22-4	SILVER	1	0.77	1	0.063	В	
7440-23-5	MUIGOS	1	160	100	0.25		E
7440-25-0	THALLIUM	1		1	0,39	υ	
7440-62-2	VANADIUM		23	1	0.033		
7440-66-G	ZINO	1	550	2	0.29		

Method SW6010 Sample Results



Lab Name: Paragon Analytics, Inc.

Work Order Number: 0010156

Client Name: Morrison Knudsen Corporation

CliantProject ID: VB/1-70 IIIB 4994

Field ID: 2-15702-I: Lub ID: 0010156-12 Sample Matrix: SOIL

% Moisture: 0.3

Date Collected: 19-Oct-00

Date Extracted; 23-Oct-00

Date Analyzed: 23-Oct-00

Prop Batch: IP001023-1

QCBatchID: IP001023-1-1 Run ID: IT001023-1A4

Cleanup: NONE
Basis: Dry Weight

Sample Aliquot:

16

Final Volume: 100 ML Result Units: MG/KG

File Name: TS01023

CASNO	Target Analyte	Dilution Factor	Result	Reporting Limit	MDL	Result Qualifler	EPA Qualifier
7429-90-8	ALUMINUM	1	6500	20	0.69		
7410-35-0	YVOMITAL	7	0.91	2	0.26	8	
7440-38-2	ARSENIC	1	24	1	0.28		
7-140-39-3	BARIUM	7	290	10	0,018		
7440-41-7	BERYLLIUM	1	0,61	0.5	0.015		
7440 43-9	CADMIUM	T	4.2	5.5	0.017		
7440-70-2	CALCIUM	1	6100	100	0.54		
7440-47-3	CHROMIUM	1	18	1	0.048		
7440-18-1	CODALT	1	4.6	1	0.05		
7440-50-8	COPPER	7	50	1	0,032		
7430-60-6	IRON	1	11000	10	0.8		The Assessment of the Control of the
7439 92-1	LEAD	1	990	0,3	0.14		<u></u>
7439 95-4	MAGNESIUM	1	1700	100	0.79		
7439-96-5	MANGANESE	1	400	1	0.025	7,7,000	
7440-02-0	NICKEL	1	84	5	0.078	4.	
7440-09-7	POTASSIUM	1	2100	100	5.3		
7762-48-2	SELENIUM	1	1.2	0.5	0.27		
7440-22-4	SILVER		0.61	1	0.063	昱	
74-10-23-5	SODIUM	1	180	100	0.25		·
7440-28-0	THALLIUM	1	1	1	0.39	U	<u> </u>
7440-62-2	VANADIUM		15	1	0.033		
440-66-6	FINO		540	2	0.29		-

Total MERCURY

Method SW7471

Sample Results

PRELIMINARY RESULTS

Lab Name: Paragon Analytics, Inc.
Client Name: Morrison Knudsen Corporation

Client Project ID: VB/I-70 III8 4984

Work Order Number: 0010156 Reporting Basis; Dry Weight Final Volume: 100 ML

Matrix: SQIL Result Units: MG/KG

Client Sample ID	LabiD	Date Gollected	Dato Prepared	Date Analyzed	Percent Moisture	Dilution Factor	fluzes	Reporting Limit	WDT	Flag	Sample Aliquot
3-16623-8	0010155-1	10/19/2000	10/23/2000	10/23/2000	0.2	1	0.63	0,1	0.0028		.5.0
J 15023-B	0010156-2	10/19/2000	10/23/2000	10/23/2000	0.2	1	0.59	0.1	0.0028		.6 C
3-15624-B	0010156-3	10/10/2000	10/23/2000	10/23/2000	0.5	1	0.68	0.1	0.0028		.03.
3-15-127-F	0010156-4	10/19/2000	10/23/2000	10/25/2000	0.5	1	1.0	0.1	0,0028		,6 Q
3-15679.F	0010156-5	10/19/2000	19/23/2000	10/25/2000	0.4	1	16	0,1	0.0028		.6 3
3-15630-F	0010156-6	10/19/2000	10/23/2000	10/23/2000	0.2	2 1	1.8 24	0.2- 24	C:CQ 0.0038	-	.6 G
3-15703-B	0010156-7	10/19/2000	10/23/2000	10/23/2000	0,7	= 1	0.22	0.1	D,0028	-	,6 €
3-15/04-6	0010156-0	10/19/2000	10/23/2000	10/23/2000	0.3	1	0.25	0.1	0.0028	~-	.5 G
3-15705-B	001015S-B	10/19/2000	10/23/2000	10/23/20/10	0.1	1	0.23	0.1	0.0028		,8 6
2-1×700-F	0010156-10	10/19/2000	10/23/2000	10/23/2000	0.3	1	0,6	0.1	0.0028		.6 G
3-15701-F	0010156-11	10/19/2000	10/23/2000	10/23/2000	0.2	1	0,56	0.1	0.0028	N	,6 G
2-15702-F	001015G-12	1079/2000	10/23/2000	10/23/2000	€.⊄		0,68	1.0	0.0028		.£ G

Comments:

1. NO or U = Not Detected at or above the client requested detection limit.

Data Package ID; HG0010156-1

Date Printed: Tuesday, October 24, 2000

Paragon Analytics Inc.

Page 1 of 1